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“Clocks” in the NAD World: NAD as a Metabolic Oscillator for the Regulation of Metabolism and Aging

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Abstract

SIR2 (silent information regulator 2) proteins, now called “sirtuins,” are an evolutionarily conserved family of NAD-dependent protein deacetylases/ADP-ribosyltransferases. Sirtuins have recently attracted major attention in the field of aging research, and it has been demonstrated that SIR2 and its orthologs regulate aging and longevity in yeast, worms, and flies. In mammals, the SIR2 ortholog SIRT1 coordinates important metabolic responses to nutritional availability in multiple tissues. Most recently, it has been demonstrated that SIRT1 regulates the amplitude and the duration of circadian gene expression through the interaction and the deacetylation of key circadian clock regulators, such as BMAL1 and PER2. More strikingly, we and others have discovered a novel circadian clock feedback loop in which both the rate-limiting enzyme in mammalian NAD biosynthesis, nicotinamide phosphoribosyltransferase (NAMPT), and NAD levels display circadian oscillations and modulate CLOCK:BMAL1-mediated circadian transcriptional regulation through SIRT1, demonstrating a new function of NAD as a “metabolic oscillator.” These findings reveal a novel system dynamics of a recently proposed systemic regulatory network regulated by NAMPT-mediated NAD biosynthesis and SIRT1, namely, the NAD World. In the light of this concept, a new connection between physiological rhythmicity, metabolism, and aging will be discussed.

Introduction

In the history of aging research, there has been an idea that the life-long sequence of age-associated events might be controlled by a sort of “clock.” This idea of “the aging clock” was first proposed by Arthur Everitt in Australia in 1973 [1]. In his article, he speculated that such a clock or even multiple clocks might be located in the hypothalamus. He also tried to integrate this idea into the hypothalamic/neuroendocrine theory of aging proposed by Vladimir Dilman in Russia in 1971 [2]. Dilman recognized aging as “a process of disordered homeostatic stability of internal environment” and assumed that “permanent deviations from the law of constancy of the internal environment” could ultimately produce “the specific lesions of ageing” [2]. Both Dilman and Everitt suspected that some feedback mechanism that involves the hypothalamus might be running “the genetic programme of development and ageing” or ticking “the aging clock” [1,2].

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Intriguingly, almost at the same time when those ideas were proposed, studies began to reveal that a hypothalamic region called the suprachiasmatic nucleus (SCN) actually has such a “clock” function [3]. For example, it was reported in 1972 that bilateral lesions in the SCN eliminated circadian rhythms in rats [4,5]. Since then, remarkable progress has been made in the biology of circadian rhythm, and major regulators in the core clock mechanism have been identified [6-8]. These findings have clearly shown that major clock regulators function not only in the SCN but also in many different peripheral tissues, such as the liver and white adipose tissue (WAT), suggesting that circadian rhythmicity of physiological events is regulated by a network of multiple oscillators throughout the body [8]. It has also been reported that both amplitude and phase of circadian rhythms are affected in the process of aging [9-11]. Although these age-associated changes in circadian rhythms are rather complex, aging might affect interactions among circadian oscillators and compromise “sustainability” of rhythmic tissues [11], which could cause the “permanent deviations” from stable rhythmicity that Dilman speculated in his idea [2].

Despite all these studies, there had been neither supportive evidence for the idea of “the aging clock” nor direct molecular connection between clocks and aging until recently. Most recently, however, there have been exciting developments in the juncture of the fields of aging and circadian rhythm researches which bring up important insights into a fascinating connection between physiological rhythmicity, metabolism, and aging. Therefore, in this review, I will first summarize these new findings and then revisit the idea of “the aging clock” in the light of a new concept of the NAD World, a systemic regulatory network for the regulation of metabolism and aging [12,13].

SIRT1 and circadian clock regulators

For the past couple of decades, important regulators and signaling pathways have been identified in model organisms that, when modified, may slow down aging [14-17]. One such regulator has recently attracted major attention in the field of aging research. It is the SIR2 (silent information regulator 2) protein family, now called “sirtuins” [15,18-21]. Sirtuins are an evolutionarily conserved family of NAD-dependent protein deacetylases/ADP-ribosyltransferases. In yeast, worms, and flies, SIR2 and its orthologs regulate aging and longevity [22-27] and in certain genetic backgrounds, also mediate the lifespan-extending effect of caloric restriction [25,28-31], the most consistent dietary regimen that retards aging and extends longevity in many diverse species. There are seven mammalian sirtuins, SIRT1 through SIRT7, and the majority of mammalian sirtuin research has so far focused on the function of the SIR2 ortholog SIRT1 [15,19,21]. Although it has not yet been proven whether SIRT1 regulates aging and longevity in mammals, it has been firmly established that SIRT1 plays an important role in the regulation of metabolism in response to nutritional availability in multiple tissues. Details of reported SIRT1 functions have already been summarized and discussed in many review articles [19,21,32-34]. The most important theme through all of these SIRT1 studies is that SIRT1 coordinates physiological programs that allow animals to survive through nutritionally scarce conditions by mediating critical metabolic responses to nutritional cues, particularly in low nutritional conditions such as fasting and caloric restriction, [13,32, 35]. This unique aspect of SIRT1 function and the absolute requirement of NAD for its activity place this protein at a central position as a key regulator that connects metabolism and aging.

In 2008, two groups further stretched the physiological importance of SIRT1 towards another fascinating direction. They reported that SIRT1 regulates the circadian clock oscillatory mechanisms and affects the expression of circadian clock genes (Fig. 1) [36,37]. Nakahata *et al.* [37] found that SIRT1 shows the oscillation of its enzymatic activity and deacetylates BMAL1, one of the critical regulators in the core clock mechanism, and histone H3 in a circadian manner. SIRT1 physically interacts with CLOCK, another key clock regulator that

heterodimerizes with and acetylates BMAL1, and is recruited to the CLOCK:BMAL1 complex at the promoters of circadian clock genes (Fig. 1). They also found that a SIRT1 deficiency causes changes in the expression of circadian clock genes, such as *Period2* (*Per2*), in mouse embryonic fibroblasts (MEFs) *in vitro* and in the liver *in vivo*, including an augmentation of the expression levels and a broadening of the oscillatory cycles. On the other hand, Asher *et al.* [36] reported that SIRT1 protein levels oscillate in MEFs and in the liver and that SIRT1 activity is required for the robust oscillatory expression patterns of circadian clock genes. They likewise found that SIRT1 interacts with the CLOCK:BMAL1 complex in a circadian manner. They also showed that SIRT1 interacts with and deacetylates PER2, resulting in its degradation (Fig. 1). Although both groups indicate the importance of SIRT1 in the regulation of circadian clock gene expression, there are some discrepancies between results from these groups. One example is whether SIRT1 protein levels or activity levels oscillate. Another example is whether SIRT1 negatively or positively regulates the amplitude of the circadian gene expression. Even though there are no reconcilable explanations for these discrepancies at this moment, it is now clear that SIRT1 regulates the amplitude and the duration of circadian gene expression through the deacetylation of key circadian clock regulators, such as BMAL1 and PER2.

These studies have demonstrated the first connection between key regulators for aging and circadian rhythm. Intriguingly, *Bmal1*-deficient mice and *Per1/2*-deficient mice develop some pathological features at earlier ages that resemble those in aged mice, providing indirect support for the connection between aging and circadian rhythm [38,39]. Furthermore, the core molecular clock machinery has also been demonstrated to be one of the most powerful modifiers of metabolism [40,41]. For example, homozygous *Clock* mutant mice exhibit metabolic dysregulation, including hyperlipidemia, hyperglycemia, hyperleptinemia, hypoinsulinemia, and hepatic steatosis [42]. Liver-specific deletion of *Bmal1* causes loss of rhythmic expression of clock-regulated metabolic genes in the liver and hypoglycemia in the fasting phase of the daily feeding cycle [43]. Hepatic *Bmal1* expression is also regulated by PGC-1 α , another SIRT1 target transcription factor that regulates glucose production in the liver, and liver-specific PGC-1 α knockdown in mice proves that this factor is required for hepatic clock function [44]. These findings, as well as new studies described above, have implicated a critical connection between physiological rhythmicity, metabolism, and aging, and SIRT1 might function at a central interface connecting these fundamental biological events. However, how exactly SIRT1 functions at such a fundamental junction was a big question back then, and indeed, this particular question awaited a much more surprising answer.

NAD as a metabolic oscillator in a novel circadian clock feedback cycle

The key to solve this question was the fact that sirtuins require NAD for its enzymatic activity [45-47]. NAD is an essential coenzyme that is synthesized from three major precursors - tryptophan, nicotinic acid, and nicotinamide [48,49]. The predominant NAD biosynthetic pathway in mammals involves the conversion of nicotinamide (a form of vitamin B₃) and 5'-phosphoribosyl-1-pyrophosphate (5'-PRPP) to nicotinamide mononucleotide (NMN) by the rate-limiting enzyme nicotinamide phosphoribosyltransferase (NAMPT) (Fig. 2). Whereas the enzymatic activity of NAMPT was originally reported in 1957 [50], it was in 2001 that the gene encoding NAMPT was first identified in *Haemophilus ducreyi* [51]. Since then, the enzymological features and the crystal structures of NAMPT have been studied extensively [52-56]. Interestingly, NAMPT has intra- and extracellular forms (iNAMPT and eNAMPT, respectively), and eNAMPT (a.k.a. PBEF/visfatin), which is positively secreted through a non-classical secretory pathway by fully differentiated mouse and human adipocytes, also exhibits robust NAD biosynthetic activity compared to iNAMPT, likely contributing to the extracellular biosynthesis of the key NAD intermediate NMN in mammals [57,58]. Indeed, NMN and other NAD intermediates, such as nicotinamide riboside (NR), can be detected in both mouse and

human plasma [58, unpublished observation]. NAMPT has recently been shown to play important roles in a variety of biological events and has drawn much attention in several different fields, including NAD biology, metabolism, and immune response [57,59]. Most importantly, accumulating lines of evidence have demonstrated that NAMPT-mediated NAD biosynthesis plays a critical role in the regulation of SIRT1 activity in many different cell types [54,60-67]. Increased dosage of iNAMPT enhances total cellular NAD levels and thereby SIRT1 activity in mouse fibroblasts [54]. In human vascular smooth muscle cells, iNAMPT promotes their maturation [66] and cellular life span [61,68] through enhanced SIRT1 activity. In cardiac myocytes, increased iNAMPT levels protect them from cell death through SIRT1 [69]. In pancreatic β cells, both SIRT1 and NAMPT-mediated NAD biosynthesis play important roles in the regulation of glucose-stimulated insulin secretion [58,70,71]. In skeletal myoblasts, glucose restriction inhibits their differentiation through the AMP-activated protein kinase (AMPK)-dependent induction of *Nampt* expression and the resultant activation of SIRT1 [60]. The NAMPT/SIRT1 pathway also mediates granulocyte colony-stimulating factor (G-CSF)-triggered granulocyte differentiation *in vitro* and *in vivo* [65]. iNAMPT also plays an important role in NAD biosynthesis and sirtuin activation in mitochondria [72]. These findings led us to hypothesize that NAMPT-mediated NAD biosynthesis may coordinate physiological rhythmicity, metabolism, and aging through the regulation of SIRT1 in metabolic tissues.

Pursuing this hypothesis with Joseph Bass's group in Northwestern University, we have made a striking discovery that levels of NAMPT and NAD display circadian oscillations that are regulated by the core clock machinery in mice (Fig. 3) [64]. Almost at the same time, Paolo Sassone-Corsi's group reached the same conclusion using MEFs [63]. The first important clue came from the observation that expression levels of *Nampt* RNA display a robust diurnal oscillation in the liver and WAT, with a peak around the beginning of the dark period (*zeitgeber* time (ZT) 14) [64]. A similar oscillatory pattern of *Nampt* expression was observed in serum-entrained wild-type MEFs [63]. Interestingly, expression levels of NAMPT protein also show a diurnal oscillation in the liver. This oscillation of NAMPT protein is bimodal, with a reduction in NAMPT protein levels prior to the onset of the dark period, which is consistent with the time when mice usually start eating. This would imply some post-translational regulation on NAMPT protein in response to a nutritional surge or ingestive behavior. The oscillation of *Nampt* RNA is still robust in the liver even when mice are maintained in constant darkness, demonstrating that *Nampt* RNA oscillation is circadian in nature. These robust diurnal and circadian oscillation patterns of *Nampt* RNA and protein are completely abolished in tissues from the circadian rhythm-deficient *Clock* ^{Δ 19} mutant mice, indicating that the core clock machinery is required for the circadian control of *Nampt* expression. Most importantly, NAD levels show a very similar bimodal circadian oscillation pattern to that of NAMPT in the liver, and NAD levels are significantly reduced in *Clock* ^{Δ 19} mutant liver during both the light (ZT2) and dark (ZT14) periods. This NAD oscillation is recapitulated in serum-entrained MEFs, although there is no bimodality in their oscillatory pattern [63]. Additionally, consistent with our *in vivo* results, NAD levels are significantly reduced in serum-stimulated *Clock* mutant cells [63]. Mice deficient in BMAL1, the heterodimeric binding partner of CLOCK, also exhibit a significant reduction in both *Nampt* RNA and NAD levels in the liver. Therefore, these findings demonstrate that the rhythmic oscillation in RNA and protein levels of NAMPT, which is regulated by the CLOCK:BMAL1-mediated core clock mechanism, leads to a circadian oscillation of NAD levels *in vivo* (Fig. 3).

Given that SIRT1 regulates CLOCK:BMAL1-mediated circadian transcription [36,37], it is conceivable that NAMPT-mediated NAD biosynthesis might also control CLOCK:BMAL1-mediated transcription through SIRT1. Indeed, inhibition of NAMPT by FK866, a potent chemical inhibitor of NAMPT [73], abrogates the SIRT1-dependent suppression of CLOCK:BMAL1-mediated transcription and promotes a more robust oscillation of clock target gene expression in primary hepatocytes [64] or clock gene oscillations with earlier onsets

and higher amplitudes in serum-entrained MEFs [63], suggesting that the NAMPT/NAD-driven pathway modulates circadian transcriptional regulation in mammals. The final important piece of evidence is that the CLOCK:BMAL1 complex binds to canonical and noncanonical E-box motifs in the promoter and the first intron of the *Nampt* gene and up-regulates *Nampt* transcription [63,64], thus completing a novel circadian clock feedback loop involving NAMPT/NAD and SIRT1/CLOCK:BMAL1 (Fig. 3).

In the NAMPT/NAD-driven feedback loop, NAD functions as a “metabolic oscillator” and regulates the core clock machinery through SIRT1 (Fig. 3) [64]. Through this tight coupling of a classical transcriptional circadian loop to a novel enzymatic feedback loop, two major biological system, metabolism and circadian clock, are interlocked [63]. Given that NAD has long been considered a stable pool of energy in cells and tissues, the circadian oscillation of NAD levels is striking, and this discovery reveals a novel system dynamics of the regulatory network comprised of NAMPT-mediated NAD biosynthesis and SIRT1. Through the regulation of SIRT1 activity, the circadian oscillatory production of NAD might convey a cascade of effects on a variety of downstream pathways, including epigenetic chromatin regulation, stress response, metabolism, and possibly aging. Therefore, in the next section, I would like to discuss a possibility that this novel circadian oscillatory mechanism might function as “the aging clock” in a recently proposed systemic regulatory network regulated by NAMPT-mediated NAD biosynthesis and SIRT1, namely, the NAD World.

The aging clock(s) in the NAD World

The NAD World is a comprehensive concept that has been proposed to explain a systemic regulatory network connecting metabolism and aging. This concept also conveys the ideas of functional hierarchy and frailty for the induction of aging in mammals [12,13]. NAMPT-mediated NAD biosynthesis and SIRT1 are two critical components that comprise the NAD World. Conceptually, NAMPT-mediated NAD biosynthesis functions as a driver or a pacemaker that keeps up the pace of metabolism throughout the body. iNAMPT and eNAMPT together play a critical role in the systemic regulation of NAD biosynthesis through the intra- and extracellular biosynthesis of NMN, promoting total NAD biosynthesis and thereby contributing to the fine-tuning of SIRT1 activity [57]. SIRT1 functions as a key downstream mediator that controls physiological responses to alterations in NAMPT-mediated NAD biosynthesis in a tissue-dependent manner. The intimate connection between these two critical components of the NAD World has been demonstrated in a number of different cell types, such as pancreatic β cells [74], vascular smooth muscle cells [61,66], skeletal myoblasts [60], cardiac myocytes [62,75], and others [63-65,67]. Thus, through this interplay between NAMPT-mediated systemic NAD biosynthesis and SIRT1, the NAD World orchestrates physiological responses to a variety of nutritional and environmental inputs and maintains the robustness of the physiological system in mammals (Fig. 4) [12,13].

As described in the previous section, we have now come to know that NAMPT/NAD drives the circadian clock feedback cycle through SIRT1 and CLOCK:BMAL1. Whereas this “clock” likely ticks in major metabolic tissues, such as liver and WAT, there are tissues/organs that do not have adequate amounts of iNAMPT and therefore rely on circulating NMN and other NAD intermediates to maintain sufficient NAD biosynthesis for their functions. Pancreatic β cells and brain (neurons), both of which have very low levels of iNAMPT [58], are likely such tissues/organs. In those tissues/organs, iNAMPT-dependent oscillatory production of NAD may not be able to influence SIRT1 activity effectively. Then, an important question is whether plasma eNAMPT and NMN levels also display circadian rhythm. Further investigation will be required to have the answer to this particular question. However, if this is the case, one could speculate that eNAMPT and/or NMN might function as a key factor that synchronizes the circadian oscillation of NAD production and thereby SIRT1 activity through the whole body.

This idea also implies that NAD-dependent “clocks” in tissues/organs like pancreatic β cells and neurons might be driven or influenced by other tissues that are capable of propagating their own rhythm through the control of eNAMPT secretion (Fig. 4). In this regard, it will be of great interest to examine whether the adipose tissue has any role in the synchronization of NAD-dependent physiological rhythmicity at a systemic level by knocking out the *Nampt* gene specifically in adipose tissues.

Another critical prediction is that those tissues/organs that do not have adequate amounts of iNAMPT are likely important frailty points in the NAD World [12,13]. If systemic NAD biosynthesis starts declining, these frailty points would be the first that responds to this change and starts having functional problems due to insufficient NAD biosynthesis and thereby reduced SIRT1 activity. Indeed, we have previously demonstrated that a decrease in NAMPT-mediated systemic NAD biosynthesis, which appears to be happening during the course of aging, causes significant reductions in SIRT1 activity and glucose-stimulated insulin secretion in pancreatic β cells in aged mice [74]. Strikingly, administration of NMN restores higher levels of glucose-stimulated insulin secretion in aged mice, providing support for the notion that NAMPT-mediated systemic NAD biosynthesis declines over age [74]. Therefore, pancreatic β cells are definitely an important frailty point in the NAD World that is susceptible to changes in NAMPT-mediated systemic NAD biosynthesis. In the case of brain or neurons, it has long been known that one of the triad symptoms in pellagra, the vitamin B₃ deficiency, is dementia [76]. Although the mechanistic connection between vitamin B₃ deficiency and dementia is unknown, it is conceivable that the age-associated decline in systemic NAD biosynthesis causes functional deficits in neurons and results in neurological problems, including dementia. To address this possibility, the connection between NAMPT-mediated NAD biosynthesis and neural function is currently under investigation.

Once pancreatic β cells and brain start having functional problems due to insufficient NAD biosynthesis, other peripheral tissues/organs would also be affected through insulin secretion and central metabolic regulation, resulting in the gradual deterioration of the physiological robustness through the entire body. Therefore, in the concept of the NAD World, aging is considered as the process in which organismal robustness gradually breaks down according to a functional hierarchy determined by the susceptibility to systemic NAD biosynthesis. Additionally, tissues that control the NAD-dependent branch of circadian clock feedback regulations throughout the body could be considered as “the aging clock”, and any imbalance in this fine-tuning system, which could be imposed by long-term nutritional and environmental perturbations, might initiate the process of organismal robustness breakdown (Fig. 4). Although further investigation will be necessary, tweaking “the aging clock”, possibly by supplementing NAD intermediates to systemic NAD biosynthesis, might be an effective way to make all of our clocks tick robustly in this otherwise inevitable process of aging.

Conclusion

This review article focuses on a novel aspect of the connection between physiological rhythmicity, metabolism, and possibly aging. Particularly, the recent finding that NAD functions as a “metabolic oscillator” opens new possibilities to further dissect the complex interlocked feedback system controlling circadian rhythm and metabolism and to better understand the pathophysiology of metabolic complications caused by dysregulation or imbalance in this feedback system. Such dysregulation/imbalance might also be caused in the process of aging, and the concept of the NAD World provides some ideas and predictions on a potential role of “aging clock” tissues/organs in the systemic regulation of aging. Of course, further investigation will be necessary to address this novel concept. For example, evidence connecting altered sirtuin function and NAD levels to aging and longevity is still not strong in mammals. The primacy of NAD for the regulation of brain and β cell functions also needs to

be studied extensively. Nonetheless, in the next decade, this new NAD World will be further explored, and we might eventually find a way to manipulate the process of aging based on our knowledge on the detailed system structure and dynamics of this complex regulatory network.

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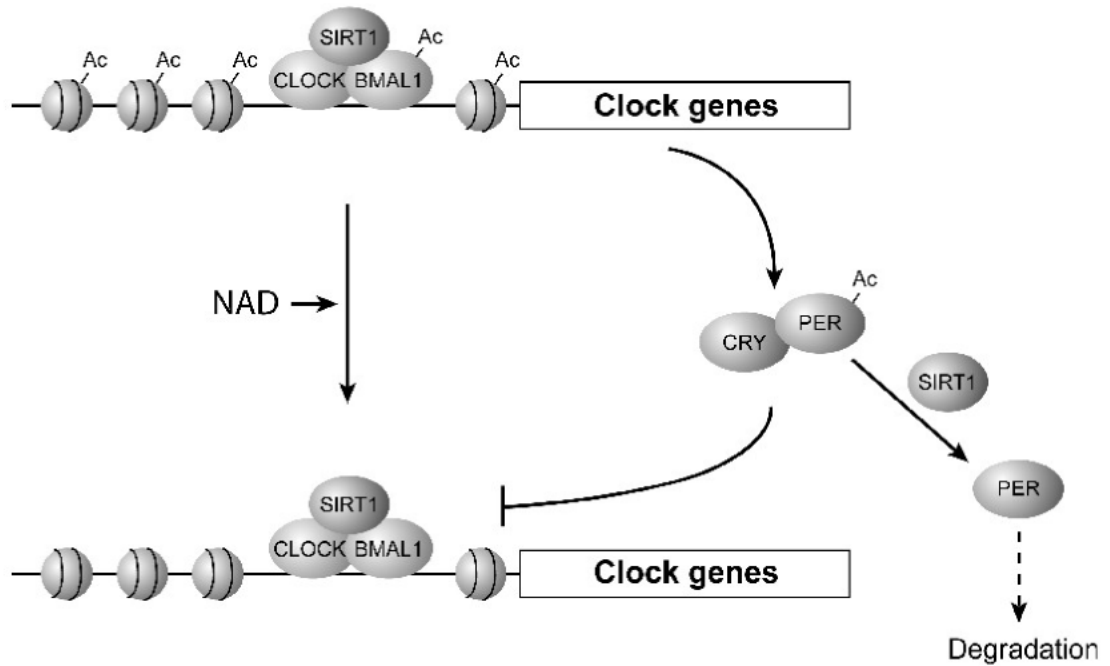


Fig. 1.

A schematic diagram of the SIRT1-mediated regulation of the CLOCK:BMAL1-dependent circadian transcription. SIRT1 physically interacts with CLOCK, a key clock regulator that heterodimerizes with BMAL1, and deacetylates BMAL1 and histone H3 in a circadian manner at the promoters of circadian clock genes. SIRT1 also interacts with and deacetylates PER2, which heterodimerizes with CRY proteins and inhibits the CLOCK:BMAL1 function, and promotes its degradation. See text for details.

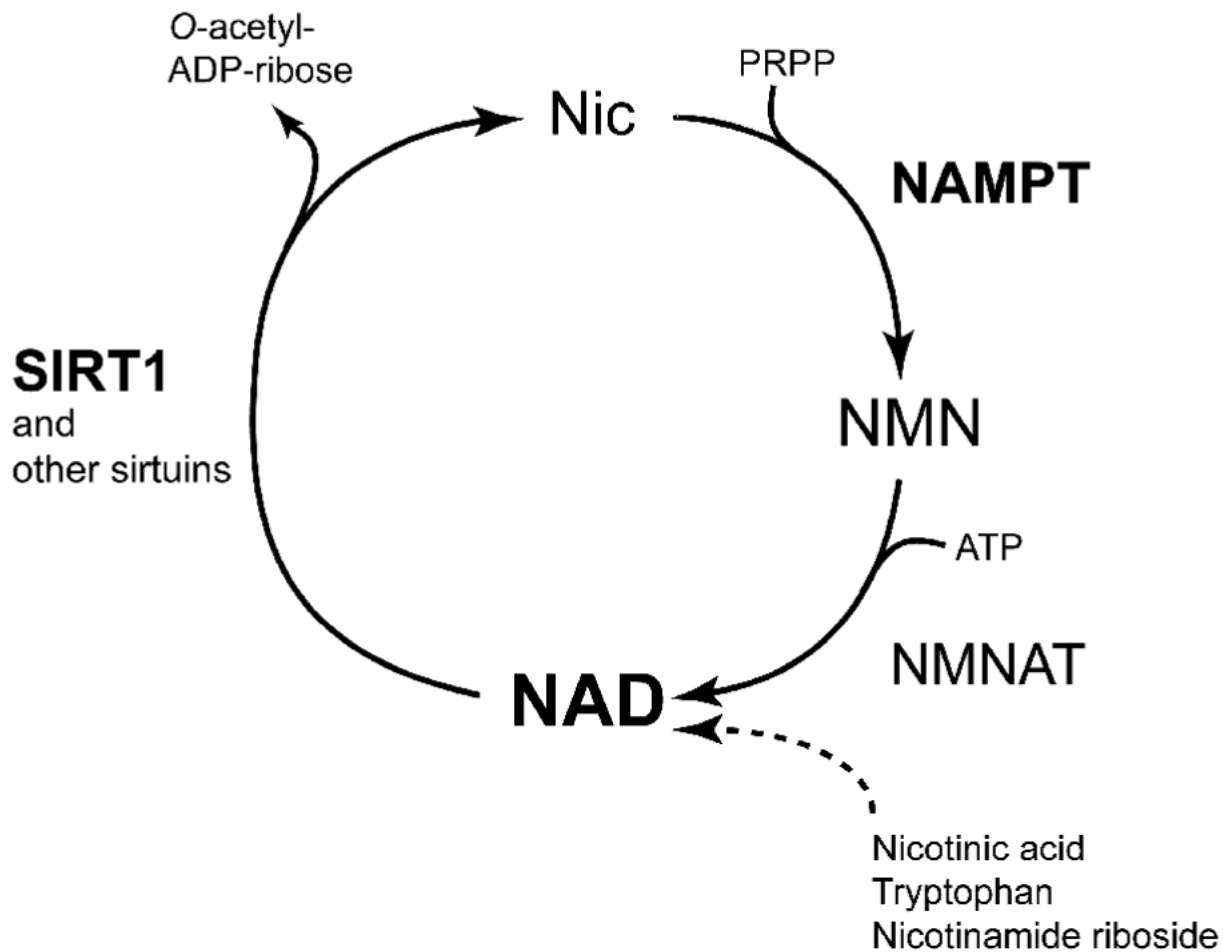


Fig. 2. NAD biosynthesis from nicotinamide in mammals. Nicotinamide (Nic) is the main precursor for mammalian NAD biosynthesis, and nicotinamide phosphoribosyltransferase (NAMPT) catalyzes the rate-limiting step in this pathway, producing nicotinamide mononucleotide (NMN) from nicotinamide and 5'-phosphoribosyl-1-pyrophosphate (PRPP). Then, nicotinamide/nicotinic acid mononucleotide adenylyltransferase (NMNAT) completes the conversion of NMN and ATP to NAD. Other pathways from nicotinic acid, tryptophan, and nicotinamide riboside are not shown. SIRT1 and other sirtuins use NAD for their functions and produce nicotinamide and *O*-acetyl-ADP-ribose. Other NAD-consuming enzymes are not shown in this figure.

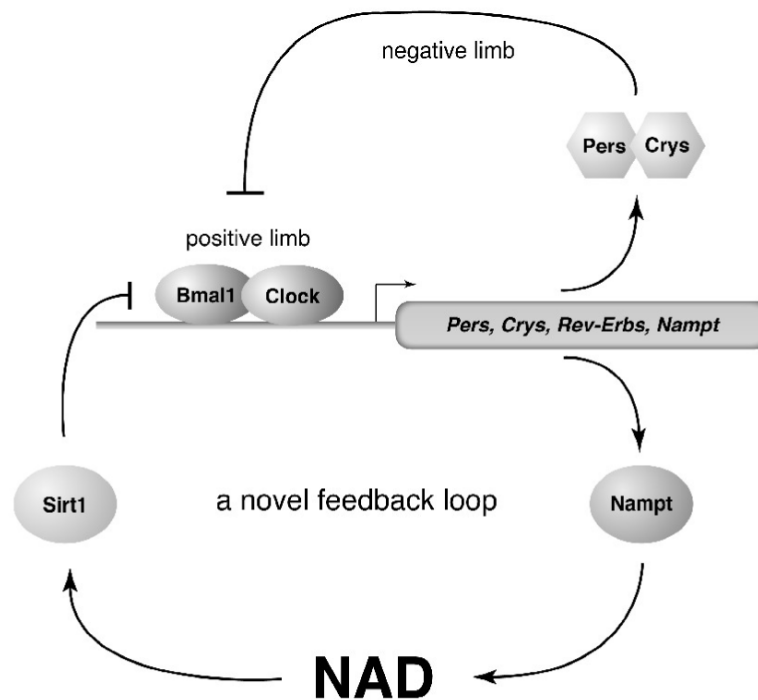


Fig. 3.

A novel NAMPT/NAD-driven feedback loop through SIRT1 and CLOCK:BMAL1. Whereas CLOCK:BMAL1-mediated circadian transcription comprises a positive limb of the transcription-translation circadian feedback loop, the complex of PER and CRY proteins inhibits the function of CLOCK:BMAL1, forming the negative limb of this feedback loop. NAMPT/NAD/SIRT1 comprises a novel circadian feedback cycle that mediates rhythmic regulation of many physiological events. In this NAMPT/NAD-driven feedback loop, NAD functions as a “metabolic oscillator.” See text for details.

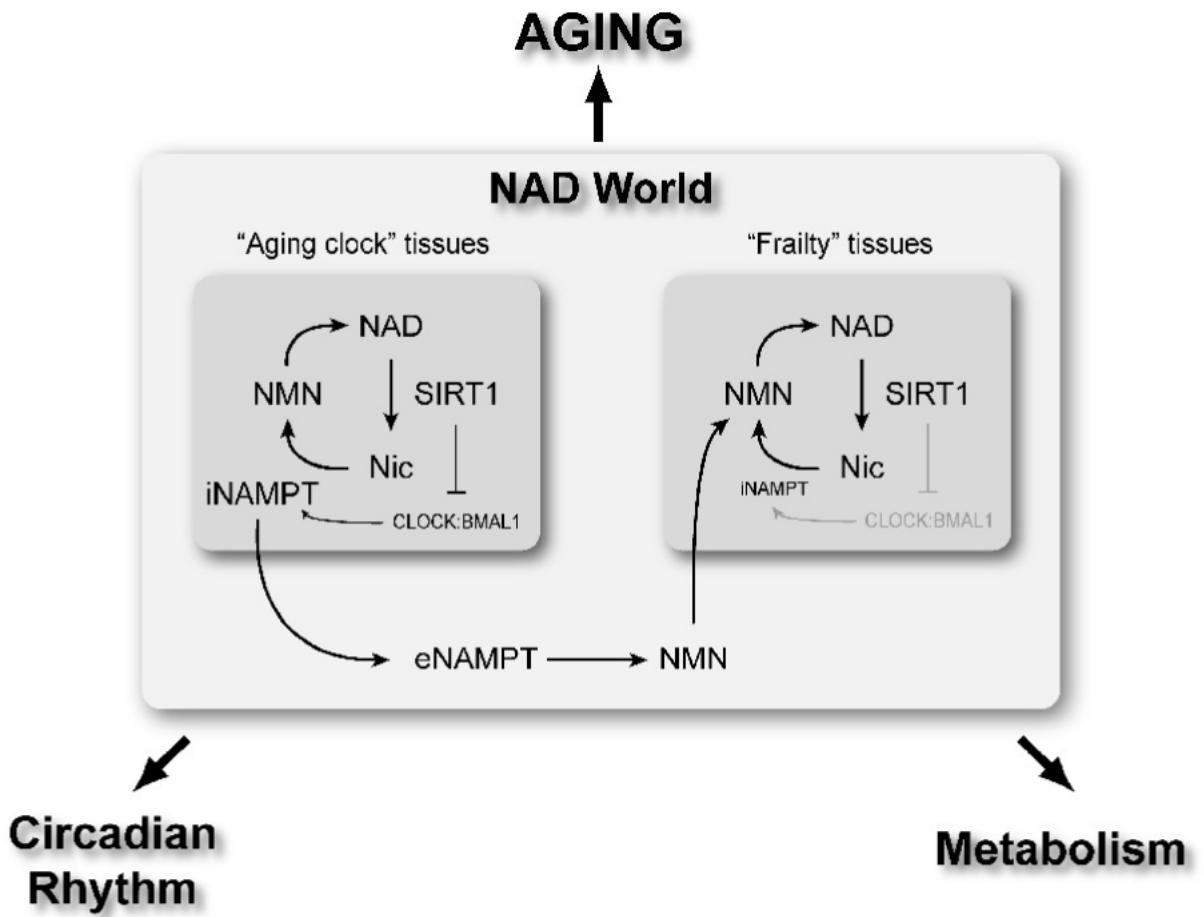


Fig. 4. A schematic model of the interplay between tissues that provide “the aging clock” function and tissues that are frailty points in the NAD World. This interplay plays a critical role in the maintenance of systemic robustness through the entire body. The NAD World functions to orchestrate physiological responses to nutritional and environmental inputs and connects physiological rhythmicity, metabolism, and aging.