

Review

# Essential Oils of Lamiaceae Family Plants as Antifungals

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**Abstract:** The incidence of fungal infections has been steadily increasing in recent years. Systemic mycoses are characterized by the highest mortality. At the same time, the frequency of infections caused by drug-resistant strains and new pathogens e.g., *Candida auris* increases. An alternative to medicines may be essential oils, which can have a broad antimicrobial spectrum. Rich in the essential oils are plants from the Lamiaceae family. In this review are presented antifungal activities of essential oils from 72 Lamiaceae plants. More than half of these have good activity (minimum inhibitory concentrations (MICs) < 1000 µg/mL) against fungi. The best activity (MICs < 100) have essential oils from some species of the genera *Clinopodium*, *Lavandula*, *Mentha*, *Thymbra*, and *Thymus*. In some cases were observed significant discrepancies between different studies. In the review are also shown the most important compounds of described essential oils. To the chemical components most commonly found as the main ingredients include β-caryophyllene (41 plants), linalool (27 plants), limonene (26), β-pinene (25), 1,8-cineole (22), carvacrol (21), α-pinene (21), p-cymene (20), γ-terpinene (20), and thymol (20).

**Keywords:** Labiatae; fungi; *Aspergillus*; *Cryptococcus*; *Penicillium*; dermatophytes; β-caryophyllene; sesquiterpene; monoterpenes; minimal inhibitory concentration (MIC)

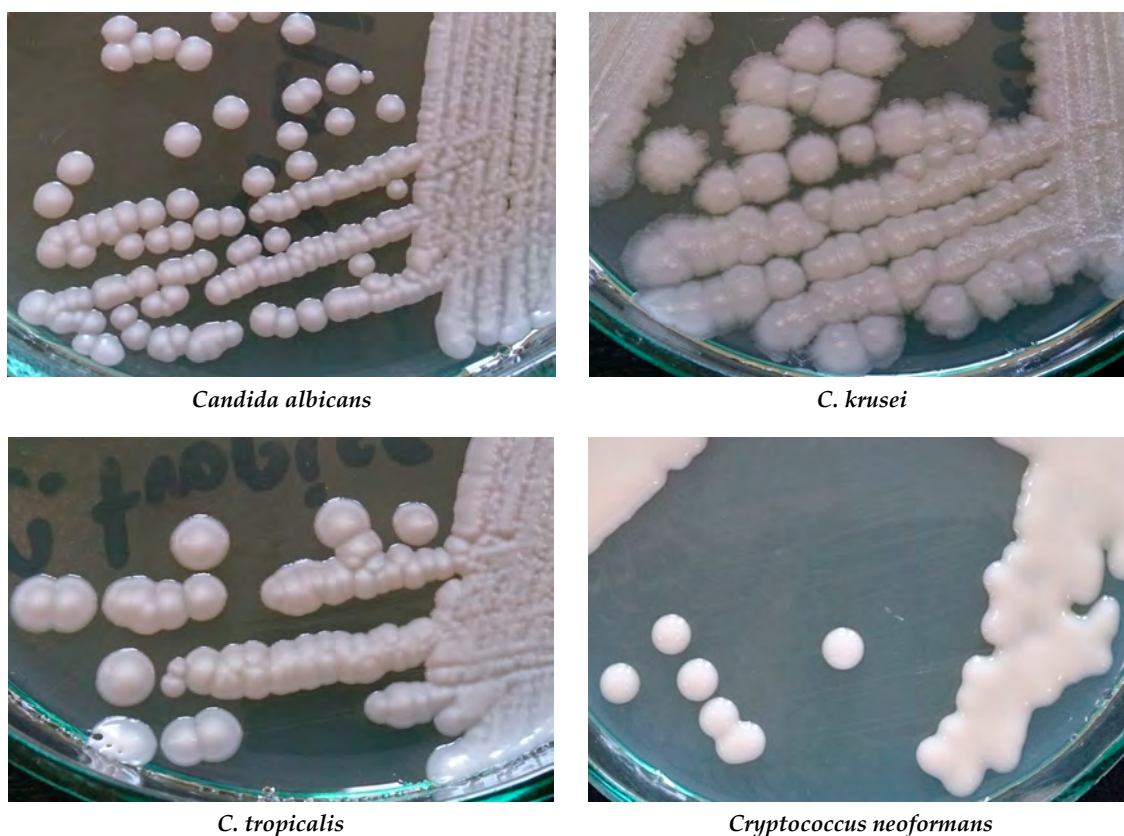
## 1. Introduction

Fungal infections belong to the most often diseases of humans. It is estimated that about 1.7 billion people (25% of the population) have skin, nail, and hair fungal infections [1]. The development of most of these infections is affected by dermatophytes, namely *Trichophyton* spp., *Microsporum* spp., and *Epidermophyton* spp. [2]. Simultaneously, mucosal infections of the oral and genital tracts caused by *Candida* spp. are very common. About 0.13 billion of women suffer from vulvovaginal candidiasis. On the other hand, oral candidiasis are common in babies and denture wearers. Fungi also cause life-threatening systemic infections, with mortality reaching >1.6 million, which is >3-fold more than malaria [3]. Among life-threatening fungal infections prevail cryptococcosis (*Cryptococcus neoformans*) with >1,000,000 cases and mortality rate 20–70%, candidiasis (*Candida albicans*) with >400,000 cases and mortality rate 46–75%, pneumocystosis (*Pneumocystis jirovecii*) with >400,000 cases and mortality rate 20–80%, and aspergillosis (*Aspergillus fumigatus*) with >200,000 cases and mortality rate 30–95% [1,4,5]. In Table 1 are presented diseases caused by some of the most often fungal pathogens among people.

**Table 1.** Fungal pathogens of humans and most often observed mycoses (based on [6,7]).

Superficial mycoses	<ul style="list-style-type: none"> <li>• <i>Hortae werneckii</i> (Tinea nigra)</li> <li>• <i>Malassezia furfur</i> (Pityriasis versicolor)</li> <li>• <i>Piedraia hortae</i> (Black piedra)</li> <li>• <i>Trichosporon</i> spp. (White piedra)</li> </ul>
Cutaneous and subcutaneous mycoses	<ul style="list-style-type: none"> <li>• <i>Aspergillus</i> spp. (Onychomycosis, Keratitis)</li> <li>• <i>Candida</i> spp. (Tinea pedis, Tinea cruris, Onychomycosis, Keratitis)</li> <li>• <i>Chaetomium</i> spp. (Subcutaneous phaeohyphomycosis)</li> <li>• <i>Curvularia</i> spp. (Subcutaneous phaeohyphomycosis)</li> <li>• <i>Epidermophyton</i> spp. (Tinea pedis, Tinea cruris, Onychomycosis)</li> <li>• <i>Exophiala</i> spp. (Chromoblastomycosis, Subcutaneous phaeohyphomycosis)</li> <li>• <i>Fonsecaea</i> spp. (Chromoblastomycosis)</li> <li>• <i>Fusarium</i> spp. (Onychomycosis, Keratitis, Eumycotic mycetoma)</li> <li>• <i>Geotrichum</i> spp. (Onychomycosis)</li> <li>• <i>Microsporum</i> spp. (Tinea corporis, Tinea capitis)</li> <li>• <i>Phaeoacremonium</i> spp. (Eumycotic mycetoma)</li> <li>• <i>Phialophora</i> spp. (Chromoblastomycosis, Subcutaneous phaeohyphomycosis)</li> <li>• <i>Scopulariopsis brevicaulis</i> (Onychomycosis)</li> <li>• <i>Sporothrix schenckii</i> (Lymphocutaneous sporotrichosis)</li> <li>• <i>Trichophyton</i> spp. (Tinea pedis, Tinea corporis, Tinea cruris, Tinea capitis, Onychomycosis)</li> <li>• <i>Trichosporon</i> spp. (Onychomycosis)</li> </ul>
Endemic mycoses	<ul style="list-style-type: none"> <li>• <i>Blastomyces dermatitidis</i> (Blastomycosis)</li> <li>• <i>Histoplasma capsulatum</i> (Histoplasmosis)</li> <li>• <i>Coccidioides immitis/posadasii</i> (Coccidioidomycosis)</li> <li>• <i>Penicillium marneffei</i> (Penicilliosis)</li> <li>• <i>Paracoccidioides brasiliensis</i> (Paracoccidioidomycosis)</li> </ul>
Opportunistic mycoses	<ul style="list-style-type: none"> <li>• <i>Acremonium</i> spp. (Hyalohyphomycosis-cutaneous, disseminated infection)</li> <li>• <i>Alternaria</i> spp. (Phaeohyphomycosis-subcutaneous, sinusitis, disseminated infection)</li> <li>• <i>Aspergillus</i> spp. (Allergic reactions, Aspergillosis-nasal, sinusitis, bronchial, pulmonary, systemic dissemination)</li> <li>• <i>Bipolaris</i> spp. (Phaeohyphomycosis-subcutaneous, sinusitis, brain abscess)</li> <li>• <i>Candida</i> spp. (Candidiasis-superficial mucosal, cutaneous, widespread hematogenous distribution involving target organs)</li> <li>• <i>Cryptococcus</i> spp. (Cryptococcosis-cutaneous, pulmonary, meningitis)</li> <li>• <i>Curvularia</i> spp. (Phaeohyphomycosis-subcutaneous, sinusitis, disseminated infection)</li> <li>• <i>Fusarium</i> spp. (Hyalohyphomycosis-cutaneous, disseminated infection)</li> <li>• <i>Lichtheimia</i> spp. (Mucormycosis-cutaneous, invasive)</li> <li>• <i>Mucor</i> spp. (Mucormycosis-cutaneous, invasive)</li> <li>• <i>Paecilomyces</i> spp. (Hyalohyphomycosis-cutaneous, disseminated infection)</li> <li>• <i>Pneumocystis jirovecii</i> (Pneumocystosis-pneumonia, extrapulmonary manifestations)</li> <li>• <i>Rhizomucor</i> spp. (Mucormycosis-cutaneous, invasive)</li> <li>• <i>Rhizopus</i> spp. (Mucormycosis-cutaneous, invasive)</li> <li>• <i>Scedosporium</i> spp. (Hyalohyphomycosis-cutaneous, disseminated infection)</li> <li>• <i>Trichosporon</i> spp. (Trichosporonosis-invasive disease)</li> <li>• <i>Wangiella</i> spp. (Phaeohyphomycosis-subcutaneous, sinusitis, brain abscess)</li> </ul>

The big problem is growing drug-resistance amid fungi. Among *Candida* and *Aspergillus* species is observed resistance to azoles, e.g., to fluconazole, voriconazole, and posaconazole. Some *Candida* species, especially *C. glabrata* and *C. parapsilosis*, can be echinocandin- and multidrug-resistant [8,9]. Acquired resistance to echinocandins has also been reported for yeasts *C. albicans*, *C. tropicalis*, *C. krusei*, *C. kefyr*, *C. lusitanae*, and *C. dubliniensis* [10]. More than 3% of *Aspergillus fumigatus* isolates are resistant to one or more azoles [11]. Polyene resistance mainly concerns amphotericin B. Resistance to this drug is observed in *Fusarium* spp., *Trichosporon* spp., *Aspergillus* spp., and *Sporothrix schenckii* [12,13]. Resistance to amphotericin B has also been reported for *C. albicans*, *C. glabrata*, and *C. tropicalis* [14–16]. Cultures of some *Candida* species and *Cryptococcus neoformans* are presented in Figure 1.



**Figure 1.** Cultures of selected yeast fungi on Sabouraud agar (Author of photos: Tomasz M. Karpiński).

The new epidemiological problem is *C. auris*, a multidrug-resistant organism first described in Japan in 2009 [17]. Recently, *C. auris* has been reported from 36 countries from six continents [18]. About 30% of isolates demonstrate reduced susceptibility to amphotericin B, and 5% can be resistant to the echinocandins [19,20]. The estimated mortality from *C. auris* fungemia range from 28% to 60% [21].

Fundamental issues are also the costs of treatment and hospitalization of patients with invasive fungal diseases. According to Drgona et al., all costs range from around €26,000 up to over €80,000 per patient [5].

Therefore, all time, new treatments for fungal infections are being sought. One option may be to apply natural products having antifungal activity. Among these, significant importances have essential oils, which can have a broad antimicrobial spectrum. Rich in the essential oils are among other plants from the Lamiaceae family.

In this review are presented antifungal activities of essential oils from seventy-two (72) plants of the Lamiaceae family. Moreover, are shown the most important compounds of these essential oils. For objective comparison of results, in this paper were included only antifungal studies specifying the minimum inhibitory concentrations (MICs) for essential oils. The MIC (expressed in  $\mu\text{g/mL}$ ) is the

lowest concentration of an antimicrobial agent in which no growth of a microorganism is observed in an agar or broth dilution susceptibility test [22–24].

## 2. Components of Essential Oils of Lamiaceae Family

The family Lamiaceae or Labiatae contains many valuable medicinal plants. In the family are 236 genera and between 6900 and 7200 species. To the most abundant genera belong *Salvia* (900 species), *Scutellaria* (360), *Stachys* (300), *Plectranthus* (300), *Hyptis* (280), *Teucrium* (250), *Vitex* (250), *Thymus* (220), and *Nepeta* (200). Lamiaceae plants rich in essential oils have great worth in natural medicine, pharmacology, cosmetology, and aromatherapy [25]. The essential oils are mostly present in leaves, however, they can be found in flowers, buds, fruits, seeds, rind, wood, or roots [26]. Essential oils are mixtures of volatile compounds, which are secondary plant metabolites. They play a role in the defense system of higher plants [27]. Essential oils may contain over 300 different compounds, mainly of molecular weight below 300 [28]. Some oils, e.g., obtained from *Lavandula*, *Geranium*, or *Rosmarinus*, contain 450 to 500 chemicals [29]. Among the active compounds of essential oils are various chemical classes, e.g., alcohols, ethers, aldehydes, ketones, esters, phenols, terpenes (monoterpenes, sesquiterpenes), and coumarins [30,31].

In Table 2 are presented the main chemical components of essential oils of selected Lamiaceae family plants. Plant names were unified according to The Plant List [32], however synonyms used in the literature were also left. Chemical component names were unified, according to PubChem [33].

**Table 2.** The main chemical components of the essential oils of selected Lamiaceae family plants.

Essential Oil	Main Chemical Components	References
<i>Aeollanthus suaveolens</i> Mart. ex Spreng. = <i>A. heliotropioides</i> Oliv.	Linalool (38.5%), $\alpha$ -Farnesene (25.1%), Massoialactone (4.5%), $\beta$ -Caryophyllene (3.6%), Germacrene D (2.0%)	[34]
<i>Agastache rugosa</i> (Fisch. and C.A.Mey.) Kuntze	Methyl chavicol (93.45%), Methyl eugenol (2.48–50.51%), Estragole (8.55%), Eugenol (0.15–7.54%), Thymol (3.62%), Pulegone (2.56%), Limonene (2.49%), $\beta$ -Caryophyllene (1.19–2.38%),	[35,36]
<i>Ballota nigra</i> subsp. <i>foetida</i> (Vis.) Hayek	$\beta$ -Caryophyllene (21.8–22.6%), Caryophyllene oxide (18.0–20.5%), Germacrene D (13.1–16.5%), 2-Hexenal (6.5–11.2%), 1-Octen-3-ol (3.5–5.5%), $\beta$ -Pinene (1.6–4.4%), Limonene (2.2–4.1%), Linalool (1.2–3.5%), $\beta$ -Bourbonene (1.5–2.7%), $\alpha$ -Humulene (2.2–2.6%), $\alpha$ -Copaene (1.5–2.2%)	[37]
<i>Clinopodium dalmaticum</i> (Benth.) Bräuchler and Heubl = <i>Micromeria dalmatica</i> Benth.	Piperitenone oxide (41.77%), Pulegone (15.94%), Piperitenone (10.19%), Limonene (5.77%), Piperitone (3.39%), $\alpha$ -Pinene (2.9%), $\beta$ -Pinene (2.16%),	[38]
<i>Clinopodium nepeta</i> subsp. <i>glandulosum</i> (Req.) Govaerts = <i>Calamintha glandulosa</i> (Req.) Bentham = <i>Calamintha officinalis</i> Moench	Piperitenone (trace–42.6%), Piperitone (0.0–40.3%), Carvone (1–38.7%), Pulegone (0.6–9.7%), Shisofuran (0.1–9.7%), Menthone (trace–8.3%), Dihydrocarveol acetate (0.1–7.6%), Dihydrocarveol (0–6.9%), 1,8-Cineole (0.0–6.4%), cis-Carvyl acetate (0.0–6.1%),	[39,40]
<i>Clinopodium nepeta</i> (L.) Kuntze = <i>Calamintha nepeta</i> (L.) Savi	Pulegone (2.4–84.7%), Isomenthone (1.9–51.3%), Menthone (0.0–35.4%), Crysanthenone (1.3–33.9%), 1,8-Cineole (0.3–21.4%), Piperitenone oxide (0.0–19.1%), Limonene (0.0–13.6%), Isopulegone (0.0–9.4%), Piperitenone (0.0–7.7%), Cinerolone (0.0–5.8%), Isopulegol (0.0–4.1%), Isomenthol (0.0–3.9%), $\beta$ -Caryophyllene (0.0–3.8%), 3-Octanol (0.0–3.0%), $\beta$ -Pinene (0.0–2.3%), cis-Piperitone oxide (0.0–2.2%)	[41,42]

Table 2. Cont.

Essential Oil	Main Chemical Components	References
<i>Clinopodium thymifolium</i> (Scop.) Kuntze = <i>Micromeria thymifolia</i> (Scop.) Fritsch	Pulegone (32.81%), Piperitenone (25.7%), Piperitone (11.71%), Isomenthone (4.98%), Limonene (2.4%), $\beta$ -Caryophyllene (2.39%)	[38]
<i>Clinopodium umbrosum</i> (M.Bieb.) Kuntze = <i>Calamintha umbrosa</i> Benth.	$\beta$ -Caryophyllene (13.9%), Germacrene D (11.6%), Spathulenol (10.6%)	[43]
<i>Dracocephalum heterophyllum</i> Benth.	Citronellol (74.2%), Geraniol (2.8%), cis-Rose oxide (2.2%), Citronellyl acetate (1.7%)	[44]
<i>Hymenocrater longiflorus</i> Benth.	$\delta$ -Cadinol (18.49%), $\alpha$ -Pinene (10.16%), p-Menth-1-en-8-ol (9.82%), Hedycaryol (6.42%), $\beta$ -Eudesmol (4.56%), Spathulenol (4.14%), $\delta$ -Cadenene (3.02%), Linalool (2.98%), Caryophyllene oxide (2.81%), $\beta$ -Bourbonene (2.72%), $\beta$ -Caryophyllene (2.29%)	[45]
<i>Hyptis ovalifolia</i> Benth.	(R)-6-[(Z)-1-Hepteny]-5,6-dihydro-2H-pyran-2-one (60.0%), $\gamma$ -Cadinene (6.6%), Viridiflorol (6.08%), Caryophyllene oxide (4.98%), $\gamma$ -Elemene (4.38%)	[46]
<i>Hyssopus officinalis</i> L.	Pinocamphone (5.78–50.77%), 1,8-Cineole (0.47–36.43%), Pinocarvone (0.44–23.4%), $\beta$ -Pinene (13.38–19.55%), Isopinocamphone (15.32%), $\alpha$ -Phellandrene (trace–3.74%), Sabinene (1.7–2.9%), Myrtenol (1.39–2.7%), $\alpha$ -Pinene (1.01–2.57%), cis-Sabinene hydrate (0.0–2.5%), Myrtenyl methyl ether (1.64–2.1%)	[44,47,48]
<i>Lavandula angustifolia</i> Mill.	Linalool (20.18–45.8%), Linalyl acetate (4.6–43.13%), Lavandulyl acetate (0–16.01%), 1,8-Cineole (0.6–13.1%), Camphor (0.52–11.2%), Borneol (0.76–7.5%), Terpinen-4-ol (1.05–5.8%), $\beta$ -Caryophyllene (0.6–4.95%), Lavandulol (0–3.09%), $\beta$ -Ocimene (1.5–2.84%), Myrcene (0.4–2.41%)	[49–51]
<i>Lavandula multifida</i> L.	Carvacrol (41.5–42.8%), $\beta$ -Ocimene (27.0–27.4%), Myrcene (5.5–5.7%), $\beta$ -Bisabolene (5.0–5.6%), Terpinolene (2.1–3.1%), $\alpha$ -Farnesene (2.6–2.8%)	[52]
<i>Lavandula pedunculata</i> (Mill.) Cav.	Fenchone (6.2–44.5%), 1,8-Cineole (5.1–34.3%), Camphor (8.7–34.0%), $\beta$ -Pinene (1.4–9.0%), $\alpha$ -Pinene (2.5–8.0%), Camphene (0.8–6.1%), Linalool (0.5–3.8%), Bornyl acetate (0.9–3.5%), Borneol (0.6–3.4%), $\alpha$ -Cadinol (0.2–3.1%), cis-Verbenol (0.2–2.8%), Myrtenal (0.8–2.4%), trans-Verbenol (1.1–2.0%)	[53]
<i>Lavandula stoechas</i> L.	Fenchone (0.0–36.2%), 1,8-Cineole (0–33.9%), Camphor (2.2–18%), $\alpha$ -trans-Necrolyl acetate (0.0–17.4%), Lavandulyl acetate (0.0–7.6%), $\alpha$ -trans-Necrodol (0.0–7.1%), Linalool (0.0–6.2%), $\alpha$ -Copaene-8-ol (0.7–4.7%), Viridiflorol (1.4–3.6%), $\alpha$ -Pinene (1.1–3.2%), 2,3,4,4-Tetramethyl-5-methylene-cyclopenten-1-one (0.0–2.8%), Lyratyl acetate (0–2.4%), Myrtenyl acetate (1.0–2.0%), 1,1,2,3-Tetramethyl-4-hydroximethyl-2-cyclopentene (0.0–2.0%)	[51,54]
<i>Lavandula viridis</i> L'Her.	1,8-Cineole (34.5–42.2%), Camphor (13.4%), $\alpha$ -Pinene (9.0%), Linalool (6.7–7.9%)	[55]
<i>Lepechinia mutica</i> (Benth.) Epling	$\Delta^3$ -Carene (8.69–24.23%), Thujopsan-2- $\alpha$ -ol (0.0–11.9%), Shyobunol (0.0–10.8%), $\beta$ -Pinene (3.78–7.96%), $\delta$ -Cadinene (0.0–6.96%), Globulol (0.0–5.91%), Valerianol (0.0–5.19%), epi-Cubebol (0.0–4.62%), $\beta$ -Caryophyllene (0.0–4.55%), Limonene (3.79–4.47%), $\alpha$ -Eudesmol (0.0–4.47%), $\alpha$ -Phellandrene (0.34–3.8%), $\beta$ -Phellandrene (3.79%), $\gamma$ -Cadinene (0.0–2.86%), $\alpha$ -Pinene (1.23–2.68%), o-Cymene (0.0–2.04%), Isobornyl acetate (0.0–2.2%)	[56,57]



Table 2. Cont.

Essential Oil	Main Chemical Components	References
<i>Marrubium vulgare</i> L.	$\gamma$ -Eudesmol (11.93%), $\beta$ -Citronellol (9.9%), Citronellyl formate (9.5%), Germacrene-D (9.37%), Geranyl formate (6.25%), Geranyl tiglate (5.53%), Ledene (5.35%), 1,8-Cineole (3.72%), Neryl acetate (3.41%), $\delta$ -Cadinene (3.3%), Cyclononasiloxane octadecamethyl (3.08%), Geraniol (2.74%), N-trimethylsilyl trifluoroacetamide (2.35%), Eicosamethylcyclodecasiloxane (2.29%), $\alpha$ -Thujone (2.29%), trans-Caryophyllene (2.15%)	[58]
<i>Melissa officinalis</i> L.	Geranial (23.4%), Neral (16.5%), Citronellal (13.7%), $\beta$ -Caryophyllene (4.6%), Geraniol (3.4%), Isomenthone (3.0%), Menthol (2.9%), Methyl citronellate (2.7%), Germacrene D (2.4%), Limonene (2.2%)	[59]
<i>Mentha cervina</i> L.	Isomenthone (8.7–77%), Pulegone (12.9–75.1%), Menthone (0.8–4.4%), Limonene (0.8–4.3%)	[60]
<i>Mentha × piperita</i> L.	Menthol (34.82–43.85%), Menthone (9.1–31.68%), Carvone (0.0–19.54%), Menthyl acetate (1.64–17.4%), Anethole (0.0–9.54%), Isomenthone (4.71–8.08%), Limonene (0.86–6.9%), Menthofuran (6.8%), Eucalyptol (4.36–6.21%), 1,8-Cineole (5.6%), Pulegone (0.47–5.15%), Isomenthol acetate (4.56–4.91%), Isomenthol (0.68–3.58%), Sabinene (0.0–2.5%)	[61–64]
<i>Mentha pulegium</i> L.	Pulegone (2.3–70.66%), Piperitone (0.24–38.0%), Piperitenone (1.58–33.0%), Neomenthol (11.21%), $\alpha$ -Terpineol (0.0–4.7%), 1,8-Cineole (0.11–4.0%), Piperitenone oxide (0.0–3.4%), Menthone (2.63–3.0%), Borneol (0.0–2.9%), Isopulegone (2.33%)	[65,66]
<i>Mentha requienii</i> Benth.	Pulegone (77.6%), Isomenthone (18.2%), Limonene (1.76%)	[67]
<i>Mentha spicata</i> L.	Pulegone (0.0–78.7%), Carvone (0.0–59.12%), Menthol (0.0–39%), Menthone (5.1–21.9%), Neomenthol (11.2%), Menthyl acetate (0.0–6.9%), Dihydrocarveol (0.0–6.27%), Limonene (1.0–5.8%), 1,8-Cineole (3.0–5.42%), cis-Dihydrocarvone (0.0–4.9%), cis-Carveol (0.0–3.9%), $\beta$ -Caryophyllene (0.7–2.8%), $\beta$ -Myrcene (0.3–2.3%)	[49,51,61,68]
<i>Mentha suaveolens</i> Ehrh.	Piperitenone oxide (0.0–87.25%), Carvone (0.0–50.59%), Pulegone (0.0–50.0%), Demelverine (0.0–43.46%), Cinerolone (0.0–38.79%), p-Cymenene (0.0–35.22%), Limonene (0.0–31.25%), Piperitone oxide (0.0–26.0%), p-Cymenol-8 (0.0–20.6%), Spathulenol (0.0–18.35%), $\beta$ -Caryophyllene oxide (0.3–17.25%), $\alpha$ -Pharnesene (0.0–16.54%), $\alpha$ -Cadinol (0.09–10.69%), Calamenene (0.44–10.63%), $\alpha$ -Cubenene (0.0–10.08%), $\alpha$ -Caryophyllene (2.0–9.8%), Veridiflorol (0.0–7.59%), Cubenol (0.0–7.46%), Verbenone (0.0–6.56%), $\delta$ -Fenchol (0.3–5.9%), Menthone (0.0–5.7%), Borneol (0.12–5.6%), Citronellyl acetate (0.0–5.45%), $\delta$ -Cadinene (0.0–4.89%), Eucalyptol (0.0–4.21%), cis-8-Menthene (0.3–4.2%), Fenchone (0.1–3.6%), Geraniol (1.0–3.4%), $\tau$ -Muurolol (0.0–3.29%), $\alpha$ -Pinene (0.1–2.7%), $\beta$ -Caryophyllene (2.56%), cis-Carveol (2.31%), Germacrene D (0.0–2.04%)	[69–71]
<i>Micromeria albanica</i> (K. Maly) Silic	Piperitenone oxide (38.73%), Pulegone (13.43%), Piperitenone (9.72%), Piperitone (5.62%), Limonene (3.2%), $\alpha$ -Copaene (2.12%)	[38]

Table 2. Cont.

Essential Oil	Main Chemical Components	References
<i>Moluccella spinosa</i> L.	$\alpha$ -Pinene (26.6%), Caryophyllene oxide (16.8%), $\beta$ -Caryophyllene (8.6%), $\alpha$ -Thujene (5.9%), Nonacosane (5.5%), Heptacosane (5.3%), Ethylbenzaldehyde (3.4%), Pentacosane (2.5%), Tetracosane (2.3%), Sabinene (2.2%)	[72]
<i>Nepeta ciliaris</i> Benth. = <i>Nepeta leucophylla</i> Benth.	Caryophyllene oxide (14.8–26.3%), $\beta$ -Caryophyllene (18.0%), $\beta$ -Sesquiphellandrene (15.0%), Iridodial b-monoenol acetate (9.8%)	[43]
<i>Nepeta clarkei</i> Hook. f.	$\beta$ -Sesquiphellandrene (22.0%), Actinidine (10.0%), Germacrene D (8.0%)	[43]
<i>Ocimum basilicum</i> L.	Linalool (18.0–68.0%), Methyl chavicol (0.0–57.3%), Geraniol (0.0–16.5%), 1,8-Cineole (1.4–15.1%), p-Allylanisole (0.2–13.8%), Eugenol (0.0–12.32%), Limonene (0.2–10.4%), $\beta$ -Farnesene (0.0–6.3%), $\tau$ -Cadinol (trace–5.8%), $\beta$ -Caryophyllene (0.0–4.5%), $\alpha$ -Bergamotene (0.0–4.34%), $\alpha$ -Cadinol (0.0–4.05%), $\beta$ -Elemene (0.0–3.62%), $\delta$ -Cadinene (0.0–3.6%), Germacrene D (0.0–3.5%), $\gamma$ -Cadinene (0.0–2.8%), Camphor (0.0–2.4%), $\beta$ -Myrcene (0.2–2.3%), Terpinen-4-ol (0.0–2.2%), Guaiene (0.0–2.1%), Estragole (0.0–2.03%), Isolimonene (0.0–2.0%), $\alpha$ -Bulnesene (0.0–2.0%), $\gamma$ -Terpinene (0.0–2.0%)	[64,68,73–76]
<i>Ocimum</i> $\times$ <i>africanum</i> Lour. = <i>Ocimum</i> $\times$ <i>citriodorum</i>	Nerol (23.0%), Geraniol (15.77%), Methyl chavicol (9.45%), Linalool (9.42%), $\beta$ -Bisabolene (8.31%), $\beta$ -Caryophyllene (7.8%), Geraniol (5.2%), Neral (4.93%), $\alpha$ -Bergamotene (3.52%), $\alpha$ -Bisabolene (2.29%), $\beta$ -Cubebene (2.26%)	[76]
<i>Ocimum campechianum</i> Mill. = <i>Ocimum micranthum</i> Willd.	Eugenol (46.55%), $\beta$ -Caryophyllene (11.94%), $\beta$ -Elemene (9.06%), 1,8-Cineole (5.35%), $\delta$ -Elemene (4.17%), Bicyclogermacrene (2.9%), cis-Ocimene (2.69%), allo-Ocimene (2.42%), $\alpha$ -Humulene (2.4%)	[73]
<i>Ocimum forskolei</i> Benth.	endo-Fenchol (31.1%), $\tau$ -Cadinol (12.2%), Fenchone (12.2%), Camphor (6.2%), Linalool (5.7%), Methyl(E)-cinnamate (5.1%), $\alpha$ -Bergamotene (3.1%), $\gamma$ -Cadinene (2.9%), endo-Fenchyl acetate (2.8%), Limonene (2.5%)	[77]
<i>Ocimum gratissimum</i> L.	Eugenol (7.42–57.82%), Ethyl cinnamate (0.0–34.0%), Linalool (30.0–32.95%), 1,8-Cineole (6.5–21.91%), $\alpha$ -Bisabolene (0.0–17.19%), Camphor (3.8–11.97%), Thymol (0.0–9.8%), $\alpha$ -Cadinol (5.18%), Germacrene D (0.79–4.76%), $\alpha$ -Terpineol (3.36%), $\gamma$ -Terpinene (0.0–3.06%), $\beta$ -Caryophyllene (1.68–3.03%), p-Cymene (0.0–2.11%)	[78–80]
<i>Ocimum tenuiflorum</i> L. = <i>Ocimum sanctum</i> L.	Eugenol (0.0–61.3%), Methyl chavicol (0.0–44.63%), Linalool (0.26–21.84%), $\alpha$ -Caryophyllene (3.3–11.89%), Germacrene D (0.37–9.14%), Carvone (0.0–6.31%), Limonene (0.71–4.39%), $\beta$ -Caryophyllene (1.4–3.3%), $\alpha$ -Cubebene (0.0–2.54%), Carvacrol (0.0–2.04%)	[81–83]
<i>Origanum compactum</i> Benth.	Carvacrol (43.26%), Thymol (21.64%), p-Cymene (13.95%), $\gamma$ -Terpinene (11.28%)	[84]
<i>Origanum majorana</i> L.	Terpinen-4-ol (6.66–33.84%), Sabinene hydrate (2.31–28.33%), 1,8-Cineole (0.0–20.9%), Carvacrol (0.0–20.8%), $\gamma$ -Terpinene (7.59–19.5%), Thymol (0.0–12.18%), $\alpha$ -Terpinene (3.03–10.08%), $\beta$ -Phellandrene (1.96–8.0%), p-Cymene (2.45–7.84%), Sabinene (3.2–6.7%), Limonene (0.0–5.3%), $\alpha$ -Terpineol (2.7–4.7%), Linalool (0.0–4.4%), Terpinolene (0.98–3.76%), Linalool acetate (1.82–3.2%), Geraniol (2.7%), $\beta$ -Caryophyllene (1.7–2.38%), $\alpha$ -Pinene (0.0–2.0%)	[62,68,85–87]

Table 2. Cont.

Essential Oil	Main Chemical Components	References
<i>Origanum vulgare</i> L.	Pulegone (0.0–77.45%), Carvacrol (0.21–65.9%), Cymenol (0.0–58.6%), Thymol (3.7–45.22%), o-Cymene (0.0–14.33%), Terpinen-4-ol (0.03–12.55%), $\beta$ -Terpineol (0.0–10.46%), p-Cymene (0.5–9.3%), $\gamma$ -Terpinene (3.1–9.12%), Borneol (0.0–6.1%), $\alpha$ -Pinene (0.0–5.1%), Menthone (0.0–4.86%), Linalool (0.0–4.8%), $\beta$ -Bisabolene (0.0–4.5%), Caryophyllene oxide (0.0–4.5%), Sabinene (0.0–3.91%), $\beta$ -Phellandrene (0.0–3.74%), $\beta$ -Caryophyllene (0.0–3.7%), $\alpha$ -Terpineol (0.0–3.35%), Sabinene hydrate (0.0–3.31%), $\alpha$ -Cadinol (0.0–3.3%), $\alpha$ -Terpinene (1.63–3.1%), Eucalyptol (0.0–2.8%), $\beta$ -Ocimene (0.0–2.77%), cis-Isopulegone (2.22%), $\beta$ -Myrcene (0.0–2.2%), Anisole (0.0–2.13%), Piperitenone (0.0–2.13%), Germacrene D (0.0–1.23%)	[49,62,64,68,74,88–91]
<i>Pogostemon cablin</i> (Blanco) Benth.	Patchouli alcohol (38.3–44.52%), $\alpha$ -Bulnesene (0.0–13.3%), $\delta$ -Guaiene (12.64%), $\alpha$ -Guaiene (8.89–9.6%), Pogostol (0.0–6.2%), Seychellene (5.8%), $\alpha$ -Bergamotene (5.76%), Eremophilene (4.34%), $\beta$ -Guaiene (3.54%), $\beta$ -Caryophyllene (1.93–3.0%), $\beta$ -Patchoulene (1.8–2.77%)	[92,93]
<i>Pogostemon heyneanus</i> Benth.	Acetophenone (51.0%), Patchouli alcohol (14.0%), Nerolidol (5.4%), $\beta$ -Pinene (5.3%), Limonene (4.0%), Benzoyl acetone (3.1%), $\alpha$ -Pinene (2.4%), $\beta$ -Caryophyllene (2.0%)	[93]
<i>Premna microphylla</i> Turcz.	Blumenol C (49.7%), $\beta$ -Cedrene (6.1%), Limonene (3.8%), $\alpha$ -Guaiene (3.3%), Cryptone (3.1%), $\alpha$ -Cyperone (2.7%), cis-14-nor-Muurool-5-en-4-one (2.4%)	[94]
<i>Rosmarinus officinalis</i> L.	$\alpha$ -Pinene (5.4–37.9%), 1,8-Cineole (0.88–26.54%), Eucalyptol (0.0–24.34%), Limonene (0.0–21.7%), Camphor (2.45–21.6%), Myrcene (0.9–20.18%), Borneol (0.0–18.08%), Bornyl acetate (0.92–14.9%), Verbenone (1.36–12.0%), Camphene (1.7–11.38%), Linalool oxide (0.0–10.8%), $\beta$ -Pinene (0.0–6.95%), $\beta$ -Caryophyllene (0.0–6.3%), Linalool (0.0–5.32%), o-Cymene (0.0–4.43%), p-Cymene (0.0–4.34%), $\beta$ -Phellandrene (0.0–3.9%), Sabinene (0.0–3.72%), $\alpha$ -Terpineol (1.19–3.36%), Isobornyl acetate (0.0–3.3%), Carvacrol (0.0–3.15%), Verbenol (0.7–3.03%), $\alpha$ -Humulene (0.0–2.6%), $\alpha$ -Terpinene (0.21–2.4%), Terpinen-4-ol (0.34–2.15%)	[51,62,68,87,91,95–98]
<i>Salvia fruticosa</i> Miller	1,8-Cineole (16.9–54.4%), Camphor (0.6–18.34%), Manool (0–11.2%), $\beta$ -Thujone (0.6–9.0%), $\beta$ -Pinene (0.0–9.0%), Sabinene (0.0–8.6%), Viridiflorol (0.0–8.4%), $\beta$ -Caryophyllene (1.53–8.3%), $\alpha$ -Thujone (trace–8.1%), Borneol (0.0–8.0%), Camphene (0.0–7.0%), $\alpha$ -Pinene (1.5–6.85%), Bornyl acetate (0.0–6.8%), $\alpha$ -Terpineol (trace–6.7%), Myrcene (1.3–5.2%), Caryophyllene oxide (0.0–3.9%), $\alpha$ -Terpinyl acetate (0.0–2.2%), $\alpha$ -Humulene (0.16–1.5%)	[49,51,99]
<i>Salvia mirzayanii</i> Rech. f. and Esfand	1,8-Cineole (41.2%), Linalool acetate (10.7%), $\alpha$ -Terpinyl acetate (5.7%), Myrcene (4.7%), Geranyl acetate (3.7%), $\gamma$ -Cadinene (3.3%), Linalool (2.5%), Neryl acetate (2.3%)	[100]



Table 2. Cont.

Essential Oil	Main Chemical Components	References
<i>Salvia officinalis</i> L.	1,8-Cineole (4.2–50.3%), Camphor (8.8–25.0%), $\alpha$ -Thujone (1.2–19.9%), Viridiflorol (0.5–17.5%), $\beta$ -Thujone (0.1–9.9%), $\beta$ -Pinene (0.8–7.3%), $\beta$ -Caryophyllene (1.4–5.5%), Borneol (1.5–5.4%), $\alpha$ -Pinene (0.5–4.8%), Camphene (0.2–3.9%), Bornyl acetate (0.2–3.3%), $\alpha$ -Terpineol (0.0–3.1%), $\alpha$ -Terpenyl acetate (1.4–2.9%), $\alpha$ -Humulene (0.4–2.6%), $\alpha$ -Farnesene (0.0–2.5%), Eicosane (0.0–2.0%)	[96,101]
<i>Salvia sclarea</i> L.	Linalyl acetate (84%), Caryophyllene oxide (24.1%), Linalool (13.6%), 1H-Naphtho(2,1,6)pyran (8.6%), Sclareol (11.5%), Spathulenol (11.4%), $\beta$ -Caryophyllene (5.1%)	[85,102]
<i>Satureja hortensis</i> L.	Thymol (23.12–29.0%), Carvacrol (24.5–26.5%), $\gamma$ -Terpinene (20.72–22.6%), p-Cymene (6.3–9.3%), $\alpha$ -Terpinene (2.2–2.93%), $\alpha$ -Pinene (2.6–2.91%), $\beta$ -Pinene (0.92–2.7%), Limonene (0.0–2.55%), $\beta$ -Bisabolene (0.2–2.2%)	[103,104]
<i>Satureja montana</i> L.	Carvacrol (47.1%), p-Cymene (9.0%), $\gamma$ -Terpinene (6.1%), $\beta$ -Caryophyllene (3.6%), Linalool (3.1%), Thymol (2.6%), Borneol (2.1%)	[68]
<i>Satureja thymbra</i> L.	Thymol (25.16–44.5%), $\gamma$ -Terpinene (11.1–39.23%), p-Cymene (7.17–21.7%), Carvacrol (4.18–5.3%), Carvacrol methyl ether (0.1–3.33%), $\alpha$ -Terpinene (1.0–3.26%), $\beta$ -Caryophyllene (1.2–2.76%), Caryophyllene oxide (0.32–2.0%)	[51,105]
<i>Stachys cretica</i> L.	Germacrene D (12.9–20.3%), $\beta$ -Caryophyllene (0.9–9.5%), $\alpha$ -Pinene (0.7–8.6%), Octacosane (0.0–7.2%), $\beta$ -Pinene (1.5–6.2%), Linalyl acetate (0.0–5.2%), Nonacosane (0.4–4.9%), 9-Geranyl-p-cymene (0.0–4.9%), Heptacosane (0.3–4.8%), cis-Chrysanthenyl acetate (0.0–4.8%), $\beta$ -Farnesene (3.1–4.0%), Hexadecanoic acid (1.3–3.5%), Caryophyllene oxide (0.5–2.9%), $\beta$ -Bisabolene (1.6–2.8%), Linalool (0.0–2.6%), Pentacosane (0.0–2.5%), Sesquisabinene (2.1%), Geranyl acetate (0.0–2.1%)	[106]
<i>Stachys officinalis</i> (L.) Trevis	Germacrene D (19.9%), $\beta$ -Caryophyllene (14.1%), $\alpha$ -Humulene (7.5%), $\delta$ -Cadinene (4.0%), $\beta$ -Bourbonene (3.8%), $\alpha$ -Selinene (3.4%), $\gamma$ -Muuroleone (3.2%), Oct-1-en-3-ol (2.9%), Caryophyllene oxide (2.5%), Hexadecanoic acid (2.4%), $\beta$ -Selinene (2.1%), $\gamma$ -Cadinene (2.0%), $\tau$ -Muurolol (2.0%)	[107]
<i>Stachys pubescens</i> Ten.	Germacrene (22.4%), $\delta$ -Cadinene (19.7%), 2,6-Octadien (11.5%), Linalool (9.7%), Limonene (6.3%), $\delta$ -Elemene (5.4%), $\beta$ -Ocimene (2.8%), $\alpha$ -Terpinene (2.7%), 2,6-Octadienal (2.1%)	[108]
<i>Teucrium sauwagei</i> Le Houerou	$\beta$ -Eudesmol (28.8%), $\tau$ -Cadinol (17.5%), $\alpha$ -Thujene (8.7%), $\gamma$ -Cadinene (5.6%), Sabinene (4.8%), $\beta$ -Selinene (4.2%), Limonene (2.8%), $\gamma$ -Selinene (2.8%), $\alpha$ -Selinene (2.8%), $\delta$ -Cadinene (2.2%), Terpinen-4-ol (2.2%), p-Cymene (2.0%),	[109]
<i>Teucrium yemense</i> Deflers.	Caryophyllene oxide (4.3–20.1%), 7-epi- $\alpha$ -Selinene (1.3–20.1%), $\beta$ -Caryophyllene (11.2–19.1%), $\alpha$ -Cadinol (2.0–9.5%), $\alpha$ -Pinene (2.3–6.6%), $\delta$ -Cadinene (0.4–6.5%), $\alpha$ -Humulene (4.0–6.4%), $\tau$ -Cadinol (2.0–5.7%), $\gamma$ -Selinene (0.4–5.5%), $\tau$ -Muurolol (0.6–4.9%), Shyobunol (0.0–4.6%), Valencene (0.0–3.7%), Ledol (0.5–3.6%), cis-Sesquisabinene hydrate (0.9–3.4%), $\beta$ -Pinene (1.1–3.1%), Germacrene D-4-ol (0.0–3.1%), $\gamma$ -Cadinene (0.0–2.7%), $\beta$ -Selinene (0.3–2.5%), Alloaromadendrene (trace–2.2%)	[77]

Table 2. Cont.

Essential Oil	Main Chemical Components	References
<i>Thymbra capitata</i> (L.) Cav. = <i>Thymus capitatus</i> (L.) Hoffmanns. and Link = <i>Coridothymus capitatus</i> (L.) Rchb.f. Solms	Carvacrol (35.6–75.0%), Thymol (0.1–29.3%), p-Cymene (5.0–21.0%), $\gamma$ -Terpinene (4.0–12.3%), $\alpha$ -Terpinene (1.0–3.0%), $\beta$ -Myrcene (0.8–3.0%), Linalool (0.5–2.9%), $\beta$ -Caryophyllene (0.2–2.5%)	[51,110–112]
<i>Thymbra spicata</i> L.	Carvacrol (20.1–64.0%), $\gamma$ -Terpinene (11.6–31.2%), p-Cymene (9.6–26.0%), $\alpha$ -Terpinene (1.2–10.1%), $\beta$ -Myrcene (0.9–7.7%), Thujene (trace–5.2%), $\beta$ -Caryophyllene (0.5–5.1%)	[51,113,114]
<i>Thymus bovei</i> Benth.	Geraniol (35.38%), $\alpha$ -Citral (20.37%), $\beta$ -Citral (14.76%), Nerol (7.38%), 3-Octanol (4.38%)	[115]
<i>Thymus daenensis</i> Celak.	Carvacrol (31.46%), $\alpha$ -Terpineol (22.95%), Thymol (20.2%), Camphene (6.27%), 2,6-Octadien (2.22%), Borneol (2.17%), Cyclohexanone (2.1%)	[108]
<i>Thymus kotschyanus</i> Boiss. and Hohen.	Thymol (46.72%), Benzene (6.88%), Carvacrol (3.73%), $\gamma$ -Terpinene (3.58%), $\beta$ -Caryophyllene (3.39%), Linalool (2.88%), Phenol (2.61%), Borneol (2.51%), Isopropyl (2.07%)	[108]
<i>Thymus mastichina</i> (L.) L.	1,8-Cineole (67.4%), Linalool (4.3%), $\beta$ -Pinene (4.0%), $\alpha$ -Terpineol (3.5%), $\alpha$ -Pinene (3.0%), Sabinene (2.4%)	[116]
<i>Thymus migricus</i> Klokov et Des.-Shost.	Thymol (44.9%), Geraniol (10.8%), $\gamma$ -Terpinene (10.3%), Citronellol (8.5%), p-Cymene (7.2%)	[117,118]
<i>Thymus pulegioides</i> L.	Thymol (26.0%), Carvacrol (21.0%), $\gamma$ -Terpinene (8.8%), p-Cymene (7.8%), Octan-3-one (3.9%), Camphor (3.9%), $\beta$ -Bisabolene (3.0%), Borneol (2.9%), Oct-1-en-3-ol (2.0%)	[119]
<i>Thymus schimperi</i> Ronniger	Carvacrol (13.91–39.07%), Thymol (11.53–34.66%), o-Cymene (18.72–27.06%), $\gamma$ -Terpinene (4.13–13.73%), Linalool (3.34–3.59%), 3-Octanone (1.05–2.67%), $\alpha$ -Terpinene (1.67–2.37%)	[120]
<i>Thymus serpyllum</i> L.	Thymol (52.6%), p-Cymene (15.3%), $\beta$ -Caryophyllene (6.8%), Sabinene hydrate (3.8%), $\gamma$ -Terpinene (2.9%), Terpinen-4-ol (2.4%)	[68]
<i>Thymus striatus</i> Vahl.	Thymol (59.5%), $\gamma$ -Terpinene (11.6%), p-Cymene (6.4%), Carvacrol methyl ether (5.9%), Carvacrol (4.9%), $\alpha$ -Terpinene (3.3%), $\beta$ -Caryophyllene (2.3%)	[121]
<i>Thymus vulgaris</i> L.	Carvacrol (3.5–70.3%), Thymol (0.6–51.8%), Borneol (0.0–40.6%), p-Cymene (2.9–38.9%), o-Cymene (0.0–31.7%), $\alpha$ -Terpineol (0.0–19.9%), Linalool (0.0–16.0%), $\gamma$ -Terpinene (0.3–12.65%), Camphene (0.0–12.3%), 1,8-Cineole (0.0–11.3%), $\alpha$ -Pinene (0.2–6.1%), $\beta$ -Caryophyllene (0.0–3.5%), Neomenthol (0.0–2.8%), $\beta$ -Cubebene (0.0–2.4%), Geraniol (0.0–2.32%), Menthone (0.0–2.2%)	[61,64,74,85,87,104,116,122–126]
<i>Thymus zygis</i> L.	Linalool (5.5–39.7%), Thymol (0.52–39.6%), p-Cymene (2.2–21.2%), Terpinen-4-ol (1.0–11.7%), $\beta$ -Myrcene (3.0–8.6%), $\gamma$ -Terpinene (7.6–7.9%), $\alpha$ -Terpinene (1.2–4.2%), $\beta$ -Caryophyllene (1.6–3.6%), $\alpha$ -Pinene (0.9–3.6%), Limonene (1.7–2.6%), Carvacrol (0.08–2.4%), Terpinolene (0.2–2.0%)	[116,127]
<i>Vitex agnus-castus</i> L.	Eucalyptol (20.5%), 1,8-Cineole (1.5–19.61%), Bicyclogermacrene (0.0–16.2%), $\beta$ -Farnesene (0.0–16.1%), Sabinene (0.0–14.57%), Sclarene (0.0–10.9%), $\alpha$ -Pinene (0.9–9.76%), Manool (0.0–8.2%), $\beta$ -Caryophyllene (3.0–6.6%), $\beta$ -Caryophyllene oxide (0.0–5.83%), Limonene (0.0–4.89%), Vulgarol B (0.0–4.7%), $\beta$ -Pinene (0.4–4.4%), $\alpha$ -Terpinyl acetate (1.2–4.21%), $\beta$ -Sitosterol (3.13%), p-Cymene (0.0–3.11%), Geranyl linalool (0.0–3.1%), $\beta$ -Phellandrene (0.0–3.0%), Cembrene A (0.7–2.8%), Beyrene (0.0–2.6%), $\beta$ -Myrcene (trace–2.12%), $\gamma$ -Elemene (2.11%), s-Cadinol (2.01%)	[51,128,129]

Table 2. Cont.

Essential Oil	Main Chemical Components	References
<i>Zataria multiflora</i> Boiss.	Thymol (25.8–48.4%), Carvacrol (1.5–34.36%), Carvacrol methyl ether (0.0–28.32%), p-Cymene (2.27–13.2%), $\gamma$ -Terpinene (0.92–10.6%), Linalool (0.9–6.52%), $\alpha$ -Terpinenyl acetate (5.4%), $\alpha$ -Terpineol (0.5–3.69%), $\alpha$ -Pinene (0.02–3.13%), $\beta$ -Caryophyllene (2.24–3.12%), Carvacrol acetate (0.0–2.26%), Terpinen-4-ol (0.0–2.21%)	[117,130]
<i>Ziziphora clinopodioides</i> L.	Carvacrol (0.63–74.29%), Thymol (7.28–55.6%), $\gamma$ -Terpinene (1.54–24.56%), p-Cymene (2.21–10.25%), $\alpha$ -Terpinene (0.39–2.77%)	[131,132]
<i>Ziziphora tenuior</i> L.	Pulegone (46.8%), p-Menth-3-en-8-ol (12.5%), Isomenthone (6.6%), 8-Hydroxymenthone (6.2%), Isomenthol (4.7%), Limonene (3.2%)	[133]

To the chemical components most commonly found as the main ingredients in essential oils, among plants presented in Table 2, include  $\beta$ -caryophyllene (41 plants), linalool (27 plants), limonene (26),  $\beta$ -pinene (25), 1,8-cineole (22), carvacrol (21),  $\alpha$ -pinene (21), p-cymene (20),  $\gamma$ -terpinene (20), and thymol (20) (Figure 2). Sesquiterpene  $\beta$ -caryophyllene seems particularly important antifungal component in the Lamiaceae family. Its activity and its derivatives, such as caryophyllene oxide is well known [134–136]. According to Bona et al. [137], essential oils containing high concentrations of phenolic monoterpenes (e.g., carvacrol, p-cymene, thymol) have great antifungal activities. Rich in these substances are, among others *Origanum* and *Thymus* plants. Important antifungal chemicals often presented in Lamiaceae are also other monoterpenes as alcohol linalool and cyclic 1,8-cineole, limonene, pinenes, and terpinenes [138–146]. Table 1 shows that all of these antifungal substances are common in presented plants.

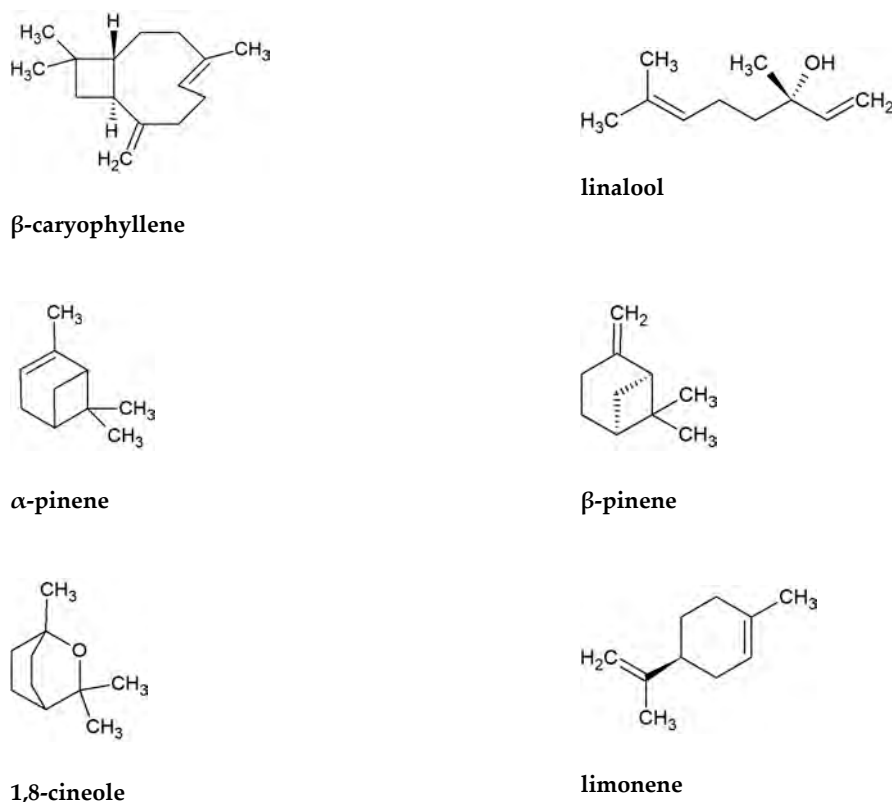
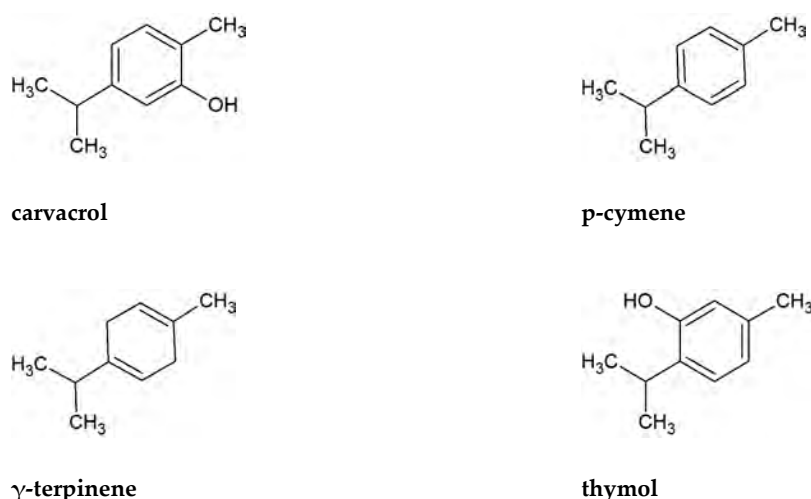


Figure 2. Cont.



**Figure 2.** Chemical formulas of ten substances the most commonly found in essential oils of Lamiaceae plants presented in Table 1.

### 3. Antifungal Activity of Essential Oils of Lamiaceae Family

In Table 3 are shown the antifungal activities of selected Lamiaceae essential oils. More than half of the essential oils have good activity ( $<1000 \mu\text{g/mL}$ ) against fungi. In some cases are observed significant discrepancies between different studies. An example could be the action of essential oils from Italian *Calamintha nepeta* against *Candida albicans*. In the work of Marongiu et al. [39], minimal inhibitory concentrations amounted to  $1.25\text{--}2.5 \mu\text{g/mL}$ , while in Božović et al. [40] MICs were between  $780$  to  $12,480 \mu\text{g/mL}$ . Differences may be related to the different biochemical composition of the examined essential oils. In results presented by Marongiu et al. [39] the main components of essential oils were pulegone ( $39.9\text{--}64.4\%$ ), piperitenone oxide ( $2.5\text{--}19.1\%$ ) and piperitenone ( $6.4\text{--}7.7\%$ ), while in Božović et al. [40] three main substances were pulegone ( $37.7\text{--}84.7\%$ ), caryophyllene ( $1.3\text{--}33.9\%$ ) and menthone ( $0.5\text{--}35.4\%$ ). Some authors have described that the content of active substances varies depending on the season. In studies of Gonçalves et al. [60] in *Mentha cervina* during the flowering phase in August amount of isomenthone and pulegone in essential oil amounted  $8.7\%$  and  $75.1\%$  respectively. Simultaneously, in the vegetative phase in February, the content of both components changed significantly and amounted to  $77.0\%$  for isomenthone and  $12.9\%$  for pulegone. Similarly, Al-Maskri et al. [75] presented essential changes in some compounds of *Ocimum basilicum* essential oil between winter and summer. In the summer essential oil, there is significantly more of linalool, p-allylanisole and  $\beta$ -farnesene, and at the same time much less content of limonene and 1,8-cineole. In this work, a seasonal variation of chemical composition is directly related to other antifungal activities. It is particularly evident in action against *Aspergillus niger*, which was lower in the summer season. Zone of growth inhibition (ZOI) for winter essential oil was  $21 \text{ mm}$  and  $\text{MIC} > 50 \mu\text{g/mL}$ , while for summer essential oil-ZOI was  $13 \text{ mm}$  and  $\text{MIC} > 100 \mu\text{g/mL}$  [75]. Influence on the content of chemical substances in essential oils also has a method of obtaining them. Čavar et al. [40] compared the composition of oils obtained from *Calamintha glandulosa* using three methods: Hydrodistillation (HD), steam distillation (SD) and aqueous reflux extraction (ARE). For example, the level of menthone was  $3.3\%$  in ARE,  $4.7\%$  in HD, and  $8.3\%$  in SD method, while for shisofuran was only  $0.1\%$  in HD and SD, and even  $9.7\%$  in ARE [40]. Additionally, many other factors can affect antimicrobial activity, such as amount and concentration of inoculum, type of culture medium, pH of the medium and incubation time. All these factors can affect the value of MIC [145]. Differences are visible in Table 2. Generally, it can be assumed that the best activity ( $\text{MICs} < 100$ ) have essential oils from *Clinopodium* spp. (excluding *C. nepeta* subsp. *glandulosum* and *C. umbrosum*), *Lavandula* spp., *Mentha* spp. (excluding *M. piperita*), *Thymbra* spp., and *Thymus* spp. (excluding *T. migricus* and *T. vulgaris*). The highest values of MICs are

presented among others for *Aeollanthus suaveolens*, *Agastache rugosa*, *Lepechinia mutica*, *Mentha × piperita*, and *Salvia sclarea*. Simultaneously, some essential oils have a very different activity, and MIC values differ depending on the region, chemical composition, research methodology, etc. Significant variations can be observed even in *Ocimum basilicum* (MICs 1–10,000), *O. sanctum* (MICs 0.1–500), *Origanum majorana* (MICs 0.5–14,400) or in *Thymus vulgaris* (MICs 0.08–3600).

The mode of action of essential oils is multidirectional. Essential oils lead to disruption of the cell wall and cell membrane through a permeabilization process. The lipophilic compounds of essential oils can pass through the cell wall and damage polysaccharides, fatty acids, and phospholipids, eventually making them permeable [146,147]. Change of the permeability for H<sup>+</sup> and K<sup>+</sup> cations affects cellular pH and damage of cellular organelles [148,149]. Additionally, essential oils inhibit the synthesis of fungal DNA, RNA, proteins, and polysaccharides [150]. Essential oils can also disintegrate mitochondrial membrane [151,152]. It has also been shown that essential oil from *Thymus vulgaris* inhibits the production of aflatoxins by *Aspergillus flavus* and leads to the reduction of ergosterol production [123].

**Table 3.** Minimal inhibitory concentrations (MICs) of essential oils against fungi.

Source of the Essential Oil	Targeted Fungus	MICs (µg/mL; µl/mL)	Reference(s)
<i>Aeollanthus suaveolens</i> Mart. ex Spreng. = <i>A. heliotropioides</i> Oliv.	<i>Candida albicans</i>	1200–5000	[34]
	<i>Candida glabrata</i>	5000	[34]
	<i>Candida krusei</i>	2500	[34]
	<i>Candida parapsilosis</i>	2500	[34]
	<i>Candida tropicalis</i>	1200	[34]
	<i>Cryptococcus neoformans</i>	600–5000	[34]
<i>Agastache rugosa</i> (Fisch. and C.A.Mey.) Kuntze	<i>Aspergillus flavus</i>	10,000	[153]
	<i>Aspergillus niger</i>	5000	[153]
	<i>Blastoschizomyces capitatus</i>	5000	[153]
	<i>Candida albicans</i>	28–5000	[153,154]
	<i>Candida utilis</i>	5000	[153]
	<i>Candida tropicalis</i>	5000	[153]
	<i>Cryptococcus neoformans</i>	10,000	[153]
	<i>Trichoderma viride</i>	5000	[153]
	<i>Trichophyton erinacei</i>	780	[153]
	<i>Trichophyton mentagrophytes</i>	3120	[153]
	<i>Trichophyton rubrum</i>	1560	[153]
	<i>Trichophyton schoenleinii</i>	1560	[153]
<i>Trichophyton soudanense</i>	1560	[153]	
<i>Trichophyton tonsurans</i>	10,000	[153]	
<i>Trichosporon mucoides</i>	5000	[153]	
<i>Ballota nigra</i> subsp. <i>foetida</i> (Vis.) Hayek	<i>Alternaria solani</i>	750	[37]
	<i>Botrytis cinerea</i>	600	[37]
	<i>Fusarium coeruleum</i>	350	[37]
	<i>Fusarium culmorum</i>	300	[37]
	<i>Fusarium oxysporum</i>	300	[37]
	<i>Fusarium solani</i>	350	[37]
	<i>Fusarium sporotrichioides</i>	350	[37]
	<i>Fusarium tabacinum</i>	350	[37]
<i>Fusarium verticillioides</i>	300	[37]	
<i>Clinopodium dalmaticum</i> (Benth.) Bräuchler and Heubl = <i>Micromeria dalmatica</i> Benth.	<i>Aspergillus niger</i>	0.4	[38]
	<i>Aspergillus ochraceus</i>	0.4	[38]
	<i>Cladosporium cladosporioides</i>	0.4	[38]
	<i>Fusarium tricinctum</i>	0.4	[38]
	<i>Penicillium ochrochloron</i>	0.4	[38]
	<i>Phomopsis helianthi</i>	0.2	[38]
<i>Trichoderma viride</i>	0.4	[38]	
<i>Clinopodium nepeta</i> subsp. <i>glandulosum</i> (Req.) Govaerts = <i>Calamintha glandulosa</i> (Req.) Bentham = <i>Calamintha officinalis</i> Moench	<i>Aspergillus niger</i>	1250	[39]
	<i>Candida albicans</i>	2500	[39]



Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
<i>Clinopodium nepeta</i> (L.) Kuntze = <i>Calamintha nepeta</i> (L.) Savi	<i>Aspergillus flavus</i>	1.25–10	[41]
	<i>Aspergillus fumigatus</i>	0.64–5	[41]
	<i>Aspergillus niger</i>	0.32–10	[41]
	<i>Candida albicans</i>	1.25–12,480	[41,42]
	<i>Candida guilliermondii</i>	1.25–2.5	[41]
	<i>Candida krusei</i>	1.25–2.5	[41]
	<i>Candida parapsilosis</i>	1.25–2.5	[41]
	<i>Candida tropicalis</i>	1.25–2.5	[41]
	<i>Cryptococcus neoformans</i>	0.32–1.25	[41]
	<i>Epidermophyton floccosum</i>	0.64–2.5	[41]
	<i>Microsporium canis</i>	0.64–2.5	[41]
	<i>Microsporium gypseum</i>	1.25–5	[41]
	<i>Trichophyton mentagrophytes</i>	0.64–5	[41]
<i>Trichophyton rubrum</i>	0.64–5	[41]	
<i>Clinopodium thymifolium</i> (Scop.) Kuntze = <i>Micromeria thymifolia</i> (Scop.) Fritsch	<i>Aspergillus niger</i>	2	[38]
	<i>Aspergillus ochraceus</i>	2	[38]
	<i>Cladosporium cladosporioides</i>	2	[38]
	<i>Fusarium tricinctum</i>	2	[38]
	<i>Penicillium ochrochloron</i>	2	[38]
	<i>Phomopsis helianthi</i>	0.4	[38]
	<i>Trichoderma viride</i>	2	[38]
<i>Clinopodium umbrosum</i> (M.Bieb.) Kuntze = <i>Calamintha umbrosa</i> Benth.	<i>Alternaria solani</i>	3000	[43]
	<i>Fusarium oxysporum</i>	2000	[43]
	<i>Helminthosporium maydis</i>	1500	[43]
<i>Dracocephalum heterophyllum</i> Benth.	<i>Alternaria solani</i>	625	[155]
	<i>Candida albicans</i>	625–1000	[44,155]
	<i>Epidermophyton floccosum</i>	2500	[155]
	<i>Fusarium semitectum</i>	313	[155]
<i>Hymenocrater longiflorus</i> Benth.	<i>Aspergillus niger</i>	480	[45]
	<i>Candida albicans</i>	240	[45]
<i>Hyptis ovalifolia</i> Benth.	<i>Microsporium canis</i>	15.6–1000	[46,156]
	<i>Microsporium gypseum</i>	7.8–1000	[46,156]
	<i>Trichophyton mentagrophytes</i>	15.6–1000	[46,156]
	<i>Trichophyton rubrum</i>	7.8–1000	[46,156]
<i>Hyssopus officinalis</i> L.	<i>Aspergillus niger</i>	52,200	[47]
	<i>Aspergillus ochraceus</i>	26,100	[47]
	<i>Aspergillus versicolor</i>	10,440	[47]
	<i>Candida albicans</i>	128–1000	[44,48]
	<i>Candida glabrata</i>	512–1024	[48]
	<i>Candida krusei</i>	128–256	[48]
	<i>Candida parapsilosis</i>	256–512	[48]
	<i>Candida tropicalis</i>	512–1024	[48]
	<i>Cladosporium cladosporioides</i>	10,440	[47]
	<i>Cladosporium fulvum</i>	26,100	[47]
	<i>Penicillium funiculosum</i>	52,200	[47]
	<i>Penicillium ochrochloron</i>	26,100	[47]
	<i>Trichoderma viride</i>	10,440	[47]
<i>Lavandula angustifolia</i> Mill.	<i>Candida albicans</i>	0.125–512	[50,51,157]
	<i>Malassezia furfur</i>	>4	[49]
	<i>Trichophyton rubrum</i>	1–512	[49,51]
	<i>Trichosporon beigelii</i>	2	[49]
<i>Lavandula multifida</i> L.	<i>Aspergillus flavus</i>	0.64	[52]
	<i>Aspergillus fumigatus</i>	0.32	[52]
	<i>Aspergillus niger</i>	0.32	[52]
	<i>Candida albicans</i>	0.32	[52]
	<i>Candida guilliermondii</i>	0.32	[52]
	<i>Candida krusei</i>	0.64	[52]
	<i>Candida parapsilosis</i>	0.32	[52]
	<i>Candida tropicalis</i>	0.32	[52]
	<i>Cryptococcus neoformans</i>	0.16	[52]
	<i>Epidermophyton floccosum</i>	0.16	[52]
	<i>Microsporium canis</i>	0.16	[52]
	<i>Microsporium gypseum</i>	0.16	[52]
	<i>Trichophyton mentagrophytes</i>	0.16	[52]
	<i>Trichophyton mentagrophytes</i> var. <i>interdigitale</i>	0.16	[52]
	<i>Trichophyton rubrum</i>	0.16	[52]
<i>Trichophyton verrucosum</i>	0.16	[52]	

Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
<i>Lavandula pedunculata</i> (Miller) Cav.	<i>Aspergillus flavus</i>	5–10	[53]
	<i>Aspergillus fumigatus</i>	2.5–5	[53]
	<i>Aspergillus niger</i>	5	[53]
	<i>Candida albicans</i>	2.5	[53]
	<i>Candida guilliermondii</i>	1.25	[53]
	<i>Candida krusei</i>	1.25–2.5	[53]
	<i>Candida parapsilosis</i>	2.5–5	[53]
	<i>Candida tropicalis</i>	1.25–2.5	[53]
	<i>Cryptococcus neoformans</i>	0.32–1.25	[53]
	<i>Epidermophyton floccosum</i>	0.32–0.64	[53]
	<i>Microsporium canis</i>	0.32–1.25	[53]
	<i>Microsporium gypseum</i>	0.64–2.5	[53]
	<i>Trichophyton mentagrophytes</i>	0.64–1.25	[53]
<i>Trichophyton rubrum</i>	0.32–1.25	[53]	
<i>Lavandula stoechas</i> L.	<i>Aspergillus flavus</i>	1.25–10	[54]
	<i>Aspergillus fumigatus</i>	0.64–1.25	[54]
	<i>Aspergillus niger</i>	0.32–1.25	[54]
	<i>Candida albicans</i>	0.64–512	[51,54]
	<i>Candida guilliermondii</i>	1.25	[54]
	<i>Candida krusei</i>	2.5	[54]
	<i>Candida parapsilosis</i>	2.5	[54]
	<i>Candida tropicalis</i>	2.5	[54]
	<i>Cryptococcus neoformans</i>	0.64	[54]
	<i>Epidermophyton floccosum</i>	0.16–0.32	[54]
	<i>Microsporium canis</i>	0.16–0.64	[54]
	<i>Microsporium gypseum</i>	0.32–0.64	[54]
	<i>Trichophyton mentagrophytes</i>	0.32–0.64	[54]
<i>Trichophyton mentagrophytes var. interdigitale</i>	0.16–0.64	[54]	
<i>Trichophyton rubrum</i>	0.16–256	[51,54]	
<i>Trichophyton verrucosum</i>	0.32	[54]	
<i>Lavandula viridis</i> L'Her.	<i>Aspergillus flavus</i>	5	[55]
	<i>Aspergillus fumigatus</i>	2.5	[55]
	<i>Aspergillus niger</i>	2.5	[55]
	<i>Candida albicans</i>	1.25–2.5	[55]
	<i>Candida guilliermondii</i>	0.64–1.25	[55]
	<i>Candida krusei</i>	1.25–2.5	[55]
	<i>Candida parapsilosis</i>	1.25	[55]
	<i>Candida tropicalis</i>	1.25–2.5	[55]
	<i>Cryptococcus neoformans</i>	0.64	[55]
	<i>Epidermophyton floccosum</i>	0.32	[55]
	<i>Microsporium canis</i>	0.32	[55]
	<i>Microsporium gypseum</i>	0.64	[55]
	<i>Trichophyton mentagrophytes</i>	0.32–0.64	[55]
<i>Trichophyton mentagrophytes var. interdigitale</i>	0.32–0.64	[55]	
<i>Trichophyton rubrum</i>	0.32	[55]	
<i>Trichophyton verrucosum</i>	0.32	[55]	
<i>Lepechinia mutica</i> (Benth.) Epling	<i>Candida albicans</i>	>9000	[56]
	<i>Fusarium graminearum</i>	>9000	[56]
	<i>Microsporium canis</i>	2200–4500	[56]
	<i>Pyricularia oryzae</i>	>9000	[56]
	<i>Trichophyton rubrum</i>	2200–4500	[56]
<i>Marrubium vulgare</i> L.	<i>Aspergillus niger</i>	>1180	[58]
	<i>Botrytis cinerea</i>	>1100	[58]
	<i>Fusarium solani</i>	>1190	[58]
	<i>Penicillium digitatum</i>	>1120	[58]
<i>Melissa officinalis</i> L.	<i>Aspergillus niger</i>	313	[158]
	<i>Candida albicans</i>	30–313	[59,158]
	<i>Cryptococcus neoformans</i>	78	[158]
	<i>Epidermophyton floccosum</i>	30	[59]
	<i>Microsporium canis</i>	30	[59]
	<i>Penicillium verrucosum</i>	125	[159]
	<i>Trichophyton mentagrophytes var. mentagrophytes</i>	15	[59]
<i>Trichophyton rubrum</i>	15	[59]	
<i>Trichophyton tonsurans</i>	15	[59]	

Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
<i>Mentha cervina</i> L.	<i>Aspergillus flavus</i>	2.5–5	[60]
	<i>Aspergillus fumigatus</i>	1.25–2.5	[60]
	<i>Aspergillus niger</i>	1.25–2.5	[60]
	<i>Candida albicans</i>	1.25–2.5	[60]
	<i>Candida guilliermondii</i>	1.25–2.5	[60]
	<i>Candida krusei</i>	1.25–2.5	[60]
	<i>Candida parapsilosis</i>	1.25–2.5	[60]
	<i>Candida tropicalis</i>	1.25–2.5	[60]
	<i>Cryptococcus neoformans</i>	1.25	[60]
	<i>Epidermophyton floccosum</i>	0.64–1.25	[60]
	<i>Microsporium canis</i>	1.25	[60]
	<i>Microsporium gypseum</i>	1.25–2.5	[60]
	<i>Trichophyton mentagrophytes</i>	1.25–2.5	[60]
<i>Trichophyton rubrum</i>	1.25	[60]	
<i>Mentha × piperita</i> L.	<i>Aspergillus flavus</i>	1450–5000	[62,64]
	<i>Aspergillus niger</i>	625–10,000	[64,158]
	<i>Aspergillus parasiticus</i>	2500	[64]
	<i>Candida albicans</i>	225–1125	[63,158,160]
	<i>Candida glabrata</i>	225	[62]
	<i>Candida tropicalis</i>	225–230	[62]
	<i>Cryptococcus neoformans</i>	313	[158]
	<i>Fusarium oxysporum</i>	125	[161]
	<i>Penicillium chrysogenum</i>	1250	[64]
	<i>Penicillium minioluteum</i>	2050–2200	[62]
	<i>Penicillium oxalicum</i>	1300–2050	[62]
	<i>Penicillium verrucosum</i>	2500	[90]
	<i>Mentha pulegium</i> L.	<i>Aspergillus niger</i>	0.25–1.25
<i>Aspergillus flavus</i>		1.25	[162]
<i>Aspergillus fumigatus</i>		1.25	[162]
<i>Candida albicans</i>		0.94–3.75	[65,66,162,163]
<i>Candida bracarensis</i>		3.75	[163]
<i>Candida guilliermondii</i>		1.25	[162]
<i>Candida krusei</i>		0.94–1.25	[162,163]
<i>Candida parapsilosis</i>		1.25	[162]
<i>Candida tropicalis</i>		1.25	[162]
<i>Cryptococcus neoformans</i>		0.64	[162]
<i>Epidermophyton floccosum</i>		1.25	[162]
<i>Microsporium canis</i>		1.25	[162]
<i>Microsporium gypseum</i>		1.25–2.5	[162]
<i>Saccharomyces cerevisiae</i>		<0.3–0.94	[66,163]
<i>Trichophyton mentagrophytes</i>		1.25–2.5	[162]
<i>Trichophyton mentagrophytes var. interdigitale</i>	2.5	[162]	
<i>Trichophyton rubrum</i>	1.25	[162]	
<i>Trichophyton verrucosum</i>	1.25	[162]	
<i>Mentha requienii</i> Bentham	<i>Alternaria</i> spp.	>40	[67]
	<i>Aspergillus fumigatus</i>	>60	[67]
	<i>Candida albicans</i>	0.94–40	[67,163]
	<i>Candida bracarensis</i>	3.75	[163]
	<i>Candida krusei</i>	0.94	[163]
	<i>Fusarium</i> spp.	>40	[67]
	<i>Penicillium</i> spp.	>60	[67]
	<i>Rhodotorula</i> spp.	45	[67]
	<i>Saccharomyces cerevisiae</i>	0.94	[163]
<i>Mentha spicata</i> L.	<i>Aspergillus flavus</i>	1.25	[162]
	<i>Aspergillus fumigatus</i>	0.64	[162]
	<i>Aspergillus niger</i>	0.64–313	[158,162]
	<i>Candida albicans</i>	1.25–625	[51,158,162]
	<i>Candida guilliermondii</i>	1.25	[162]
	<i>Candida krusei</i>	1.25	[162]
	<i>Candida parapsilosis</i>	1.25	[162]
	<i>Candida tropicalis</i>	1.25	[162]
	<i>Cryptococcus neoformans</i>	0.32–313	[158,162]
	<i>Epidermophyton floccosum</i>	0.64	[162]
	<i>Fusarium graminearum</i>	2.5	[164]
<i>Fusarium moniliforme</i>	2.5	[164]	
<i>Malassezia furfur</i>	>4	[49]	
<i>Microsporium canis</i>	0.64–2	[68,162]	

Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
	<i>Microsporium gypseum</i>	0.64–3	[162]
	<i>Penicillium corylophilum</i>	0.625	[165]
	<i>Penicillium expansum</i>	2.5	[164]
	<i>Trichophyton erinacei</i>	3	[68]
	<i>Trichophyton mentagrophytes</i>	0.64–3	[68,162]
	<i>Trichophyton mentagrophytes</i> var. <i>interdigitale</i>	0.64	[162]
	<i>Trichophyton rubrum</i>	0.25–512	[49,51,162]
	<i>Trichophyton terrestre</i>	3	[68]
	<i>Trichophyton verrucosum</i>	0.32	[162]
	<i>Trichosporon beigeli</i>	0.25	[49]
	<i>Candida albicans</i>	0.34–1250	[69,71,166]
	<i>Candida glabrata</i>	0.69–2.77	[69]
	<i>Cryptococcus neoformans</i>	300	[167]
	<i>Microsporium canis</i>	1250	[167]
	<i>Microsporium gypseum</i>	1250	[167]
	<i>Trichophyton mentagrophytes</i>	600–1250	[167]
	<i>Trichophyton rubrum</i>	5000	[167]
	<i>Trichophyton violaceum</i>	600	[167]
	<i>Aspergillus niger</i>	0.2	[38]
	<i>Aspergillus ochraceus</i>	0.2	[38]
	<i>Cladosporium cladosporioides</i>	0.2	[38]
	<i>Fusarium tricinctum</i>	0.4	[38]
	<i>Penicillium ochrochloron</i>	0.2	[38]
	<i>Phomopsis helianthi</i>	0.2	[38]
	<i>Trichoderma viride</i>	0.4	[38]
	<i>Aspergillus niger</i>	50	[72]
	<i>Candida albicans</i>	100	[72]
	<i>Fusarium oxysporum</i>	100	[72]
	<i>Alternaria solani</i>	3000	[43]
	<i>Candida albicans</i>	0.78	[168]
	<i>Fusarium oxysporum</i>	1000	[43]
	<i>Trichophyton rubrum</i>	0.19	[168]
	<i>Helminthosporium maydis</i>	1500	[43]
	<i>Alternaria solani</i>	3000	[43]
	<i>Fusarium oxysporum</i>	2000	[43]
	<i>Helminthosporium maydis</i>	2000	[43]
	<i>Aspergillus flavus</i>	10,000	[64]
	<i>Aspergillus fumigatus</i>	>50	[75]
	<i>Aspergillus niger</i>	>50–10,000	[64,75,158]
	<i>Aspergillus parasiticus</i>	5000	[64]
	<i>Candida albicans</i>	30–625	[73,74,158]
	<i>Candida guilliermondii</i>	3.125–6.25	[76]
	<i>Cryptococcus neoformans</i>	313–1250	[158,169]
	<i>Debaryomyces hansenii</i>	6.25	[76]
	<i>Epidermophyton floccosum</i>	15	[74]
	<i>Microsporium canis</i>	1–15.2	[68,74]
	<i>Microsporium gypseum</i>	3	[68]
	<i>Penicillium chrysogenum</i>	10,000	[64]
	<i>Penicillium italicum</i>	>50	[75]
	<i>Rhizopus stolonifer</i>	>50	[75]
	<i>Rhodotorula glutinis</i>	86	[73]
	<i>Trichophyton erinacei</i>	2.5	[68]
	<i>Trichophyton mentagrophytes</i>	2.5–8.3	[68,74]
	<i>Trichophyton terrestre</i>	3	[68]
	<i>Saccharomyces cerevisiae</i>	28	[73]
	<i>Schizosaccharomyces pombe</i>	86	[73]
	<i>Trichophyton rubrum</i>	8.3	[74]
	<i>Trichophyton tonsurans</i>	8	[74]
	<i>Yarrowia lipolytica</i>	57	[73]
	<i>Candida guilliermondii</i>	3.125	[76]
	<i>Debaryomyces hansenii</i>	1.56	[76]

Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
<i>Ocimum campechianum</i> Mill. = <i>Ocimum micranthum</i> Willd.	<i>Candida albicans</i>	69	[73]
	<i>Rhodotorula glutinis</i>	139	[73]
	<i>Saccharomyces cerevisiae</i>	69	[73]
	<i>Schizosaccharomyces pombe</i>	104	[73]
	<i>Yarrowia lipolytica</i>	69	[73]
<i>Ocimum forskolei</i> Benth.	<i>Candida albicans</i>	35.3–8600	[77,170]
<i>Ocimum gratissimum</i> L.	<i>Aspergillus fumigatus</i>	>1000	[78]
	<i>Candida albicans</i>	350–1500	[78,171]
	<i>Candida krusei</i>	750	[171]
	<i>Candida parapsilosis</i>	380	[171]
	<i>Candida tropicalis</i>	1500	[171]
	<i>Cryptococcus neoformans</i>	250–300	[78,79]
	<i>Fusarium oxysporum</i> f. sp. <i>cubense</i>	62.5	[80]
	<i>Fusarium oxysporum</i> f. sp. <i>lycopersici</i>	31.25	[80]
	<i>Fusarium oxysporum</i> f. sp. <i>tracheiphilum</i>	62.5	[80]
	<i>Fusarium solani</i>	62.5	[80]
	<i>Macrophomina phaseolina</i>	62.5–125	[80]
	<i>Malassezia pachydermatis</i>	300	[78]
	<i>Microsporum canis</i>	200–500	[78,172]
	<i>Microsporum gypseum</i>	150–250	[78,172]
	<i>Rhizoctonia solani</i>	31.25	[80]
	<i>Scopulariopsis brevicaulis</i>	400	[78]
	<i>Trichophyton interdigitale</i>	250	[78]
<i>Trichophyton mentagrophytes</i>	200–250	[78,172]	
<i>Trichophyton rubrum</i>	150–250	[78,172]	
<i>Ocimum tenuiflorum</i> L. = <i>Ocimum sanctum</i> L.	<i>Aspergillus flavus</i>	300	[83]
	<i>Candida albicans</i>	0.1–300	[81,82]
	<i>Candida glabrata</i>	0.15–300	[81,82]
	<i>Candida krusei</i>	0.35–450	[81,82]
	<i>Candida parapsilosis</i>	0.25–500	[81,82]
	<i>Candida tropicalis</i>	0.1–300	[81,82]
<i>Origanum compactum</i> Benth.	<i>Alternaria alternata</i>	300	[84]
	<i>Bipolaris oryzae</i>	300	[84]
	<i>Fusarium equiseti</i>	300	[84]
	<i>Fusarium graminearum</i>	300	[84]
	<i>Fusarium verticillioides</i>	300	[84]
<i>Origanum majorana</i> L.	<i>Aspergillus flavus</i>	450–650	[62]
	<i>Aspergillus niger</i>	625	[158]
	<i>Botrytis cinerea</i>	5000	[87]
	<i>Candida albicans</i>	625	[158]
	<i>Cryptococcus neoformans</i>	313	[158]
	<i>Fusarium delphinoides</i>	1800–14,400	[85]
	<i>Fusarium incarnatum-equiseti</i>	450–3600	[85]
	<i>Fusarium napiforme</i>	3600–14,400	[85]
	<i>Fusarium oxysporum</i>	900–3600	[85]
	<i>Fusarium solani</i>	900–3600	[85]
	<i>Fusarium verticillioides</i>	14,400	[85]
	<i>Microsporum canis</i>	0.5	[68]
	<i>Microsporum gypseum</i>	2	[68]
	<i>Penicillium expansum</i>	10,000	[87]
	<i>Penicillium minioluteum</i>	400–500	[62]
	<i>Penicillium oxalicum</i>	350–400	[62]
	<i>Sporothrix brasiliensis</i>	$\leq 2250$ –9000	[86]
	<i>Sporothrix schenckii</i>	$\leq 2250$ –9000	[86]
	<i>Trichophyton erinacei</i>	1	[68]
<i>Trichophyton mentagrophytes</i>	1.5	[68]	
<i>Trichophyton terrestris</i>	2	[68]	



Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
<i>Origanum vulgare</i> L.	<i>Aspergillus flavus</i>	0.64–2500	[64,89,91]
	<i>Aspergillus fumigatus</i>	0.32–0.64	[89]
	<i>Aspergillus niger</i>	0.32–623	[62,89,91,158]
	<i>Aspergillus ochraceus</i>	470	[91]
	<i>Aspergillus parasiticus</i>	2500	[64]
	<i>Candida albicans</i>	0.32–700	[74,88,89,91,158]
	<i>Candida glabrata</i>	350	[88]
	<i>Candida guilliermondii</i>	0.64–1.25	[89]
	<i>Candida krusei</i>	0.64–700	[88,89]
	<i>Candida parapsilosis</i>	0.64–170	[88,89]
	<i>Candida tropicalis</i>	0.32–700	[88,89]
	<i>Cladosporium</i> sp.	0.05–0.3	[173]
	<i>Cryptococcus neoformans</i>	0.16–78	[89,158]
	<i>Epidermophyton floccosum</i>	0.32–2	[74,89]
	<i>Fusarium</i> sp.	0.1–0.5	[173]
	<i>Malassezia furfur</i>	1–780	[49,174]
	<i>Microsporium canis</i>	0.025–2	[68,74,89]
	<i>Microsporium gypseum</i>	0.025–1.25	[68,89]
	<i>Penicillium</i> sp.	0.1–0.5	[173]
	<i>Penicillium chrysogenum</i>	625	[64]
	<i>Penicillium corylophilum</i>	0.625	[165]
	<i>Penicillium funiculosum</i>	610	[91]
	<i>Penicillium ochrochloron</i>	710	[91]
	<i>Penicillium verrucosum</i>	1.1719	[90,91]
	<i>Trichophyton mentagrophytes</i>	0.32–1.25	[74,89]
	<i>Trichophyton rubrum</i>	0.16–1.25	[49,74,89]
	<i>Trichophyton tonsurans</i>	1	[74]
	<i>Trichosporon beigeli</i>	0.25	[49]
<i>Trichophyton erinacei</i>	0.5	[68]	
<i>Trichophyton mentagrophytes</i>	0.5	[68]	
<i>Trichophyton terrestre</i>	0.25	[68]	
<i>Pogostemon cablin</i> (Blanco) Benth.	<i>Aspergillus flavus</i>	>1500	[92]
	<i>Aspergillus niger</i>	156	[158]
	<i>Aspergillus oryzae</i>	>1500	[92]
	<i>Candida albicans</i>	32–625	[158,175]
	<i>Candida krusei</i>	64–257	[175]
	<i>Candida tropicalis</i>	32–257	[175]
<i>Pogostemon heyneanus</i> Benth.	<i>Cryptococcus neoformans</i>	20	[158]
	<i>Candida albicans</i>	6000	[176]
	<i>Candida glabrata</i>	6000	[176]
<i>Premna microphylla</i> Turcz.	<i>Candida tropicalis</i>	10,000	[176]
	<i>Aspergillus niger</i>	>500	[94]
	<i>Candida albicans</i>	>500	[94]
<i>Rosmarinus officinalis</i> L.	<i>Fusarium oxysporum</i>	>500	[94]
	<i>Aspergillus flavus</i>	330	[91]
	<i>Aspergillus ochraceus</i>	590	[91]
	<i>Aspergillus niger</i>	380–10,000	[91,98,158]
	<i>Botrytis cinerea</i>	2500	[87]
	<i>Candida albicans</i>	30.2–1000	[51,91,96,98,158]
	<i>Cryptococcus neoformans</i>	313	[158]
	<i>Epidermophyton floccosum</i>	30	[96]
	<i>Microsporium canis</i>	2.5–30.2	[68,96]
	<i>Microsporium gypseum</i>	2.5	[68]
	<i>Penicillium expansum</i>	5000	[87]
	<i>Penicillium ochrochloron</i>	470	[91]
	<i>Penicillium funiculosum</i>	570	[91]
	<i>Trichophyton erinacei</i>	1.5	[68]
<i>Trichophyton mentagrophytes</i>	5–15.3	[68,96]	
<i>Trichophyton rubrum</i>	15–256	[51,96]	
<i>Trichophyton terrestre</i>	5	[68]	
<i>Trichophyton tonsurans</i>	15.2	[96]	

Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
<i>Salvia fruticosa</i> Miller	<i>Candida albicans</i>	512	[51]
	<i>Fusarium oxysporum</i> f. sp. <i>dianthi</i>	>2000	[99]
	<i>Fusarium proliferatum</i>	>2000	[99]
	<i>Fusarium solani</i> f. sp. <i>cucurbitae</i>	>2000	[99]
	<i>Malassezia furfur</i>	>4	[99]
	<i>Rhizoctonia solani</i>	>2000	[99]
	<i>Sclerotinia sclerotiorum</i>	>2000	[99]
	<i>Trichophyton rubrum</i>	2–256	[49,99]
	<i>Trichosporon beigelii</i>	4	[49]
<i>Salvia mirzayanii</i> Rech. f. and Esfand	<i>Candida albicans</i>	0.5–2	[100]
	<i>Candida krusei</i>	1	[100]
	<i>Candida dubliniensis</i>	0.06–0.5	[100]
	<i>Candida glabrata</i>	0.06–1	[100]
	<i>Candida parapsilosis</i>	0.25–1	[100]
	<i>Candida tropicalis</i>	0.25–2	[100]
	<i>Trichosporon</i> sp.	1	[100]
<i>Salvia officinalis</i> L.	<i>Aspergillus flavus</i>	5–10	[101]
	<i>Aspergillus fumigatus</i>	2.5–5	[101]
	<i>Aspergillus niger</i>	5–1250	[101,158]
	<i>Candida albicans</i>	2.5–2780	[96,101,158,177]
	<i>Candida guilliermondii</i>	1.25–2.5	[101]
	<i>Candida krusei</i>	2.5–5	[101]
	<i>Candida parapsilosis</i>	5	[101]
	<i>Candida tropicalis</i>	5	[101]
	<i>Cryptococcus neoformans</i>	0.64–625	[101,158]
	<i>Epidermophyton floccosum</i>	0.64–100	[96,101]
	<i>Microsporium canis</i>	1.25–100.2	[96,101]
	<i>Microsporium gypseum</i>	1.25–2.5	[101]
	<i>Trichophyton mentagrophytes</i>	1.25–60	[96,101]
	<i>Trichophyton mentagrophytes</i> var. <i>interdigitale</i>	1.25	[101]
	<i>Trichophyton rubrum</i>	0.64–60	[96,101]
<i>Trichophyton tonsurans</i>	60	[96]	
<i>Trichophyton verrucosum</i>	1.25–2.5	[101]	
<i>Salvia sclarea</i> L.	<i>Aspergillus niger</i>	1250	[158]
	<i>Candida albicans</i>	1250	[158]
	<i>Cryptococcus neoformans</i>	313	[158]
	<i>Fusarium delphinoides</i>	1800–3600	[85]
	<i>Fusarium incarnatum-equiseti</i>	1800–3600	[85]
	<i>Fusarium napiforme</i>	1800–3600	[85]
	<i>Fusarium oxysporum</i>	1800–3600	[85]
	<i>Fusarium solani</i>	3600–7200	[85]
<i>Fusarium verticillioides</i>	1800	[85]	
<i>Satureja hortensis</i> L.	<i>Alternaria alternata</i>	62.5	[103]
	<i>Aspergillus flavus</i>	31.25–500	[103,104,117]
	<i>Aspergillus niger</i>	471	[117]
	<i>Aspergillus ochraceus</i>	423	[117]
	<i>Aspergillus parasiticus</i>	373	[117]
	<i>Aspergillus terreus</i>	389	[117]
	<i>Aspergillus varicolor</i>	125	[103]
	<i>Candida albicans</i>	200–400	[103,178]
	<i>Fusarium culmorum</i>	125	[103]
	<i>Fusarium oxysporum</i>	250	[103]
	<i>Microsporium canis</i>	62.5	[103]
	<i>Monilia fructicola</i>	31.25	[103]
	<i>Penicillium</i> spp.	125	[103]
	<i>Rhizoctonia solani</i>	125	[103]
	<i>Rhizopus</i> spp.	250	[103]
<i>Sclerotinia minor</i>	250	[103]	
<i>Sclerotinia sclerotiorum</i>	125	[103]	
<i>Trichophyton mentagrophytes</i>	62.5	[103]	
<i>Trichophyton rubrum</i>	31.25	[103]	
<i>Satureja montana</i> L.	<i>Microsporium canis</i>	0.5	[68]
	<i>Microsporium gypseum</i>	2	[68]
	<i>Trichophyton erinacei</i>	2	[68]
	<i>Trichophyton mentagrophytes</i>	2	[68]
	<i>Trichophyton terrestris</i>	3	[68]

Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
<i>Satureja thymbra</i> L.	<i>Aspergillus flavus</i>	25	[105]
	<i>Aspergillus fumigatus</i>	1.25–25	[105,179]
	<i>Aspergillus niger</i>	2.5–25	[105,179]
	<i>Aspergillus ochraceus</i>	2.5–25	[105,179]
	<i>Aspergillus versicolor</i>	1.25	[179]
	<i>Candida albicans</i>	25–128	[51,105]
	<i>Penicillium funiculosum</i>	2.5–25	[105,179]
	<i>Penicillium ochrochloron</i>	1–1.25	[105,179]
	<i>Trichoderma viride</i>	1.25–25	[105,179]
	<i>Trichophyton rubrum</i>	128	[51]
<i>Stachys cretica</i> L.	<i>Candida albicans</i>	625	[106]
<i>Stachys officinalis</i> (L.) Trevis	<i>Aspergillus niger</i>	2500	[107]
	<i>Candida albicans</i>	5000	[107]
<i>Stachys pubescens</i> Ten.	<i>Alternaria alternata</i>	1	[108]
	<i>Aspergillus flavus</i>	0–5	[108]
	<i>Fusarium oxysporum</i>	1	[108]
<i>Teucrium sauvagei</i> Le Houerou	<i>Aspergillus fumigatus</i>	>1000	[109]
	<i>Candida albicans</i>	>1000	[109]
	<i>Cryptococcus neoformans</i>	>1000	[109]
	<i>Epidermophyton floccosum</i>	850	[109]
	<i>Microsporum canis</i>	800	[109]
	<i>Microsporum gypseum</i>	900	[109]
	<i>Scopulariopsis brevicaulis</i>	>1000	[109]
	<i>Scytalidium dimidiatum</i>	>1000	[109]
	<i>Trichophyton mentagrophytes</i> var. <i>interdigitale</i>	950	[109]
	<i>Trichophyton mentagrophytes</i> var. <i>mentagrophytes</i>	900	[109]
<i>Teucrium yemense</i> Deflers.	<i>Aspergillus niger</i>	313	[77]
	<i>Botrytis cinerea</i>	313	[77]
	<i>Candida albicans</i>	1250	[77]
<i>Thymbra capitata</i> (L.) Cav. = <i>Thymus capitatus</i> (L.) Hoffmanns. and Link = <i>Coridothymus capitatus</i> (L.) Rchb.f. Solms	<i>Aspergillus flavus</i>	0.32	[111]
	<i>Aspergillus fumigatus</i>	0.16–0.32	[111]
	<i>Aspergillus niger</i>	0.1–0.16	[111,180]
	<i>Aspergillus oryzae</i>	0.2	[180]
	<i>Candida albicans</i>	0.16–128	[51,110–112]
	<i>Candida glabrata</i>	0.32	[111,112]
	<i>Candida guilliermondii</i>	0.16–0.32	[111,112]
	<i>Candida krusei</i>	0.32	[111]
	<i>Candida parapsilosis</i>	0.32	[111,112]
	<i>Candida tropicalis</i>	0.32	[111,112]
	<i>Epidermophyton floccosum</i>	0.08	[111]
	<i>Fusarium solani</i>	0.2	[180]
	<i>Microsporum canis</i>	0.08	[111]
	<i>Microsporum gypseum</i>	0.08	[111]
	<i>Penicillium digitatum</i>	0.5	[180]
	<i>Trichophyton mentagrophytes</i>	0.08	[111]
<i>Trichophyton rubrum</i>	0.16–64	[51,111]	
<i>Thymbra spicata</i> L.	<i>Aspergillus fumigatus</i>	0.3	[179]
	<i>Aspergillus niger</i>	0.6	[179]
	<i>Aspergillus versicolor</i>	0.3	[179]
	<i>Aspergillus ochraceus</i>	0.6	[179]
	<i>Candida albicans</i>	1.12–3750	[51,113,114]
	<i>Candida krusei</i>	1.12	[114]
	<i>Candida parapsilosis</i>	0.6–1.12	[114]
	<i>Penicillium funiculosum</i>	0.3	[179]
	<i>Penicillium ochrochloron</i>	0.3	[179]
<i>Trichoderma viride</i>	0.3	[179]	
<i>Trichophyton rubrum</i>	64	[51]	
<i>Thymus bovei</i> Benth.	<i>Candida albicans</i>	250	[115]
<i>Thymus daenensis</i> Celak.	<i>Alternaria alternata</i>	>8	[108]
	<i>Aspergillus flavus</i>	1	[108]
	<i>Fusarium oxysporum</i>	4	[108]

Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
<i>Thymus kotschyanus</i> Boiss. and Hohen.	<i>Alternaria alternata</i>	1	[108]
	<i>Aspergillus flavus</i>	0.5	[108]
	<i>Fusarium oxysporum</i>	0–5	[108]
<i>Thymus mastichina</i> (L.) L.	<i>Candida albicans</i>	1.25–2.5	[116]
	<i>Candida glabrata</i>	1.25–1.5	[116]
	<i>Candida guilliermondii</i>	1.25	[116]
	<i>Candida krusei</i>	1.25–2.5	[116]
	<i>Candida parapsilosis</i>	2.5–5	[116]
<i>Thymus migricus</i> Klokov et Des.-Shost.	<i>Candida tropicalis</i>	2.5–10	[116]
	<i>Aspergillus flavus</i>	452	[117]
<i>Thymus migricus</i> Klokov et Des.-Shost.	<i>Aspergillus niger</i>	460	[117]
	<i>Aspergillus ochraceus</i>	430	[117]
	<i>Aspergillus parasiticus</i>	581	[117]
<i>Thymus pulegioides</i> L.	<i>Aspergillus terreus</i>	447	[117]
	<i>Aspergillus flavus</i>	0.32	[119]
	<i>Aspergillus fumigatus</i>	0.16	[119]
	<i>Aspergillus niger</i>	0.32	[119]
	<i>Candida albicans</i>	0.32–0.64	[119]
	<i>Candida glabrata</i>	0.32–0.64	[119]
	<i>Candida guilliermondii</i>	0.32	[119]
	<i>Candida krusei</i>	0.32–0.64	[119]
	<i>Candida parapsilosis</i>	0.64	[119]
	<i>Candida tropicalis</i>	0.32–0.64	[119]
	<i>Epidermophyton floccosum</i>	0.16	[119]
	<i>Microsporium canis</i>	0.16	[119]
	<i>Microsporium gypseum</i>	0.16	[119]
	<i>Trichophyton mentagrophytes</i>	0.16	[119]
<i>Thymus schimperi</i> Ronninger	<i>Trichophyton rubrum</i>	0.32	[119]
	<i>Aspergillus minutus</i>	0.512–2	[120]
	<i>Aspergillus niger</i>	0.16	[181]
	<i>Aspergillus tubingensis</i>	1–4	[120]
	<i>Beauveria bassiana</i>	0.128–1	[120]
	<i>Candida albicans</i>	0.16	[181]
	<i>Microsporium</i> spp.	0.08	[181]
	<i>Microsporium gypseum</i>	0.128–1	[120]
	<i>Penicillium chrysogenum</i>	0.512–2	[120]
	<i>Rhodotorula</i> spp.	0.08	[181]
	<i>Trichophyton</i> spp.	0.08–0.31	[181]
<i>Verticillium</i> sp.	0.512–2	[120]	
<i>Thymus serpyllum</i> L.	<i>Aspergillus carbonarius</i>	1.25	[182]
	<i>Aspergillus ochraceus</i>	0.625	[182]
	<i>Aspergillus niger</i>	2.5	[182]
	<i>Microsporium canis</i>	0.025	[68]
	<i>Microsporium gypseum</i>	0.25	[68]
	<i>Trichophyton erinacei</i>	0.1	[68]
	<i>Trichophyton mentagrophytes</i>	0.2	[68]
<i>Thymus striatus</i> Vahl.	<i>Trichophyton terrestre</i>	0.1	[68]
	<i>Alternaria alternata</i>	1	[121]
	<i>Aspergillus flavus</i>	1.5	[121]
	<i>Aspergillus niger</i>	1	[121]
	<i>Aspergillus ochraceus</i>	1	[121]
	<i>Aspergillus terreus</i>	1	[121]
	<i>Aspergillus versicolor</i>	1	[121]
	<i>Cladosporium cladosporioides</i>	0.5	[121]
	<i>Epidermophyton floccosum</i>	1	[121]
	<i>Microsporium canis</i>	1.5	[121]
<i>Thymus striatus</i> Vahl.	<i>Penicillium funiculosum</i>	2	[121]
	<i>Penicillium ochrochloron</i>	2	[121]
	<i>Phomopsis helianthi</i>	0.5	[121]
	<i>Trichoderma viride</i>	2	[121]
	<i>Trichophyton mentagrophytes</i>	1	[121]

Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
<i>Thymus vulgaris</i> L.	<i>Absidia</i> spp.	7 $\pm$ 4	[122]
	<i>Alternaria</i> spp.	9.4 $\pm$ 4.5	[122]
	<i>Alternaria alternata</i>	4.7–500	[122,183]
	<i>Aspergillus</i> spp.	3.2	[122]
	<i>Aspergillus flavus</i>	9.35–1500	[64,104,122,125,184]
	<i>Aspergillus fumigatus</i>	144–1000	[124,184]
	<i>Aspergillus niger</i>	9.35–1250	[64,122,158,184]
	<i>Aspergillus ochraceus</i>	2.5–750	[164,184]
	<i>Aspergillus parasiticus</i>	1250	[64]
	<i>Aspergillus sulphureus</i>	10.88 $\pm$ 3.1	[122]
	<i>Aspergillus versicolor</i>	9.6 $\pm$ 9.25	[122]
	<i>Botrytis cinerea</i>	312	[87]
	<i>Candida albicans</i>	0.16–313	[73,74,116,158]
	<i>Candida glabrata</i>	0.16–0.32	[116]
	<i>Candida krusei</i>	0.08–0.16	[116]
	<i>Candida guilliermondii</i>	0.16	[116]
	<i>Candida parapsilosis</i>	0.16–0.32	[116]
	<i>Candida tropicalis</i>	0.16–0.32	[116]
	<i>Chaetomium globosum</i>	1.6	[122]
	<i>Cladosporium</i> spp.	12.8	[122]
	<i>Cladosporium sphaerospermum</i>	19.6	[122]
	<i>Cryptococcus neoformans</i>	78	[158]
	<i>Epidermophyton floccosum</i>	4	[74]
	<i>Fusarium</i> spp.	62.5	[185]
	<i>Fusarium delphinoides</i>	900–1800	[85]
	<i>Fusarium incarnatum-equiseti</i>	450–3600	[85]
	<i>Fusarium napiforme</i>	900	[85]
	<i>Fusarium oxysporum</i>	5–900	[85,126]
	<i>Fusarium solani</i>	1800–3600	[85]
	<i>Fusarium verticillioides</i>	900	[85]
	<i>Malassezia furfur</i>	920	[174]
	<i>Microsporium canis</i>	2.2	[74]
	<i>Mortierella</i> spp.	250	[185]
	<i>Mucor</i> spp.	50.2 $\pm$ 8.4	[122]
	<i>Penicillium</i> spp.	18.95–500	[122,185]
	<i>Penicillium brevicompactum</i>	19.6	[122]
	<i>Penicillium chrysogenum</i>	312.5–1750	[64,184]
	<i>Penicillium chrysogenum</i>	19.6	[122]
	<i>Penicillium citrinum</i>	1250	[184]
	<i>Penicillium expansum</i>	625	[87]
	<i>Penicillium griseofulvum</i>	19.6	[122]
	<i>Rhizopus</i> spp.	12.6	[122]
	<i>Rhodotorula glutinis</i>	72	[73]
<i>Rhizopus oryzae</i>	256–512	[123]	
<i>Saccharomyces cerevisiae</i>	72	[73]	
<i>Schizosaccharomyces pombe</i>	36	[73]	
<i>Stachybotrys chartarum</i>	6.2	[122]	
<i>Trichoderma</i> spp.	16.8	[122]	
<i>Trichophyton mentagrophytes</i>	2.2	[74]	
<i>Trichophyton rubrum</i>	2–72	[74,124]	
<i>Trichophyton tonsurans</i>	2.2	[74]	
<i>Ulocladium</i> spp.	5.45 $\pm$ 1.5	[122]	
<i>Yarrowia lypolytica</i>	36	[73]	
<i>Thymus zygis</i> L.	<i>Candida albicans</i>	0.16–0.32	[116]
	<i>Candida glabrata</i>	0.32	[116]
	<i>Candida krusei</i>	0.16–0.32	[116]
	<i>Candida guilliermondii</i>	0.16	[116]
	<i>Candida parapsilosis</i>	0.32	[116]
	<i>Candida tropicalis</i>	0.16–0.32	[116]
	<i>Penicillium corylophilum</i>	0.3125–0.625	[165]



Table 3. Cont.

Source of the Essential Oil	Targeted Fungus	MICs ( $\mu\text{g/mL}$ ; $\mu\text{l/mL}$ )	Reference(s)
<i>Vitex agnus-castus</i> L.	<i>Candida albicans</i>	0.53–512	[51,129]
	<i>Candida dubliniensis</i>	0.27	[129]
	<i>Candida famata</i>	2.13	[129]
	<i>Candida glabrata</i>	0.27	[129]
	<i>Candida krusei</i>	0.27	[129]
	<i>Candida lusitanae</i>	2.13	[129]
	<i>Candida parapsilosis</i>	1.06	[129]
	<i>Candida tropicalis</i>	0.13	[129]
	<i>Epidermophyton floccosum</i>	0.64–2.5	[128]
	<i>Microsporium canis</i>	0.64–5	[128]
	<i>Microsporium gypseum</i>	1.25–10	[128]
	<i>Trichophyton mentagrophytes</i>	1.25–10	[128]
	<i>Trichophyton rubrum</i>	0.64–512	[51,128]
<i>Zataria multiflora</i> Boiss.	<i>Aspergillus flavus</i>	358	[117]
	<i>Aspergillus niger</i>	358	[117]
	<i>Aspergillus ochraceus</i>	341	[117]
	<i>Aspergillus parasiticus</i>	367	[117]
	<i>Aspergillus terreus</i>	447	[117]
	<i>Microsporium canis</i>	0.125–0.25	[130]
	<i>Microsporium gypseum</i>	0.03–0.06	[130]
	<i>Trichophyton mentagrophytes</i>	0.03	[130]
	<i>Trichophyton rubrum</i>	0.03–0.06	[130]
	<i>Trichophyton schoenleinii</i>	0.125–0.6	[130]
<i>Ziziphora clinopodioides</i> Lam.	<i>Aspergillus flavus</i>	48.82	[184,186]
	<i>Aspergillus fumigatus</i>	1750	[184]
	<i>Aspergillus niger</i>	3000	[184]
	<i>Aspergillus ochraceus</i>	1500	[184]
	<i>Aspergillus parasiticus</i>	48.82	[186]
	<i>Penicillium chrysogenum</i>	3000	[184]
	<i>Penicillium citrinum</i>	1750	[184]
<i>Ziziphora tenuior</i> L.	<i>Aspergillus flavus</i>	1.25	[133]
	<i>Aspergillus fumigatus</i>	0.64	[133]
	<i>Aspergillus niger</i>	0.64	[133]
	<i>Candida albicans</i>	1.25	[133]
	<i>Candida guilliermondii</i>	1.25	[133]
	<i>Candida krusei</i>	1.25	[133]
	<i>Candida parapsilosis</i>	1.25	[133]
	<i>Candida tropicalis</i>	1.25	[133]
	<i>Cryptococcus neoformans</i>	0.16	[133]
	<i>Epidermophyton floccosum</i>	0.64	[133]
	<i>Microsporium canis</i>	0.64–1.25	[133]
	<i>Microsporium gypseum</i>	1.25	[133]
	<i>Trichophyton mentagrophytes</i>	1.25	[133]
	<i>Trichophyton mentagrophytes</i> var. <i>interdigitale</i>	1.254	[133]
	<i>Trichophyton rubrum</i>	0.64	[133]
<i>Trichophyton verrucosum</i>	0.64	[133]	

#### 4. Essential Oils of Lamiaceae Plants in Cosmetics and Medicines

Some essential oils of Lamiaceae family plants and/or their components are commonly used in cosmetics and less often in medicine. Essential oils from *Thymus vulgaris*, *Origanum vulgare*, *Rosmarinus officinalis*, *Calamintha officinalis*, *Salvia officinalis*, or *Lavandula officinalis* are in cosmetic formulations as natural preservatives [187]. *Lavandula angustifolia* oil is commonly used as a fragrance in cosmetics, soaps, perfumes and pharmaceutical products. It also acts as an anti-inflammatory, and is calming, headache relieving, is a sedative and is skin healing. Essential oils from *Lavandula hybrida* and *L. angustifolia* also have anti-lice activity. Compounds (essential oils and mainly menthol) extracted from *Mentha piperita* are commonly used as a fragrance in soaps, cosmetics and as well as in the kitchen as a spice and refreshing products. Moreover, they are often found in chewing gums, toothpastes, and mouthwashes. For medical use, it can be taken orally in gastrointestinal complications. *Rosmarinus officinalis* essential oil is often an ingredient as a fragrance in cosmetics, soaps, bath salts and oils, gels and ointments. It is widely used for hair care and hair-loss treatment because it promotes hair growth and helps against dandruff [188]. In medicine, essential oils from Lamiaceae family are used in

aromatherapy (*Salvia sclarea*, *Lavandula officinalis*, *Mentha piperita*, *Rosmarinus officinalis*) [189], sinusitis (*Lavandula officinalis*, *Thymus vulgaris*) [190], and in upper respiratory tract for treatment of catarrh (*Mentha piperita*, *Mentha arvensis*, *Thymus* spp.) [191]. Both essential oils from Lamiaceae plants and mono-substances are used in toothpastes and mouthwashes. In many of these the following chemicals, like limonene, linalool, menthol, and thymol, are presented as flavorings and fragrances [192,193]. Additionally, in some toothpastes are essential oils, e.g., in “Parodontax®” occurs *Salvia officinalis* oil, *Mentha piperita* oil, and *Mentha arvensis* oil; in “Lacalut Active Herbal” is *Mentha arvensis* oil, *Thymus vulgaris* oil, and *Salvia officinalis* oil, while in “Signal Family Herbal Fresh” are oils from *Mentha piperita* and *Salvia officinalis* [194]. Literature data confirm a strong antifungal effect against *C. albicans* and anti-inflammatory activity of “Parodontax” toothpaste [195,196]. Besides toothpastes, also some medicines used to rinse the oral cavity or throat contain a large number of essential oils. Mention may be made of “Salviasept” having in its composition the oils from *Mentha × piperita*, *Thymus vulgaris*, *Thymus zygis*, *Origanum majorana*, and *Salvia officinalis* or “Dentosept Complex” containing oils from *Mentha piperita*, *Thymus vulgaris*, *Salvia* sp., *Lavandula* sp., and *Eucalyptus globulus*. Among the antifungal medicines in “Acerin Talk” antifungal foot deodorant are present *Lavandula* sp. oil, menthol, linalool, limonene, and geraniol, while in “Podoflex Tincture” for nails mucosis occur among others oils from *Salvia sclarea* and *Lavandula angustifolia* and mono-substances current in Lamiaceae plants: geraniol, limonene, linalool, citral, and eugenol [194].

## 5. Conclusions

More than half of the essential oils from Lamiaceae family plants have good antifungal activity (MICs < 1000 µg/mL). The microbiological data indicate that they could be used alone or in combination with antifungal drugs in the treatment of fungal infections, especially of the skin and mucous membranes. Some essential oils and their components extracted from Lamiaceae plants are used in cosmetics and medicines. Essential oils may be of future relevance in the treatment of multi-drug resistant fungi.

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## References

1. Brown, G.D.; Denning, D.W.; Gow, N.A.; Levitz, S.M.; Netea, M.G.; White, T.C. Hidden killers: Human fungal infections. *Sci. Transl. Med.* **2012**, *4*. [[CrossRef](#)] [[PubMed](#)]
2. White, T.C.; Findley, K.; Dawson, T.L., Jr.; Scheynius, A.; Boekhout, T.; Cuomo, C.A.; Xu, J.; Saunders, C.W. Fungi on the skin: Dermatophytes and *Malassezia*. *Cold Spring Harb. Perspect. Med.* **2014**, *4*. [[CrossRef](#)]
3. Bongomin, F.; Gago, S.; Oladele, R.O.; Denning, D.W. Global and multi-national prevalence of fungal diseases—estimate precision. *J. Fungi* **2017**, *3*, 57. [[CrossRef](#)] [[PubMed](#)]
4. Park, B.J.; Wannemuehler, K.A.; Marston, B.J.; Govender, N.; Pappas, P.G.; Chiller, T.M. Estimation of the current global burden of cryptococcal meningitis among persons living with HIV/AIDS. *AIDS* **2009**, *23*, 525–530. [[CrossRef](#)] [[PubMed](#)]
5. Drgona, L.; Khachatryan, A.; Stephens, J.; Charbonneau, C.; Kantecki, M.; Haider, S.; Barnes, R. Clinical and economic burden of invasive fungal diseases in Europe: Focus on pre-emptive and empirical treatment of *Aspergillus* and *Candida* species. *Eur. J. Clin. Microbiol. Infect. Dis.* **2014**, *33*, 7–21. [[CrossRef](#)] [[PubMed](#)]
6. Murray, P.R.; Rosenthal, K.S.; Pfaller, M.A. Section 6. Mycology. In *Medical Microbiology*, 7th ed.; Saunders: Philadelphia, PA, USA, 2013; pp. 605–711.
7. Reddy, K.R. Fungal infections (Mycoses): Dermatophytoses (Tinea, Ringworm). *J. Gandaki Med. Coll. Nepal* **2017**, *10*. [[CrossRef](#)]
8. Lortholary, O.; Desnos-Ollivier, M.; Sitbon, K.; Fontanet, A.; Bretagne, S.; Dromer, F. Recent exposure to caspofungin or fluconazole influences the epidemiology of candidemia: A prospective multicenter study involving 2,441 patients. *Antimicrob. Agents Chemother.* **2011**, *55*, 532–538. [[CrossRef](#)]

9. Alexander, B.D.; Johnson, M.D.; Pfeiffer, C.D.; Jiménez-Ortigosa, C.; Catania, J.; Booker, R.; Castanheira, M.; Messer, S.A.; Perlin, D.S.; Pfaller, M.A. Increasing echinocandin resistance in *Candida glabrata*: Clinical failure correlates with presence of FKS mutations and elevated minimum inhibitory concentrations. *Clin. Infect. Dis.* **2013**, *56*, 1724–1732. [CrossRef]
10. Arendrup, M.C.; Perlin, D.S. Echinocandin resistance: An emerging clinical problem? *Curr. Opin. Infect. Dis.* **2014**, *27*, 484–492. [CrossRef]
11. Van der Linden, J.W.; Arendrup, M.C.; Warris, A.; Lagrou, K.; Pelloux, H.; Hauser, P.M.; Chryssanthou, E.; Mellado, E.; Kidd, S.E.; Tortorano, A.M.; et al. Prospective multicenter international surveillance of azole resistance in *Aspergillus fumigatus*. *Emerg. Infect. Dis.* **2015**, *21*, 1041–1044. [CrossRef]
12. Pfaller, M.A.; Diekema, D.J. Rare and emerging opportunistic fungal pathogens: Concern for resistance beyond *Candida albicans* and *Aspergillus fumigatus*. *J. Clin. Microbiol.* **2004**, *42*, 4419–4431. [CrossRef] [PubMed]
13. Perlin, D.S.; Rautemaa-Richardson, R.; Alastruey-Izquierdo, A. The global problem of antifungal resistance: Prevalence, mechanisms, and management. *Lancet Infect. Dis.* **2017**, *17*, e383–e392. [CrossRef]
14. Krcmery, V., Jr.; Spanik, S.; Kunova, A.; Trupl, J. Breakthrough fungemia appearing during empiric therapy with amphotericin B. *Chemotherapy* **1997**, *43*, 367–370. [CrossRef] [PubMed]
15. Hull, C.M.; Bader, O.; Parker, J.E.; Weig, M.; Gross, U.; Warrilow, A.G.; Kelly, D.E.; Kelly, S.L. Two clinical isolates of *Candida glabrata* exhibiting reduced sensitivity to amphotericin B both harbor mutations in ERG2. *Antimicrob. Agents Chemother.* **2012**, *56*, 6417–6421. [CrossRef]
16. Woods, R.A.; Bard, M.; Jackson, I.E.; Drutz, D.J. Resistance to polyene antibiotics and correlated sterol changes in two isolates of *Candida tropicalis* from a patient with an amphotericin B-resistant funguria. *J. Infect. Dis.* **1974**, *129*, 53–58. [CrossRef]
17. Satoh, K.; Makimura, K.; Hasumi, Y.; Nishiyama, Y.; Uchida, K.; Yamaguchi, H. *Candida auris* sp. nov, a novel ascomycetous yeast isolated from the external ear canal of an inpatient in a Japanese hospital. *Microbiol. Immunol.* **2009**, *53*, 41–44. [CrossRef]
18. Tracking *Candida Auris*. Case Count Updated as of July 31; 2019; CDC. Available online: <https://www.cdc.gov/fungal/candida-auris/tracking-c-auris.html> (accessed on 9 September 2019).
19. Lockhart, S.R.; Etienne, K.A.; Vallabhaneni, S.; Farooqi, J.; Chowdhary, A.; Govender, N.P.; Colombo, A.L.; Calvo, B.; Cuomo, C.A.; Desjardins, C.A.; et al. Simultaneous emergence of multidrug-resistant *Candida auris* on 3 continents confirmed by whole-genome sequencing and epidemiological analyses. *Clin. Infect. Dis.* **2017**, *64*, 134–140. [CrossRef]
20. Friedman, D.Z.P.; Schwartz, I.S. Emerging fungal infections: New patients, new patterns, and new pathogens. *J. Fungi* **2019**, *5*, 67. [CrossRef]
21. Hata, D.J.; Humphries, R.; Lockhart, S.R.; College of American Pathologists Microbiology Committee. *Candida auris*: An emerging yeast pathogen posing distinct challenges for laboratory diagnostics, treatment, and infection prevention. *Arch. Pathol. Lab. Med.* **2019**. [CrossRef]
22. Clinical and Laboratory Standards Institute. *Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeast*, 3rd ed.; Approved Standard, CLSI document M27-A3, Clinical and Laboratory Standards Institute: Wayne, PA, USA, 2008.
23. Clinical and Laboratory Standards Institute. *Reference Method for Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi*, 2nd ed.; Approved Standard, CLSI document M38-A2, Clinical and Laboratory Standards Institute: Wayne, PA, USA, 2008.
24. Przybyłek, I.; Karpiński, T.M. Antibacterial properties of propolis. *Molecule* **2019**, *24*, 47. [CrossRef]
25. Ramasubramania Raja, R. Medicinally potential plants of Labiateae (Lamiaceae) family: An overview. *Res. J. Med. Plant.* **2012**, *6*, 203–213.
26. Carović-Stanko, K.; Petek, M.; Grdiša, M.; Pintar, J.; Bedeković, D.; Herak Ćustić, M.; Satovic, Z. Medicinal plants of the family Lamiaceae as functional foods—A review. *Czech J. Food Sci.* **2016**, *34*, 377–390. [CrossRef]
27. Radulović, N.S.; Blagojević, P.D.; Stojanović-Radić, Z.Z.; Stojanović, N.M. Antimicrobial plant metabolites: Structural diversity and mechanism of action. *Curr. Med. Chem.* **2013**, *20*, 932–952. [PubMed]
28. Vainstein, A.; Lewinsohn, E.; Pichersky, E.; Weiss, D. Floral fragrance. New inroads into an old commodity. *Plant Physiol.* **2001**, *127*, 1383–1389. [CrossRef]

29. De Groot, A.C.; Schmidt, E. Essential oils, Part III: Chemical composition. *Dermatitis* **2016**, *27*, 161–619. [[CrossRef](#)]
30. Piątkowska, E.; Rusiecka-Ziółkowska, J. Influence of essential oils on infectious agents. *Adv. Clin. Exp. Med.* **2016**, *25*, 989–995. [[CrossRef](#)]
31. Dhifi, W.; Bellili, S.; Jazi, S.; Bahloul, N.; Mnif, W. Essential oils' chemical characterization and investigation of some biological activities: A critical review. *Medicines* **2016**, *3*, 25. [[CrossRef](#)]
32. The Plant List. Available online: <http://www.theplantlist.org> (accessed on 12 September 2019).
33. PubChem. Available online: <https://pubchem.ncbi.nlm.nih.gov> (accessed on 23 September 2019).
34. Ngo Mback, M.N.; Agnani, H.; Nguimatsia, F.; Jazet Dongmo, P.M.; Houndou Fokou, J.B.; Bakarnga-Via, I.; Fekam Boyom, F.; Menut, C. Optimization of antifungal activity of *Aeollanthus heliotropioides* oliv essential oil and Time Kill Kinetic Assay. *J. Mycol. Med.* **2016**, *26*, 233–243. [[CrossRef](#)]
35. Ivanov, I.G.; Vrancheva, R.Z.; Petkova, N.T.; Tumbarski, Y.; Dincheva, I.N.; Badjakov, I.K. Phytochemical compounds of anise hyssop (*Agastache foeniculum*) and antibacterial, antioxidant, and acetylcholinesterase inhibitory properties of its essential oil. *J. Appl. Pharmac. Sci.* **2019**, *9*, 72–78.
36. Li, H.Q.; Liu, Q.Z.; Liu, Z.L.; Du, S.S.; Deng, Z.W. Chemical composition and nematocidal activity of essential oil of *Agastache rugosa* against *Meloidogyne incognita*. *Molecules* **2013**, *18*, 4170–4180. [[CrossRef](#)]
37. Fraternal, D.; Ricci, D. Essential oil composition and antifungal activity of aerial parts of *Ballota nigra* ssp *foetida* collected at flowering and fruiting times. *Nat. Prod. Commun.* **2014**, *9*, 1015–1018. [[CrossRef](#)] [[PubMed](#)]
38. Marinković, B.; Marin, P.D.; Knezević-Vukčević, J.; Soković, M.D.; Brkić, D. Activity of essential oils of three *Micromeria* species (Lamiaceae) against micromycetes and bacteria. *Phytother. Res.* **2002**, *16*, 336–339. [[CrossRef](#)] [[PubMed](#)]
39. Monforte, M.T.; Tzakou, O.; Nostro, A.; Zimbalatti, V.; Galati, E.M. Chemical composition and biological activities of *Calamintha officinalis* Moench essential oil. *J. Med. Food* **2011**, *14*, 297–303. [[CrossRef](#)] [[PubMed](#)]
40. Čavar, S.; Vidic, D.; Maksimović, M. Volatile constituents, phenolic compounds, and antioxidant activity of *Calamintha glandulosa* (Req.) Benth. *J. Sci. Food Agric.* **2013**, *93*, 1758–1764. [[CrossRef](#)]
41. Marongiu, B.; Piras, A.; Porcedda, S.; Falconieri, D.; Maxia, A.; Gonçalves, M.J.; Cavaleiro, C.; Salgueiro, L. Chemical composition and biological assays of essential oils of *Calamintha nepeta* (L.) Savi subsp. *nepeta* (Lamiaceae). *Nat. Prod. Res.* **2010**, *24*, 1734–1742. [[CrossRef](#)] [[PubMed](#)]
42. Božović, M.; Garzoli, S.; Sabatino, M.; Pepi, F.; Baldisserotto, A.; Andreotti, E.; Romagnoli, C.; Mai, A.; Manfredini, S.; Ragno, R. Essential oil extraction, chemical analysis and anti-*Candida* activity of *Calamintha nepeta* (L.) Savi subsp. *landulosa* (Req.) Ball—New approaches. *Molecules* **2017**, *22*, 203. [[CrossRef](#)]
43. Kumar, V.; Mathela, C.S.; Tewari, A.K.; Bisht, K.S. In vitro inhibition activity of essential oils from some Lamiaceae species against phytopathogenic fungi. *Pestic. Biochem. Physiol.* **2014**, *114*, 67–71. [[CrossRef](#)]
44. Stappen, I.; Wanner, J.; Tabanca, N.; Wedge, D.E.; Ali, A.; Kaul, V.K.; Lal, B.; Jaitak, V.; Gochev, V.K.; Schmidt, E.; et al. Chemical composition and biological activity of essential oils of *Dracocephalum heterophyllum* and *Hyssopus officinalis* from Western Himalaya. *Nat. Prod. Commun.* **2015**, *10*, 133–138. [[CrossRef](#)]
45. Ahmadi, F.; Sadeghi, S.; Modarresi, M.; Abiri, R.; Mikaeli, A. Chemical composition, in vitro anti-microbial, antifungal and antioxidant activities of the essential oil and methanolic extract of *Hymenocrater longiflorus* Benth., of Iran. *Food Chem. Toxicol.* **2010**, *48*, 1137–1144. [[CrossRef](#)]
46. De Oliveira, C.M.A.; Silva, M.R.R.; Kato, L.; da Silva, C.C.; Ferreira, H.D.; Souza, L.K.H. Chemical composition and antifungal activity of the essential oil of *Hyptis ovalifolia* Benth. (Lamiaceae). *J. Braz. Chem. Soc.* **2004**, *15*, 756–759. [[CrossRef](#)]
47. Džamić, A.M.; Soković, M.D.; Novaković, M.; Jadranin, M.; Ristić, M.S.; Tešević, V.; Marin, P.D. Composition, antifungal and antioxidant properties of *Hyssopus officinalis* L. subsp. *pilifer* (Pant.) Murb. essential oil and deodorized extracts. *Ind. Crops Prod.* **2013**, *51*, 401–407.
48. Hristova, Y.; Wanner, J.; Jirovetz, L.; Stappen, I.; Iliev, I.; Gochev, V. Chemical composition and antifungal activity of essential oil of *Hyssopus officinalis* L. from Bulgaria against clinical isolates of *Candida* species. *Biotechnol. Biotechnol. Equip.* **2015**, *29*, 592–601. [[CrossRef](#)]
49. Adam, K.; Sivropoulou, A.; Kokkini, S.; Lanaras, T.; Arsenakis, M. Antifungal activities of *Origanum vulgare* subsp. *hirtum*, *Mentha spicata*, *Lavandula angustifolia*, and *Salvia fruticosa* essential oils against human pathogenic fungi. *J. Agric. Food Chem.* **1998**, *46*, 1739–1745.

50. D'Auria, F.D.; Tecca, M.; Strippoli, V.; Salvatore, G.; Battinelli, L.; Mazzanti, G. Antifungal activity of *Lavandula angustifolia* essential oil against *Candida albicans* yeast and mycelial form. *Med. Mycol.* **2005**, *43*, 391–396. [[CrossRef](#)] [[PubMed](#)]
51. Houry, M.; Stien, D.; Eparvier, V.; Ouaini, N.; El Beyrouthy, M. Report on the medicinal use of eleven Lamiaceae species in Lebanon and rationalization of their antimicrobial potential by examination of the chemical composition and antimicrobial activity of their essential oils. *Evid. Based Compl. Altern. Med.* **2016**, *2016*. [[CrossRef](#)]
52. Zuzarte, M.; Vale-Silva, L.; Gonçalves, M.J.; Cavaleiro, C.; Vaz, S.; Canhoto, J.; Pinto, E.; Salgueiro, L. Antifungal activity of phenolic-rich *Lavandula multifida* L. essential oil. *Eur. J. Clin. Microbiol. Infect. Dis.* **2012**, *31*, 1359–1366. [[CrossRef](#)]
53. Zuzarte, M.; Gonçalves, M.J.; Cavaleiro, C.; Dinis, A.M.; Canhoto, J.M.; Salgueiro, L.R. Chemical composition and antifungal activity of the essential oils of *Lavandula pedunculata* (Miller) Cav. *Chem. Biodivers.* **2009**, *6*, 1283–1292. [[CrossRef](#)]
54. Zuzarte, M.; Gonçalves, M.J.; Cruz, M.T.; Cavaleiro, C.; Canhoto, J.; Vaz, S.; Pinto, E.; Salgueiro, L. *Lavandula luisieri* essential oil as a source of antifungal drugs. *Food Chem.* **2012**, *135*, 1505–1510. [[CrossRef](#)]
55. Zuzarte, M.; Gonçalves, M.J.; Cavaleiro, C.; Canhoto, J.; Vale-Silva, L.; Silva, M.J.; Pinto, E.; Salgueiro, L. Chemical composition and antifungal activity of the essential oils of *Lavandula viridis* L'Her. *J. Med. Microbiol.* **2011**, *60*, 612–618. [[CrossRef](#)]
56. Ramírez, J.; Gilardoni, G.; Jácome, M.; Montesinos, J.; Rodolfi, M.; Guglielminetti, M.L.; Cagliero, C.; Bicchi, C.; Vidari, G. Chemical composition, enantiomeric analysis, AEDA sensorial evaluation and antifungal activity of the essential oil from the Ecuadorian plant *Lepechinia mutica* Benth (Lamiaceae). *Chem. Biodivers.* **2017**, *14*, e1700292. [[CrossRef](#)]
57. Ramírez, J.; Gilardoni, G.; Ramón, E.; Tosi, S.; Picco, A.M.; Bicchi, C.; Vidari, G. Phytochemical study of the Ecuadorian species *Lepechinia mutica* (Benth.) Epling and high antifungal activity of carnosol against *Pyricularia oryzae*. *Pharmaceuticals* **2018**, *11*, 33. [[CrossRef](#)] [[PubMed](#)]
58. Zarai, Z.; Kadri, A.; Ben Chobba, I.; Ben Mansour, R.; Bekir, A.; Mejdoub, H.; Gharsallah, N. The in-vitro evaluation of antibacterial, antifungal and cytotoxic properties of *Marrubium vulgare* L. essential oil grown in Tunisia. *Lipids Health Dis.* **2011**, *10*, 161. [[CrossRef](#)] [[PubMed](#)]
59. Mimica-Dukic, N.; Bozin, B.; Sokovic, M.; Simin, N. Antimicrobial and antioxidant activities of *Melissa officinalis* L. (Lamiaceae) essential oil. *J. Agric. Food Chem.* **2004**, *52*, 2485–2489. [[CrossRef](#)] [[PubMed](#)]
60. Gonçalves, M.J.; Vicente, A.M.; Cavaleiro, C.; Salgueiro, L. Composition and antifungal activity of the essential oil of *Mentha cervina* from Portugal. *Nat. Prod. Res.* **2007**, *21*, 867–871. [[CrossRef](#)] [[PubMed](#)]
61. Soković, M.D.; Vukojević, J.; Marin, P.D.; Brkić, D.D.; Vajs, V.; van Griensven, L.J. Chemical composition of essential oils of *Thymus* and *Mentha* species and their antifungal activities. *Molecules* **2009**, *14*, 238–249. [[CrossRef](#)] [[PubMed](#)]
62. Camiletti, B.X.; Asensio, C.M.; Pecci Mde, L.; Lucini, E.I. Natural control of corn postharvest fungi *Aspergillus flavus* and *Penicillium* sp. using essential oils from plants grown in Argentina. *J. Food Sci.* **2014**, *79*, M2499–M2506. [[CrossRef](#)]
63. Samber, N.; Khan, A.; Varma, A.; Manzoor, N. Synergistic anti-candidal activity and mode of action of *Mentha piperita* essential oil and its major components. *Pharm. Biol.* **2015**, *53*, 1496–1504. [[CrossRef](#)]
64. Hossain, F.; Follett, P.; Dang Vu, K.; Harich, M.; Salmieri, S.; Lacroix, M. Evidence for synergistic activity of plant-derived essential oils against fungal pathogens of food. *Food Microbiol.* **2016**, *53*, 24–30. [[CrossRef](#)]
65. Mahboubi, M.; Haghi, G. Antimicrobial activity and chemical composition of *Mentha pulegium* L. essential oil. *J. Ethnopharmacol.* **2008**, *119*, 325–327. [[CrossRef](#)]
66. Abdelli, M.; Moghrani, H.; Aboun, A.; Maachi, R. Algerian *Mentha pulegium* L. leaves essential oil: Chemical composition, antimicrobial, insecticidal and antioxidant activities. *Ind. Crops Prod.* **2016**, *94*, 197–205. [[CrossRef](#)]
67. Chessa, M.; Sias, A.; Piana, A.; Mangano, G.S.; Petretto, G.L.; Masia, M.D.; Tirillini, B.; Pintore, G. Chemical composition and antibacterial activity of the essential oil from *Mentha requienii* Benth. *Nat. Prod. Res.* **2013**, *27*, 93–99. [[CrossRef](#)] [[PubMed](#)]
68. Nardoni, S.; Giovanelli, S.; Pistelli, L.; Mugnaini, L.; Profili, G.; Pisseri, F.; Mancianti, F. In vitro activity of twenty commercially available, plant-derived essential oils against selected dermatophyte species. *Nat. Prod. Commun.* **2015**, *10*, 1473–1478. [[CrossRef](#)] [[PubMed](#)]



69. Oumzil, H.; Ghouami, S.; Rhajaoui, M.; Ildrissi, A.; Fkih-Tetouani, S.; Faid, M.; Benjouad, A. Antibacterial and antifungal activity of essential oils of *Mentha suaveolens*. *Phytother. Res.* **2002**, *16*, 727–731. [[CrossRef](#)] [[PubMed](#)]
70. El-Kashoury, S.A.; El-Askary, H.I.; Kandil, Z.A.; Salem, M.A. Chemical composition of the essential oil and botanical study of the flowers of *Mentha suaveolens*. *Pharm. Biol.* **2014**, *52*, 688–697. [[CrossRef](#)] [[PubMed](#)]
71. Garzoli, S.; Pirolli, A.; Vavala, E.; Di Sotto, A.; Sartorelli, G.; Božović, M.; Angiolella, L.; Mazzanti, G.; Pepi, F.; Ragno, R. Multidisciplinary approach to determine the optimal time and period for extracting the essential oil from *Mentha suaveolens* Ehrh. *Molecules* **2015**, *20*, 9640–9655. [[CrossRef](#)] [[PubMed](#)]
72. Casiglia, S.; Jemia, M.B.; Riccobono, L.; Bruno, M.; Scandolera, E.; Senatore, F. Chemical composition of the essential oil of *Moluccella spinosa* L. (Lamiaceae) collected wild in Sicily and its activity on microorganisms affecting historical textiles. *Nat. Prod. Res.* **2015**, *29*, 1201–1206. [[CrossRef](#)]
73. Sacchetti, G.; Medici, A.; Maietti, S.; Radice, M.; Muzzoli, M.; Manfredini, S.; Braccioli, E.; Bruni, R. Composition and functional properties of the essential oil of amazonian basil, *Ocimum micranthum* Willd., Labiatae in comparison with commercial essential oils. *J. Agric. Food Chem.* **2004**, *52*, 3486–3491. [[CrossRef](#)]
74. Bozin, B.; Mimica-Dukic, N.; Simin, N.; Anackov, G. Characterization of the volatile composition of essential oils of some Lamiaceae spices and the antimicrobial and antioxidant activities of the entire oils. *J. Agric. Food Chem.* **2006**, *54*, 1822–1828. [[CrossRef](#)]
75. Al-Maskri, A.Y.; Hanif, M.A.; Al-Maskari, M.Y.; Abraham, A.S.; Al-sabahi, J.N.; Al-Mantheri, O. Essential oil from *Ocimum basilicum* (Omani Basil): A desert crop. *Nat. Prod. Commun.* **2011**, *6*, 1487–1490.
76. Avetisyan, A.; Markosian, A.; Petrosyan, M.; Sahakyan, N.; Babayan, A.; Aloyan, S.; Trchounian, A. Chemical composition and some biological activities of the essential oils from basil *Ocimum* different cultivars. *BMC Complement Altern. Med.* **2017**, *17*, 60. [[CrossRef](#)]
77. Ali, N.A.A.; Chhetri, B.K.; Dosoky, N.S.; Shari, K.; Al-Fahad, A.J.A.; Wessjohann, L.; Setzer, W.N. Antimicrobial, antioxidant, and cytotoxic activities of *Ocimum forskolei* and *Teucrium yemense* (Lamiaceae) essential oils. *Medicines* **2017**, *4*, 17. [[CrossRef](#)] [[PubMed](#)]
78. Dubey, N.K.; Tiwari, T.N.; Mandin, D.; Andriamboavonjy, H.; Chaumont, J.P. Antifungal properties of *Ocimum gratissimum* essential oil (ethyl cinnamate chemotype). *Fitoterapia* **2000**, *71*, 567–569. [[CrossRef](#)]
79. Lemos Jde, A.; Passos, X.S.; Fernandes Ode, F.; Paula, J.R.; Ferri, P.H.; Souza, L.K.; Lemos Ade, A.; Silva Mdo, R. Antifungal activity from *Ocimum gratissimum* L. towards *Cryptococcus neoformans*. *Mem. Inst. Oswaldo Cruz* **2005**, *100*, 55–58. [[CrossRef](#)] [[PubMed](#)]
80. Mohr, F.B.; Lermen, C.; Gazim, Z.C.; Gonçalves, J.E.; Alberton, O. Antifungal activity, yield, and composition of *Ocimum gratissimum* essential oil. *Genet. Mol. Res.* **2017**, *16*. [[CrossRef](#)]
81. Amber, K.; Aijaz, A.; Immaculata, X.; Luqman, K.A.; Nikhat, M. Anticandidal effect of *Ocimum sanctum* essential oil and its synergy with fluconazole and ketoconazole. *Phytomedicine* **2010**, *17*, 921–925. [[CrossRef](#)]
82. Khan, A.; Ahmad, A.; Akhtar, F.; Yousuf, S.; Xess, I.; Khan, L.A.; Manzoor, N. *Ocimum sanctum* essential oil and its active principles exert their antifungal activity by disrupting ergosterol biosynthesis and membrane integrity. *Res. Microbiol.* **2010**, *161*, 816–823. [[CrossRef](#)]
83. Kumar, A.; Shukla, R.; Singh, P.; Dubey, N.K. Chemical composition, antifungal and antiaflatoxic activities of *Ocimum sanctum* L. essential oil and its safety assessment as plant based antimicrobial. *Food Chem. Toxicol.* **2010**, *48*, 539–543. [[CrossRef](#)]
84. Santamarina, M.P.; Roselló, J.; Sempere, F.; Giménez, S.; Blázquez, M.A. Commercial *Origanum compactum* Benth. and *Cinnamomum zeylanicum* Blume essential oils against natural mycoflora in Valencia rice. *Nat. Prod. Res.* **2015**, *29*, 2215–2258. [[CrossRef](#)]
85. Homa, M.; Fekete, I.P.; Böszörményi, A.; Singh, Y.R.; Selvam, K.P.; Shobana, C.S.; Manikandan, P.; Kredics, L.; Vágvolgyi, C.; Galgóczy, L. Antifungal effect of essential oils against *Fusarium keratitis* isolates. *Planta Med.* **2015**, *81*, 1277–1284. [[CrossRef](#)]
86. Waller, S.B.; Madrid, I.M.; Ferraz, V.; Picoli, T.; Cleff, M.B.; de Faria, R.O.; Meireles, M.C.; de Mello, J.R. Cytotoxicity and anti-*Sporothrix brasiliensis* activity of the *Origanum majorana* Linn. oil. *Braz. J. Microbiol.* **2016**, *47*, 896–901. [[CrossRef](#)]
87. Nikkhah, M.; Hashemi, M.; Habibi Najafi, M.B.; Farhoosh, R. Synergistic effects of some essential oils against fungal spoilage on pear fruit. *Int. J. Food Microbiol.* **2017**, *257*, 285–294. [[CrossRef](#)] [[PubMed](#)]



88. Rosato, A.; Vitali, C.; Piarulli, M.; Mazzotta, M.; Argentieri, M.P.; Mallamaci, R. In vitro synergic efficacy of the combination of Nystatin with the essential oils of *Origanum vulgare* and *Pelargonium graveolens* against some *Candida* species. *Phytomedicine* **2009**, *16*, 972–975. [[CrossRef](#)] [[PubMed](#)]
89. Vale-Silva, L.; Silva, M.J.; Oliveira, D.; Gonçalves, M.J.; Cavaleiro, C.; Salgueiro, L.; Pinto, E. Correlation of the chemical composition of essential oils from *Origanum vulgare* subsp. *virens* with their in vitro activity against pathogenic yeasts and filamentous fungi. *J. Med. Microbiol.* **2012**, *61*, 252–260. [[PubMed](#)]
90. Jeršek, B.; Poklar Ulrih, N.; Skrt, M.; Gavarić, N.; Božin, B.; Smole Možina, S. Effects of selected essential oils on the growth and production of ochratoxin A by *Penicillium verrucosum*. *Arhiv Higijenu i Toksikologiju* **2014**, *65*, 199–208. [[CrossRef](#)]
91. Elansary, H.O.; Abdelgaleil, S.A.M.; Mahmoud, E.A.; Yessoufou, K.; Elhindi, K.; El-Hendawy, S. Effective antioxidant, antimicrobial and anticancer activities of essential oils of horticultural aromatic crops in northern Egypt. *BMC Complement Altern. Med.* **2018**, *18*, 214. [[CrossRef](#)]
92. Kocevski, D.; Du, M.; Kan, J.; Jing, C.; Lačanin, I.; Pavlović, H. Antifungal effect of *Allium tuberosum*, *Cinnamomum cassia*, and *Pogostemon cablin* essential oils and their components against population of *Aspergillus* species. *J. Food Sci.* **2013**, *78*, M731–M737. [[CrossRef](#)]
93. Murugan, R.; Mallavarapu, G.R.; Padmashree, K.V.; Rao, R.R.; Livingstone, C. Volatile oil composition of *Pogostemon heyneanus* and comparison of its composition with patchouli oil. *Nat. Prod. Commun.* **2010**, *5*, 1961–1964. [[CrossRef](#)]
94. Zhang, H.Y.; Gao, Y.; Lai, P.X. Chemical composition, antioxidant, antimicrobial and cytotoxic activities of essential oil from *Premna microphylla* Turczaninow. *Molecules* **2017**, *22*, 381. [[CrossRef](#)]
95. Angioni, A.; Barra, A.; Cereti, E.; Barile, D.; Coisson, J.D.; Arlorio, M.; Dessi, S.; Coroneo, V.; Cabras, P. Chemical composition, plant genetic differences, antimicrobial and antifungal activity investigation of the essential oil of *Rosmarinus officinalis* L. *J. Agric. Food Chem.* **2004**, *52*, 3530–3535. [[CrossRef](#)]
96. Božin, B.; Mimica-Dukic, N.; Samojlik, I.; Jovin, E. Antimicrobial and antioxidant properties of rosemary and sage (*Rosmarinus officinalis* L. and *Salvia officinalis* L., Lamiaceae) essential oils. *J. Agric. Food Chem.* **2007**, *55*, 7879–7885. [[CrossRef](#)]
97. Ozcan, M.M.; Chalchat, J.C. Chemical composition and antifungal activity of rosemary (*Rosmarinus officinalis* L.) oil from Turkey. *Int. J. Food Sci. Nutr.* **2008**, *59*, 691–698. [[CrossRef](#)] [[PubMed](#)]
98. Jiang, Y.; Wu, N.; Fu, Y.J.; Wang, W.; Luo, M.; Zhao, C.J.; Zu, Y.G.; Liu, X.L. Chemical composition and antimicrobial activity of the essential oil of Rosemary. *Environ. Toxicol. Pharmacol.* **2011**, *32*, 63–68. [[CrossRef](#)] [[PubMed](#)]
99. Pitarokili, D.; Tzakou, O.; Loukis, A.; Harvala, C. Volatile metabolites from *Salvia fruticosa* as antifungal agents in soilborne pathogens. *J. Agric. Food Chem.* **2003**, *51*, 3294–3301. [[CrossRef](#)] [[PubMed](#)]
100. Zomorodian, K.; Moein, M.; Pakshir, K.; Karami, F.; Sabahi, Z. Chemical composition and antimicrobial activities of the essential oil from *Salvia mirzayanii* leaves. *J. Evid. Based Complementary Altern. Med.* **2017**, *22*, 770–776. [[CrossRef](#)]
101. Abu-Darwish, M.S.; Cabral, C.; Ferreira, I.V.; Gonçalves, M.J.; Cavaleiro, C.; Cruz, M.T.; Al-bdour, T.H.; Salgueiro, L. Essential oil of common sage (*Salvia officinalis* L.) from Jordan: Assessment of safety in mammalian cells and its antifungal and anti-inflammatory potential. *BioMed Res. Int.* **2013**, *2013*. [[CrossRef](#)]
102. Yuce, E.; Yildirim, N.; Yildirim, N.C.; Paksoy, M.Y.; Bagci, E. Essential oil composition, antioxidant and antifungal activities of *Salvia sclarea* L. from Munzur Valley in Tunceli, Turkey. *Cell. Mol. Biol.* **2014**, *60*, 1–5.
103. Güllüce, M.; Sökmen, M.; Daferera, D.; Ađar, G.; Ozkan, H.; Kartal, N.; Polissiou, M.; Sökmen, A.; Sahin, F. In vitro antibacterial, antifungal, and antioxidant activities of the essential oil and methanol extracts of herbal parts and callus cultures of *Satureja hortensis* L. *J. Agric. Food Chem.* **2003**, *51*, 3958–3965. [[CrossRef](#)]
104. Omidbeygi, M.; Barzegar, M.; Hamidi, Z.; Naghdibadi, H. Antifungal activity of thyme, summer savory and clove essential oils against *Aspergillus flavus* in liquid medium and tomato paste. *Food Control* **2007**, *18*, 1518–1523. [[CrossRef](#)]
105. Giweli, A.; Džamić, A.M.; Soković, M.; Ristić, M.S.; Marin, P.D. Antimicrobial and antioxidant activities of essential oils of *Satureja thymbra* growing wild in Libya. *Molecules* **2012**, *17*, 4836–4850. [[CrossRef](#)]
106. Serbetçi, T.; Demirci, B.; Güzel, C.B.; Kültür, S.; Ergüven, M.; Başer, K.H. Essential oil composition, antimicrobial and cytotoxic activities of two endemic *Stachys cretica* subspecies (Lamiaceae) from Turkey. *Nat. Prod. Commun.* **2010**, *5*, 1369–1374. [[CrossRef](#)]

107. Lazarević, J.S.; Đorđević, A.S.; Kitić, D.V.; Zlatković, B.K.; Stojanović, G.S. Chemical composition and antimicrobial activity of the essential oil of *Stachys officinalis* (L.) Trevis. (Lamiaceae). *Chem. Biodivers.* **2013**, *10*, 1335–1349. [[CrossRef](#)] [[PubMed](#)]
108. Mohammadi, A.; Nazari, H.; Imani, S.; Amrollahi, H. Antifungal activities and chemical composition of some medicinal plants. *J. Mycol. Med.* **2014**, *24*, e1–e8. [[CrossRef](#)] [[PubMed](#)]
109. Salah, K.B.; Mahjoub, M.A.; Chaumont, J.P.; Michel, L.; Millet-Clerc, J.; Chraeif, I.; Ammar, S.; Mighri, Z.; Aouni, M. Chemical composition and in vitro antifungal and antioxidant activity of the essential oil and methanolic extract of *Teucrium sauvagei* Le Houerou. *Nat. Prod. Res.* **2006**, *20*, 1089–1097. [[CrossRef](#)] [[PubMed](#)]
110. Goren, A.C.; Bilsel, G.; Bilsel, M.; Demir, H.; Kocabaş, E.E. Analysis of essential oil of *Coridothymus capitatus* (L.) and its antibacterial and antifungal activity. *Zeitschrift für Naturforschung C* **2003**, *58*, 687–690. [[CrossRef](#)]
111. Salgueiro, L.R.; Pinto, E.; Gonçalves, M.J.; Pina-Vaz, C.; Cavaleiro, C.; Rodrigues, A.G.; Palmeira, A.; Tavares, C.; Costa-de-Oliveira, S.; Martinez-de-Oliveira, J. Chemical composition and antifungal activity of the essential oil of *Thymbra capitata*. *Planta Med.* **2004**, *70*, 572–575. [[CrossRef](#)]
112. Palmeira-de-Oliveira, A.; Gaspar, C.; Palmeira-de-Oliveira, R.; Silva-Dias, A.; Salgueiro, L.; Cavaleiro, C.; Pina-Vaz, C.; Martinez-de-Oliveira, J.; Queiroz, J.A.; Rodrigues, A.G. The anti-*Candida* activity of *Thymbra capitata* essential oil: Effect upon pre-formed biofilm. *J. Ethnopharmacol.* **2012**, *140*, 379–383. [[CrossRef](#)]
113. Kiliç, T. Analysis of essential oil composition of *Thymbra spicata* var. *spicata*: Antifungal, antibacterial and antimycobacterial activities. *Z. Naturforsch. C* **2006**, *61*, 324–328.
114. Unlü, M.; Vardar-Unlü, G.; Vural, N.; Dönmez, E.; Ozbaş, Z.Y. Chemical composition, antibacterial and antifungal activity of the essential oil of *Thymbra spicata* L. from Turkey. *Nat. Prod. Res.* **2009**, *23*, 572–579. [[CrossRef](#)]
115. Jaradat, N.; Adwan, L.; Kaibni, S.; Shraim, N.; Zaid, A.N. Chemical composition, anthelmintic, antibacterial and antioxidant effects of *Thymus bovei* essential oil. *BMC Complement Altern. Med.* **2016**, *16*, 418. [[CrossRef](#)]
116. Pina-Vaz, C.; Gonçalves Rodrigues, A.; Pinto, E.; Costa-de-Oliveira, S.; Tavares, C.; Salgueiro, L.; Cavaleiro, C.; Gonçalves, M.J.; Martinez-de-Oliveira, J. Antifungal activity of *Thymus* oils and their major compounds. *J. Eur. Acad. Dermatol. Venereol.* **2004**, *18*, 73–78. [[CrossRef](#)]
117. Alizadeh, A.; Zamani, E.; Sharaifi, R.; Javan-Nikkhah, M.; Nazari, S. Antifungal activity of some essential oils against toxigenic *Aspergillus* species. *Commun. Agric. Appl. Biol. Sci.* **2010**, *75*, 761–767. [[PubMed](#)]
118. Alizadeh, A.; Sharaifi, R.; Javan-Nikkhah, M.; Sedaghat, N. Survey of *Thymus migricus* essential oil on aflatoxin inhibition in *Aspergillus flavus*. *Commun. Agric. Appl. Biol. Sci.* **2010**, *75*, 769–776. [[PubMed](#)]
119. Pinto, E.; Pina-Vaz, C.; Salgueiro, L.; Gonçalves, M.J.; Costa-de-Oliveira, S.; Cavaleiro, C.; Palmeira, A.; Rodrigues, A.; Martinez-de-Oliveira, J. Antifungal activity of the essential oil of *Thymus pulegioides* on *Candida*, *Aspergillus* and dermatophyte species. *J. Med. Microbiol.* **2006**, *55*, 1367–1373. [[CrossRef](#)] [[PubMed](#)]
120. Pagiotti, R.; Angelini, P.; Rubini, A.; Tirillini, B.; Granetti, B.; Venanzoni, R. Identification and characterisation of human pathogenic filamentous fungi and susceptibility to *Thymus schimperii* essential oil. *Mycoses* **2011**, *54*, e364–e376. [[CrossRef](#)]
121. Couladis, M.; Tzakou, O.; Kujundzic, S.; Sokovic, M.; Mimica-Dukic, N. Chemical analysis and antifungal activity of *Thymus striatus*. *Phytother. Res.* **2004**, *18*, 40–42. [[CrossRef](#)]
122. Segvić Klarić, M.; Kosalec, I.; Mastelić, J.; Piecková, E.; Pepeljnak, S. Antifungal activity of thyme (*Thymus vulgaris* L.) essential oil and thymol against moulds from damp dwellings. *Lett. Appl. Microbiol.* **2007**, *44*, 36–42. [[CrossRef](#)]
123. De Lira Mota, K.S.; de Oliveira Pereira, F.; de Oliveira, W.A.; Lima, I.O.; de Oliveira Lima, E. Antifungal activity of *Thymus vulgaris* L. essential oil and its constituent phytochemicals against *Rhizopus oryzae*: Interaction with ergosterol. *Molecules* **2012**, *17*, 14418–14433. [[CrossRef](#)]
124. Khan, M.S.; Ahmad, I.; Cameotra, S.S. *Carum copticum* and *Thymus vulgaris* oils inhibit virulence in *Trichophyton rubrum* and *Aspergillus* spp. *Braz. J. Microbiol.* **2014**, *45*, 523–531. [[CrossRef](#)]
125. Kohiyama, C.Y.; Yamamoto Ribeiro, M.M.; Mossini, S.A.; Bando, E.; Bomfim Nda, S.; Nerilo, S.B.; Rocha, G.H.; Grespan, R.; Mikcha, J.M.; Machinski, M., Jr. Antifungal properties and inhibitory effects upon aflatoxin production of *Thymus vulgaris* L. by *Aspergillus flavus* Link. *Food Chem.* **2015**, *173*, 1006–1010. [[CrossRef](#)]
126. Divband, K.; Shokri, H.; Khosravi, A.R. Down-regulatory effect of *Thymus vulgaris* L. on growth and Tri4 gene expression in *Fusarium oxysporum* strains. *Microb. Pathog.* **2017**, *104*, 1–5. [[CrossRef](#)]

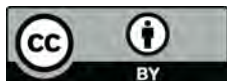
127. Lagha, R.; Ben Abdallah, F.; Al-Sarhan, B.O.; Al-Sodany, Y. Antibacterial and biofilm inhibitory activity of medicinal plant essential oils against *Escherichia coli* isolated from UTI patients. *Molecules* **2019**, *24*, 1161. [[CrossRef](#)] [[PubMed](#)]
128. Marongiu, B.; Piras, A.; Porcedda, S.; Falconieri, D.; Goncalves, M.J.; Salgueiro, L.; Maxia, A.; Lai, R. Extraction, separation and isolation of volatiles from *Vitex agnus-castus* L. (Verbenaceae) wild species of Sardinia, Italy, by supercritical CO<sub>2</sub>. *Nat. Prod. Res.* **2010**, *24*, 569–579. [[CrossRef](#)] [[PubMed](#)]
129. Asdadi, A.; Hamdouch, A.; Oukacha, A.; Moutaj, R.; Gharby, S.; Harhar, H.; El Hadek, M.; Chebli, B.; Idrissi Hassani, L.M. Study on chemical analysis, antioxidant and in vitro antifungal activities of essential oil from wild *Vitex agnus-castus* L. seeds growing in area of Argan Tree of Morocco against clinical strains of *Candida* responsible for nosocomial infections. *J. Mycol. Med.* **2015**, *25*, e118–e127. [[CrossRef](#)] [[PubMed](#)]
130. Mahboubi, M.; Heidary Tabar, R.; Mahdizadeh, E. The anti-dermatophyte activity of *Zataria multiflora* essential oils. *J. Mycol. Med.* **2017**, *27*, 232–237. [[CrossRef](#)] [[PubMed](#)]
131. Shahbazi, Y. Chemical compositions, antioxidant and antimicrobial properties of *Ziziphora clinopodioides* Lam. essential oils collected from different parts of Iran. *J. Food Sci. Technol.* **2017**, *54*, 3491–3503. [[CrossRef](#)] [[PubMed](#)]
132. Mohammadifard, F.; Alimohammadi, S. Chemical composition and role of opioidergic system in antinociceptive effect of *Ziziphora clinopodioides* essential oil. *Basic Clin. Neurosci.* **2018**, *9*, 357–366. [[CrossRef](#)] [[PubMed](#)]
133. Abu-Darwish, M.S.; Cabral, C.; Gonçalves, M.J.; Cavaleiro, C.; Cruz, M.T.; Paoli, M.; Tomi, F.; Efferth, T.; Salgueiro, L. *Ziziphora tenuior* L. essential oil from Dana Biosphere Reserve (Southern Jordan); Chemical characterization and assessment of biological activities. *J. Ethnopharmacol.* **2016**, *194*, 963–970. [[CrossRef](#)]
134. Yang, D.; Michel, L.; Chaumont, J.P.; Millet-Clerc, J. Use of caryophyllene oxide as an antifungal agent in an in vitro experimental model of onychomycosis. *Mycopathologia* **1999**, *148*, 79–82. [[CrossRef](#)]
135. Dahham, S.S.; Tabana, Y.M.; Iqbal, M.A.; Ahamed, M.B.; Ezzat, M.O.; Majid, A.S.; Majid, A.M. The anticancer, antioxidant and antimicrobial properties of the sesquiterpene  $\beta$ -caryophyllene from the essential oil of *Aquilaria crassna*. *Molecules* **2015**, *20*, 11808–11829. [[CrossRef](#)]
136. Selestino Neta, M.C.; Vittorazzi, C.; Guimarães, A.C.; Martins, J.D.; Fronza, M.; Endringer, D.C.; Scherer, R. Effects of  $\beta$ -caryophyllene and *Murraya paniculata* essential oil in the murine hepatoma cells and in the bacteria and fungi 24-h time-kill curve studies. *Pharm. Biol.* **2017**, *55*, 190–197. [[CrossRef](#)]
137. Bona, E.; Cantamessa, S.; Pavan, M.; Novello, G.; Massa, N.; Rocchetti, A.; Berta, G.; Gamalero, E. Sensitivity of *Candida albicans* to essential oils: Are they an alternative to antifungal agents? *J. Appl. Microbiol.* **2016**, *121*, 1530–1545. [[CrossRef](#)]
138. Kordali, S.; Cakir, A.; Ozer, H.; Cakmakci, R.; Kesdek, M.; Mete, E. Antifungal, phytotoxic and insecticidal properties of essential oil isolated from Turkish *Origanum acutidens* and its three components, carvacrol, thymol and p-cymene. *Bioresour. Technol.* **2008**, *99*, 8788–8795. [[CrossRef](#)]
139. Marei, G.I.K.; Abdel Rasoul, M.A.; Abdelgaleil, S.A.M. Comparative antifungal activities and biochemical effects of monoterpenes on plant pathogenic fungi. *Pesticide Biochem. Physiol.* **2012**, *103*, 56–61. [[CrossRef](#)]
140. Abbaszadeh, S.; Sharifzadeh, A.; Shokri, H.; Khosravi, A.R.; Abbaszadeh, A. Antifungal efficacy of thymol, carvacrol, eugenol and menthol as alternative agents to control the growth of food-relevant fungi. *J. Mycol. Med.* **2014**, *24*, e51–e56. [[CrossRef](#)]
141. Rivera-Yañez, C.R.; Terrazas, L.I.; Jimenez-Estrada, M.; Campos, J.E.; Flores-Ortiz, C.M.; Hernandez, L.B.; Cruz-Sanchez, T.; Garrido-Fariña, G.I.; Rodriguez-Monroy, M.A.; Canales-Martinez, M.M. Anti-*Candida* activity of *Bursera morelensis* Ramirez essential oil and two compounds,  $\alpha$ -pinene and  $\gamma$ -terpinene—an in vitro study. *Molecules* **2017**, *22*, 95. [[CrossRef](#)]
142. de Oliveira Lima, M.I.; Araújo de Medeiros, A.C.; Souza Silva, K.V.; Cardoso, G.N.; de Oliveira Lima, E.; de Oliveira Pereira, F. Investigation of the antifungal potential of linalool against clinical isolates of fluconazole resistant *Trichophyton rubrum*. *J. Mycol. Med.* **2017**, *27*, 195–202. [[CrossRef](#)] [[PubMed](#)]
143. de Macêdo Andrade, A.C.; Rosalen, P.L.; Freires, I.A.; Scotti, L.; Scotti, M.T.; Aquino, S.G.; de Castro, R.D. Antifungal activity, mode of action, docking prediction and anti-biofilm effects of (+)- $\beta$ -pinene enantiomers against *Candida* spp. *Curr. Top. Med. Chem.* **2018**, *18*, 2481–2490. [[CrossRef](#)] [[PubMed](#)]
144. Wang, K.; Jiang, S.; Pu, T.; Fan, L.; Su, F.; Ye, M. Antifungal activity of phenolic monoterpenes and structure-related compounds against plant pathogenic fungi. *Nat. Prod. Res.* **2019**, *33*, 1423–1430. [[CrossRef](#)]

145. Shi, Y.; Si, H.; Wang, P.; Chen, S.; Shang, S.; Song, Z.; Wang, Z.; Liao, S. Derivatization of natural compound  $\beta$ -pinene enhances its in vitro antifungal activity against plant pathogens. *Molecules* **2019**, *24*, 3144. [[CrossRef](#)]
146. Wojtunik-Kulesza, K.A.; Kasprzak, K.; Oniszczyk, T.; Oniszczyk, A. Natural monoterpenes: Much more than only a scent. *Chem. Biodiv.* **2019**, *16*, e19004. [[CrossRef](#)]
147. Burt, S. Essential oils: Their antibacterial properties and potential applications in foods—A review. *Int. J. Food Microbiol.* **2004**, *94*, 223–253. [[CrossRef](#)] [[PubMed](#)]
148. Helal, G.A.; Sarhan, M.M.; Abu Shahla, A.N.K.; Abou El-Khair, E.K. Effects of *Cymbopogon citratus* L. essential oil on the growth, lipid content and morphogenesis of *Aspergillus niger* ML2-strain. *J. Basic Microbiol.* **2006**, *46*, 456–469. [[CrossRef](#)] [[PubMed](#)]
149. Rammanee, K.; Hongpattarakere, T. Effects of tropical citrus essential oils on growth, aflatoxin production, and ultrastructure alterations of *Aspergillus flavus* and *Aspergillus parasiticus*. *Food Bioprocess Technol.* **2011**, *4*, 1050–1059. [[CrossRef](#)]
150. Hyldgaard, M.; Mygind, T.; Meyer, R.L. Essential oils in food preservation: Mode of action, synergies, and interactions with food matrix components. *Front. Microbiol.* **2012**, *3*, 1–24. [[CrossRef](#)] [[PubMed](#)]
151. Basak, S.; Guha, P. A review on antifungal activity and mode of action of essential oils and their delivery as nano-sized oil droplets in food system. *J. Food Sci. Technol.* **2018**, *55*, 4701–4710. [[CrossRef](#)]
152. Tariq, S.; Wani, S.; Rasool, W.; Shafi, K.; Bhat, M.A.; Prabhakar, A.; Shalla, A.H.; Rather, M.A. A comprehensive review of the antibacterial, antifungal and antiviral potential of essential oils and their chemical constituents against drug-resistant microbial pathogens. *Microb. Pathog.* **2019**, *134*. [[CrossRef](#)]
153. Shin, S.; Kang, C.A. Antifungal activity of the essential oil of *Agastache rugosa* Kuntze and its synergism with ketoconazole. *Lett. Appl. Microbiol.* **2003**, *36*, 111–115. [[CrossRef](#)]
154. Gong, H.; Li, S.; He, L.; Kasimu, R. Microscopic identification and in vitro activity of *Agastache rugosa* (Fisch. et Mey) from Xinjiang, China. *BMC Complement Altern. Med.* **2017**, *17*, 95. [[CrossRef](#)]
155. Zhang, C.; Li, H.; Yun, T.; Fu, Y.; Liu, C.; Gong, B.; Neng, B. Chemical composition, antimicrobial and antioxidant activities of the essential oil of Tibetan herbal medicine *Dracocephalum heterophyllum* Benth. *Nat. Prod. Res.* **2008**, *22*, 1–11. [[CrossRef](#)]
156. Souza, L.K.; de Oliveira, C.M.; Ferri, P.H.; de Oliveira Júnior, J.G.; de Souza Júnior, A.H.; Fernandes Ode, F.; Silva Mdo, R. Antimicrobial activity of *Hyptis ovalifolia* towards dermatophytes. *Memórias do Instituto Oswaldo Cruz* **2003**, *98*, 963–965. [[CrossRef](#)]
157. Dolatabadi, S.; Salari, Z.; Mahboubi, M. Antifungal effects of *Ziziphora tenuior*, *Lavandula angustifolia*, *Cuminum cyminum* essential oils against clinical isolates of *Candida albicans* from women suffering from vulvovaginal candidiasis. *Infect* **2019**, *23*, 222–226. [[CrossRef](#)]
158. Powers, C.N.; Osier, J.L.; McFeeters, R.L.; Brazell, C.B.; Olsen, E.L.; Moriarity, D.M.; Satyal, P.; Setzer, W.N. Antifungal and cytotoxic activities of sixty commercially-available essential oils. *Molecules* **2018**, *23*, 1549. [[CrossRef](#)] [[PubMed](#)]
159. Ozcakmak, S.; Dervisoglu, M.; Yilmaz, A. Antifungal activity of lemon balm and sage essential oils on the growth of ochratoxigenic *Penicillium verrucosum*. *Afr. J. Microbiol. Res.* **2012**, *6*, 3079–3084. [[CrossRef](#)]
160. Tyagi, A.K.; Malik, A. Liquid and vapour-phase antifungal activities of selected essential oils against *Candida albicans*: Microscopic observations and chemical characterization of *Cymbopogon citratus*. *BMC Complement Altern. Med.* **2010**, *10*, 65. [[CrossRef](#)] [[PubMed](#)]
161. Sharma, A.; Rajendran, S.; Srivastava, A.; Sharma, S.; Kundu, B. Antifungal activities of selected essential oils against *Fusarium oxysporum* f. sp. lycopersici 1322, with emphasis on *Syzygium aromaticum* essential oil. *J. Biosci. Bioeng.* **2017**, *123*, 308–313.
162. Piras, A.; Porcedda, S.; Falconieri, D.; Maxia, A.; Gonçalves, M.; Cavaleiro, C.; Salgueiro, L. Antifungal activity of essential oil from *Mentha spicata* L. and *Mentha pulegium* L. growing wild in Sardinia island (Italy). *Nat. Prod. Res.* **2019**. [[CrossRef](#)]
163. Fancello, F.; Zara, S.; Petretto, G.L.; Chessa, M.; Addis, R.; Rourke, J.P.; Pintore, G. Essential oils from three species of *Mentha* harvested in Sardinia: Chemical characterization and evaluation of their biological activity. *Int. J. Food Prop.* **2017**, *20*, 1751–1761. [[CrossRef](#)]
164. Houicher, A.; Hechachna, H.; Teldji, H.; Ozogul, F. In vitro study of the antifungal activity of essential oils obtained from *Mentha spicata*, *Thymus vulgaris*, and *Laurus nobilis*. *Recent Pat. Food Nutr. Agric.* **2016**, *8*, 99–106. [[CrossRef](#)]



165. Ji, H.; Kim, H.; Beuchat, L.R.; Ryu, J.H. Synergistic antimicrobial activities of essential oil vapours against *Penicillium corylophilum* on a laboratory medium and beef jerky. *Int. J. Food Microbiol.* **2019**, *291*, 104–110. [[CrossRef](#)]
166. Pietrella, D.; Angiolella, L.; Vavala, E.; Rachini, A.; Mondello, F.; Ragno, R.; Bistoni, F.; Vecchiarelli, A. Beneficial effect of *Mentha suaveolens* essential oil in the treatment of vaginal candidiasis assessed by real-time monitoring of infection. *BMC Complement Altern. Med.* **2011**, *11*, 18. [[CrossRef](#)]
167. Angiolella, L.; Vavala, E.; Sivric, S.; D'Auria, F.D.; Ragno, R. In vitro activity of *Mentha suaveolens* essential oil against *Cryptococcus neoformans* and dermatophytes. *Int. J. Essent. Oil Ther.* **2010**, *4*, 35–36.
168. Bisht, D.S.; Padalia, R.C.; Singh, L.; Pande, V.; Lal, P.; Mathela, C.S. Constituents and antimicrobial activity of the essential oils of six Himalayan *Nepeta* species. *J. Serb. Chem. Soc.* **2010**, *75*, 739–747. [[CrossRef](#)]
169. Cardoso, N.N.; Alviano, C.S.; Blank, A.F.; Arrigoni-Blank, M.F.; Romanos, M.T.; Cunha, M.M.; da Silva, A.J.; Alviano, D.S. Anti-cryptococcal activity of ethanol crude extract and hexane fraction from *Ocimum basilicum* var. Maria bonita: Mechanisms of action and synergism with amphotericin B and *Ocimum basilicum* essential oil. *Pharm. Biol.* **2017**, *55*, 1380–1388. [[CrossRef](#)] [[PubMed](#)]
170. Al-Hajj, N.Q.M.; Wang, H.X.; Ma, C.; Lou, Z.; Bashari, M.; Thabit, R. Antimicrobial and antioxidant activities of the essential oils of some aromatic medicinal plants (*Pulicaria inuloides*-Asteraceae and *Ocimum forskolei*-Lamiaceae). *Trop. J. Pharmaceut. Res.* **2014**, *13*, 1287–1293. [[CrossRef](#)]
171. Nakamura, C.V.; Ishida, K.; Faccin, L.C.; Filho, B.P.; Cortez, D.A.; Rozental, S.; de Souza, W.; Ueda-Nakamura, T. In vitro activity of essential oil from *Ocimum gratissimum* L. against four *Candida* species. *Res. Microbiol.* **2004**, *155*, 579–586. [[CrossRef](#)]
172. Silva, M.R.; Oliveira, J.G., Jr.; Fernandes, O.F.; Passos, X.S.; Costa, C.R.; Souza, L.K.; Lemos, J.A.; Paula, J.R. Antifungal activity of *Ocimum gratissimum* towards dermatophytes. *Mycoses* **2005**, *48*, 172–175. [[CrossRef](#)]
173. Bedoya-Serna, C.M.; Dacanal, G.C.; Fernandes, A.M.; Pinho, S.C. Antifungal activity of nanoemulsions encapsulating oregano (*Origanum vulgare*) essential oil: In vitro study and application in Minas Padrão cheese. *Braz. J. Microbiol.* **2018**, *49*, 929–935. [[CrossRef](#)]
174. Vinciguerra, V.; Rojas, F.; Tedesco, V.; Giusiano, G.; Angiolella, L. Chemical characterization and antifungal activity of *Origanum vulgare*, *Thymus vulgaris* essential oils and carvacrol against *Malassezia furfur*. *Nat. Prod. Res.* **2018**, *33*, 3273–3277. [[CrossRef](#)]
175. Wang, G.S.; Deng, J.H.; Ma, Y.H.; Shi, M.; Li, B. Mechanisms, clinically curative effects, and antifungal activities of cinnamon oil and pogostemon oil complex against three species of *Candida*. *J. Tradit. Chin. Med.* **2012**, *32*, 19–24. [[CrossRef](#)]
176. Farisa Banu, S.; Rubini, D.; Shanmugavelan, P.; Murugan, R.; Gowrishankar, S.; Karutha Pandian, S.; Nithyanand, P. Effects of patchouli and cinnamon essential oils on biofilm and hyphae formation by *Candida* species. *J. Mycol. Med.* **2018**, *28*, 332–339. [[CrossRef](#)]
177. Sookto, T.; Srithavaj, T.; Thaweboon, S.; Thaweboon, B.; Shrestha, B. In vitro effects of *Salvia officinalis* L. essential oil on *Candida albicans*. *Asian Pac. J. Trop. Biomed.* **2013**, *3*, 376–380. [[CrossRef](#)]
178. Sharifzadeh, A.; Khosravi, A.R.; Ahmadian, S. Chemical composition and antifungal activity of *Satureja hortensis* L. essential oil against planktonic and biofilm growth of *Candida albicans* isolates from buccal lesions of HIV(+) individuals. *Microb. Pathog.* **2016**, *96*, 1–9. [[CrossRef](#)]
179. Marković, T.; Chatzopoulou, P.; Šiljegović, J.; Nikolić, M.; Glamočlija, J.; Ćirić, A.; Soković, M. Chemical analysis and antimicrobial activities of the essential oils of *Satureja thymbra* L. and *Thymbra spicata* L. and their main components. *Arch. Biol. Sci. Belgrade* **2011**, *63*, 457–464. [[CrossRef](#)]
180. Tabti, L.; Dib Mel, A.; Gaouar, N.; Samira, B.; Tabti, B. Antioxidant and antifungal activity of extracts of the aerial parts of *Thymus capitatus* (L.) Hoffmanns against four phytopathogenic fungi of *Citrus sinensis*. *Jundishapur J. Nat. Pharm. Prod.* **2014**, *9*, 49–54. [[CrossRef](#)] [[PubMed](#)]
181. Nasir, M.; Tafess, K.; Abate, D. Antimicrobial potential of the Ethiopian *Thymus schimperi* essential oil in comparison with others against certain fungal and bacterial species. *BMC Complement Altern. Med.* **2015**, *15*, 260. [[CrossRef](#)] [[PubMed](#)]
182. Sokolić-Mihalak, D.; Frece, J.; Slavica, A.; Delaš, F.; Pavlović, H.; Markov, K. The effects of wild thyme (*Thymus serpyllum* L.) essential oil components against ochratoxin-producing Aspergilli. *Arhiv za Higijenu i Toksikologiju* **2012**, *63*, 457–462. [[CrossRef](#)]

183. Perina, F.J.; Amaral, D.C.; Fernandes, R.S.; Labory, C.R.; Teixeira, G.A.; Alves, E. *Thymus vulgaris* essential oil and thymol against *Alternaria alternata* (Fr.) Keissler: Effects on growth, viability, early infection and cellular mode of action. *Pest Manag. Sci.* **2015**, *71*, 1371–1378. [[CrossRef](#)]
184. Sharifzadeh, A.; Javan, A.J.; Shokri, H.; Abbaszadeh, S.; Keykhosravi, K. Evaluation of antioxidant and antifungal properties of the traditional plants against foodborne fungal pathogens. *J. Mycol. Med.* **2016**, *26*, e11–e17. [[CrossRef](#)]
185. Liu, J.; Sui, G.; He, Y.; Liu, D.; Yan, J.; Liu, S.; Qin, W. Prolonging storage time of baby ginger by using a sand-based storage medium and essential oil treatment. *J. Food Sci.* **2014**, *79*, M593–M599. [[CrossRef](#)]
186. Moghadam, H.D.; Sani, A.M.; Sangatash, M.M. Antifungal activity of essential oil of *Ziziphora clinopodioides* and the inhibition of aflatoxin B1 production in maize grain. *Toxicol. Ind. Health* **2016**, *32*, 493–499. [[CrossRef](#)]
187. Dreger, M.; Wielgus, K. Application of essential oils as natural cosmetic preservatives. *Herba Pol.* **2013**, *59*, 142–156. [[CrossRef](#)]
188. Sarkic, A.; Stappen, I. Essential oils and their single compounds in cosmetics—A critical review. *Cosmetics* **2018**, *5*, 11. [[CrossRef](#)]
189. Ali, B.; Al-Wabel, N.A.; Shams, S.; Ahamad, A.; Khan, S.A.; Anwar, F. Essential oils used in aromatherapy: A systemic review. *Asian Pac. J. Trop. Biomed.* **2015**, *5*, 601–611. [[CrossRef](#)]
190. Helms, S.; Miller, A.L. Natural treatment of chronic rhinosinusitis. *Altern. Med. Rev.* **2006**, *11*, 196–207.
191. Schilcher, H. Efficient phytotherapy. Herbal medicines in the upper respiratory tract for catarrh. *Herba Pol.* **2000**, *46*, 52–57.
192. Vranic, E.; Lacević, A.; Mehmedagić, A.; Uzunovic, A. Formulation ingredients for toothpastes and mouthwashes. *Bosnian J. Basic Med. Sci.* **2004**, *4*, 51–58. [[CrossRef](#)]
193. Guven, Y.; Ustun, N.; Tuna, E.B.; Aktoren, O. Antimicrobial effect of newly formulated toothpastes and a mouthrinse on specific microorganisms: An in vitro study. *Eur. J. Dent.* **2019**, *13*, 172–177. [[CrossRef](#)]
194. DOZ Pharmacy. Available online: <https://www.doz.pl/> (accessed on 17 December 2019).
195. Ehlers, V.; Helm, S.; Kasaj, A.; Willershausen, B. The effect of Parodontax® on the MMP-8 concentration in gingivitis patients. *Schweiz Monatsschr. Zahnmed.* **2011**, *121*, 1041–1051.
196. Adwan, G.; Salameh, Y.; Adwan, K.; Barakat, A. Assessment of antifungal activity of herbal and conventional toothpastes against clinical isolates of *Candida albicans*. *Asian Pac. J. Trop. Biomed.* **2012**, *2*, 375–379. [[CrossRef](#)]



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