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Spatial structure, cooperation and competition in biofilms

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Abstract

Bacteria often live within matrix-embedded communities, termed biofilms, which are now understood to be a major mode of microbial life. The study of biofilms has revealed their vast complexity both in terms of resident species composition and phenotypic diversity. Despite this complexity, theoretical and experimental work in the past decade has identified common principles for understanding microbial biofilms. In this Review, we discuss how the spatial arrangement of genotypes within a community influences the coc

and competitive cell–cell interactions that define biofilm form and function. Furthermore, we argue that a perspective rooted in ecology and evolution is fundamental to progress in microbiology.

Key points

- Bacteria often exist in biofilms, which are surface-adhering or free-floating groups of cells that are bound together by a secreted polymer matrix. These microbial collectives are important for bacterial occupation of diverse ecological niches, they contribute to biogeochemical cycling, and they cause disease in multicellular organisms.
- Within biofilms, bacteria interact with each other closely through cooperative phenotypes, such as the production of digestive enzymes, and antagonistic phenotypes, such as the expression of type V or type VI secretion systems. The evolutionary dynamics of these social phenotypes depend on their costs and their effects on other cells.
- Many bacterial social phenotypes involve the secretion of products that affect neighbours in a distance-dependent manner. As a result, interaction networks within biofilms are largely determined by the spatial structure of the biofilms — that is, the arrangement in space of different clones, strains and species.
- When biofilms are segregated into clonal clusters, the neighbourhood of a given cell mostly contains clonemates, and natural selection often favours the secretion of compounds that benefit all recipient cells. However, when different strains and species are spatially mixed within biofilms, cells primarily interact with other genotypes and antagonistic behaviour is often favoured. Under certain circumstances, between-species commensalism or mutualism can also evolve and remain stable against cheating.
- Cooperative and antagonistic phenotypes fall under the control of sophisticated sensory mechanisms, such as competition sensing and quorum sensing, that evolved to help account for the variation in exposure to other strains and species in space and time. These regulatory mechanisms help to reduce the marginal costs of social phenotypes, maximize their fitness impacts and ensure that the correct

recipient cells are targeted.

- Both cooperative and antagonistic behaviours feed back onto population spatial structure by locally altering the growth rates of other cells and thus changing local biofilm composition.
- Many bacteria and unicellular eukaryotes have evolved strategies for actively altering biofilm population structure, either through selective adhesion that spatially sorts the biofilm into groups that contain one or more specific genotypes or through the secretion of extracellular matrix components that spatially organize biofilm-dwelling cells.

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References

1. Hall-Stoodley, L., Costerton, J. W. & Stoodley, P. Bacterial biofilms: from the natural environment to infectious diseases. *Nat. Rev. Microbiol.* **2**, 95–108 (2004).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

2. Hobley, L., Harkins, C., MacPhee, C. E. & Stanley-Wall, N. R. Giving structure to the biofilm matrix: an overview of individual strategies and emerging common themes. *FEMS Microbiol. Rev.* **39**, 649–669 (2015).

[Show context](#)[PubMed](#) [Article](#) [Google Scholar](#)

3. Arnosti, C. Microbial extracellular enzymes and the marine carbon cycle. *Annu. Rev. Mar. Sci.* **3**, 401–425 (2011).

[Show context](#)[Article](#) [Google Scholar](#)

4. Battin, T. J., Kaplan, L. A., Newbold, J. D. & Hansen, C. M. E. Contributions of microbial biofilms to ecosystem processes in stream mesocosms. *Nature* **426**, 439–442 (2003).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

5. Macfarlane, S., Bahrami, B. & Macfarlane, G. T. Mucosal biofilm communities in the human intestinal tract. *Adv. Appl. Microbiol.* **75**, 111–143 (2011).

[Show context](#)[CAS](#) [PubMed](#) [Google Scholar](#)

-
6. Hoiby, N., Bjarnsholt, T., Givskov, M., Molin, S. & Ciofu, O. Antibiotic resistance of bacterial biofilms. *Int. J. Antimicrob. Agents* **35**, 322–332 (2010).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

7. Bixler, G. D. & Bhushan, B. Biofouling: lessons from nature. *Philos. Trans. A Math. Phys. Eng. Sci.* **370**, 2381–2417 (2012).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

8. Drescher, K., Shen, Y., Bassler, B. L. & Stone, H. A. Biofilm streamers cause catastrophic disruption of flow with consequences for environmental and medical systems. *Proc. Natl Acad. Sci. USA* **110**, 4345–4350 (2013).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

9. Harding, J. L. & Reynolds, M. M. Combating medical device fouling. *Trends Biotechnol.* **32**, 140–146 (2014).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

10. Nadell, C. D. *et al.* Cutting through the complexity of cell collectives. *Proc. Biol. Sci.* **280**, 20122770 (2013).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

11. Nadell, C. D., Xavier, J. B. & Foster, K. R. The sociobiology of biofilms. *FEMS Microbiol. Rev.* **33**, 206–224 (2009).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
12. Visca, P., Imperi, F. & Lamont, I. Pyoverdine siderophores: from biogenesis to biosignificance. *Trends Microbiol.* **15**, 22–30 (2007).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
13. Griffin, A. S., West, S. A. & Buckling, A. Cooperation and competition in pathogenic bacteria. *Nature* **430**, 1024–1027 (2004). **A key proof-of-principle investigation finding that secreted siderophores can act as public goods that are susceptible to the evolution of cheating behaviour.**

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
14. Allison, S. D. Cheaters, diffusion and nutrients constrain decomposition by microbial enzymes in spatially structured environments. *Ecol. Lett.* **8**, 626–635 (2005).

[Show context](#)[Article](#)[Google Scholar](#)

-
15. Absalon, C., Van Dellen, K. & Watnick, P. I. A communal bacterial adhesin anchors biofilm and bystander cells to surfaces. *PLoS Pathog.* **7**, e1002210 (2011).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
16. Xavier, J. B., Kim, W. & Foster, K. R. A molecular mechanism that stabilizes cooperative secretions in *Pseudomonas aeruginosa*. *Mol. Microbiol.* **79**, 166–179 (2011).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
17. Flemming, H.-C. & Wingender, J. The biofilm matrix. *Nat. Rev. Microbiol.* **8**, 623–633 (2010).

[Show context](#)[CAS](#) [PubMed](#) [Google Scholar](#)

-
18. West, S. A., Diggle, S. P., Buckling, A., Gardner, A. & Griffins, A. S. The social lives of microbes. *Annu. Rev. Ecol. Evol. Syst.* **38**, 53–77 (2007).

[Show context](#)[Article](#) [Google Scholar](#)

-
19. Meibom, K. L. *et al.* The *Vibrio cholerae* chitin utilization program. *Proc. Natl Acad. Sci. USA* **101**, 2524–2529 (2004).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
20. Drescher, K., Nadell, C., Stone, H., Wingreen, N. & Bassler, B. Solutions to the public goods dilemma in bacterial biofilms. *Curr. Biol.* **24**, 50–55 (2014).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
21. Cordero, O. X., Ventouras, L. A., DeLong, E. F. & Polz, M. F. Public good dynamics drive evolution of iron acquisition strategies in natural bacterioplankton populations. *Proc. Natl Acad. Sci. USA* **109**, 20059–20064 (2012).

[Show context](#)[PubMed](#) [Article](#) [Google Scholar](#)

-
22. Matz, C. *et al.* Biofilm formation and phenotypic variation enhance predation-driven persistence of *Vibrio cholerae*. *Proc. Natl Acad. Sci. USA* **102**, 16819–16824 (2005).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
23. Hibbing, M. E., Fuqua, C., Parsek, M. R. & Peterson, S. B. Bacterial competition: surviving and thriving in the microbial jungle. *Nat. Rev. Microbiol.* **8**, 15–25 (2010).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
24. Rendueles, O. & Ghigo, J. M. Mechanisms of competition in biofilm communities. *Microbiol. Spectr.* **3**, 3 (2015).

[Show context](#)[Google Scholar](#)

-
25. Riley, M. A. & Wertz, J. E. Bacteriocins: evolution, ecology, and application. *Annu. Rev. Microbiol.* **56**, 117–137 (2002).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
26. Hayes, C. S., Aoki, S. K. & Low, D. A. Bacterial contact-dependent delivery systems. *Annu. Rev. Genet.* **44**, 71–90 (2010).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
27. Ho, B. T., Dong, T. G. & Mekalanos, J. J. A view to a kill: the bacterial type VI secretion system. *Cell Host Microbe.* **15**, 9–21 (2014).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
28. Russell, A. B., Peterson, S. B. & Mougous, J. D. Type VI secretion system effectors: poisons with a purpose. *Nat. Rev. Microbiol.* **12**, 137–148 (2014).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
29. Basler, M., Ho, B. T. & Mekalanos, J. J. Tit-for-tat: type VI secretion system counterattack during bacterial cell-cell interactions. *Cell* **152**, 884–894 (2013). **A report showing that the T6SS of *P.***

***aeruginosa* is deployed in response to the T6SS-mediated attack from other species in the vicinity.**

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

30. Nadell, C. D. & Bassler, B. L. A fitness trade-off between local competition and dispersal in *Vibrio cholerae* biofilms. *Proc. Natl Acad. Sci. USA* **108**, 14181–14185 (2011).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

31. Schluter, J., Nadell, C. D., Bassler, B. L. & Foster, K. R. Adhesion as a weapon in microbial competition. *ISME J.* **9**, 139–149 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

32. Kim, W., Racimo, F., Schluter, J., Levy, S. B. & Foster, K. R. Importance of positioning for microbial evolution. *Proc. Natl Acad. Sci. USA* **111**, E1639–E1647 (2014).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

33. Rumbaugh, K. P. *et al.* Quorum sensing and the social evolution of bacterial virulence. *Curr. Biol.* **19**, 341–345 (2009). **A study demonstrating that phenotypes that are regulated by quorum sensing can be exploited by cheating mutants within a population of *P. aeruginosa* during infection of a mouse model system.**

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

34. Inglis, R. F., Gardner, A., Cornelis, P. & Buckling, A. Spite and virulence in the bacterium *Pseudomonas aeruginosa*. *Proc. Natl Acad. Sci. USA* **106**, 5703–5707 (2009).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

-
35. Brown, S. P., Inglis, R. F. & Taddei, F. Evolutionary ecology of microbial wars: within-host competition and (incidental) virulence. *Evol. Appl.* **2**, 32–39 (2009).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

36. Levin, S. A. Complex adaptive systems: exploring the known, the unknown, and the unknowable. *Bull. Am. Math. Soc.* **40**, 3–19 (2003).

[Show context](#)

[Article](#) [Google Scholar](#)

37. Persat, A. *et al.* The mechanical world of bacteria. *Cell* **161**, 988–997 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

38. Stacy, A., McNally, L., Darch, S., Brown, S. P. & Whiteley, M. The biogeography of polymicrobial infection. *Nat. Rev. Microbiol.* **14**, 93–105 (2015). **A major recent review of processes that generate the spatial structure of different bacterial strains and species in microbial communities associated with infection.**

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

39. Driscoll, W. W. & Pepper, J. W. Theory for the evolution of diffusible external goods. *Evolution* **64**, 2682–2687 (2010).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

40. Lion, S. & van Baalen, M. Self-structuring in spatial evolutionary ecology. *Ecol. Lett.* **11**, 277–295 (2008).

[Show context](#)[PubMed](#) [Article](#) [Google Scholar](#)

-
41. O'Toole, G. A. & Wong, G. C. Sensational biofilms: surface sensing in bacteria. *Curr. Opin. Microbiol.* **30**, 139–146 (2016).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
42. Millet, Y. A. *et al.* Insights into *Vibrio cholerae* intestinal colonization from monitoring fluorescently labeled bacteria. *PLoS Pathog.* **10**, e1004405 (2014).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
43. Anderson, M. S., Garcia, E. C. & Cotter, P. A. Kind discrimination and competitive exclusion mediated by contact-dependent growth inhibition systems shape biofilm community structure. *PLoS Pathog.* **10**, e1004076 (2014).

[Show context](#)[PubMed](#) [Article](#) [Google Scholar](#)

-
44. Nadell, C. D., Foster, K. R. & Xavier, J. B. Emergence of spatial structure in cell groups and the evolution of cooperation. *PLoS Comput. Biol.* **6**, e1000716 (2010).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
45. Thomas, C. D. & Kunin, W. E. The spatial structure of populations. *J. Animal Ecol.* **68**, 647–657 (1999).

[Show context](#)[Article](#) [Google Scholar](#)

-
46. Hallatschek, O., Hersen, P., Ramanathan, S. & Nelson, D. R. Genetic drift at expanding frontiers promotes gene segregation. *Proc. Natl Acad. Sci. USA* **104**, 19926–19930 (2007). **A theoretical and**

experimental paper that outlines how spatial structure emerges along the leading edge of expanding bacterial colonies owing to genetic drift, which generates clonal patches of one genotype.

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

-
47. Weber, M. F., Poxleitner, G., Hebisch, E., Frey, E. & Opitz, M. Chemical warfare and survival strategies in bacterial range expansions. *J. R. Soc. Interface* **11**, 20140172 (2014).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

-
48. van Gestel, J., Weissing, F. J., Kuipers, O. P. & Kovacs, A. T. Density of founder cells affects spatial pattern formation and cooperation in *Bacillus subtilis* biofilms. *ISME J.* **8**, 2069–2079 (2014).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
49. Mitri, S., Clarke, E. & Foster, K. R. Resource limitation drives spatial organization in microbial groups. *ISME J.* **10**, 1471–1482 (2016).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
50. Van Dyken, J. D., Muller, M. J. I., Mack, K. M. L. & Desai, M. M. Spatial population expansion promotes the evolution of cooperation in an experimental prisoner's dilemma. *Curr. Biol.* **23**, 919–923 (2013).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
51. Müller, M., Neugeboren, B. I., Nelson, D. R. & Murray, A. W. Genetic drift opposes mutualism during spatial population expansion. *Proc. Natl Acad. Sci. USA* **111**, 1037–1042 (2014). **Research demonstrating that genetic drift in expanding *S. cerevisiae* colonies generates a spatial structure that inhibits cooperation between two genotypes in a synthetic system. By contrast, strong**

mutualism is shown to counteract the lineage-segregating influence of radial population growth.[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
52. Buttery, N. *et al.* Structured growth and genetic drift raise relatedness in the social amoeba *Dictyostelium discoideum*. *Biol. Lett.* **8**, 794–797 (2012).

[Show context](#)[PubMed](#) [Article](#) [Google Scholar](#)

-
53. Poilane, I., Karjalainen, T., Barc, M.-C., Bourlioux, P. & Collignon, A. Protease activity of *Clostridium difficile* strains. *Can. J. Microbiol.* **44**, 157–161 (1998).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
54. Hungate, R. The anaerobic mesophilic cellulolytic bacteria. *Bacteriol. Rev.* **14**, 1 (1950).

[Show context](#)[CAS](#) [PubMed](#) [Google Scholar](#)

-
55. Gilbert, H. J. & Hazlewood, G. P. Bacterial cellulases and xylanases. *Microbiology* **139**, 187–194 (1993).

[Show context](#)[CAS](#) [Google Scholar](#)

-
56. Ross-Gillespie, A., Gardner, A., West, S. A. & Griffin, A. S. Frequency dependence and cooperation: theory and a test with bacteria. *Am. Nat.* **170**, 331–342 (2007).

[Show context](#)[PubMed](#) [Article](#) [Google Scholar](#)

-
57. Kümmerli, R., Schiessl, K. T., Waldvogel, T., McNeill, K. & Ackermann, M. Habitat structure and

the evolution of diffusible siderophores in bacteria. *Ecol. Lett.* **17**, 1536–1544 (2014).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

58. West, S. A., Griffin, A. S., Gardner, A. & Diggle, S. P. Social evolution theory for microorganisms. *Nat. Rev. Microbiol.* **4**, 597–607 (2006).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

59. Köhler, T., Buckling, A. & van Delden, C. Cooperation and virulence of clinical *Pseudomonas aeruginosa* populations. *Proc. Natl Acad. Sci. USA* **106**, 6339–6344 (2009).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

60. Andersen, S. B. *et al.* Long-term social dynamics drive loss of function in pathogenic bacteria. *Proc. Natl Acad. Sci. USA* **112**, 10756–10761 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

61. Allen, B., Gore, J. & Nowak, M. A. Spatial dilemmas of diffusible public goods. *eLife* **2**, e01169 (2013).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

62. Borenstein, D. B., Meir, Y., Shaevitz, J. W. & Wingreen, N. S. Non-local interaction via diffusible resource prevents coexistence of cooperators and cheaters in a lattice model. *PLoS ONE* **8**, e63304 (2013).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

63. Damore, J. A. & Gore, J. Understanding microbial cooperation. *J. Theor. Biol.* **299**, 31–41 (2012).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

64. Dobay, A., Bagheri, H. C., Messina, A., Kümmerli, R. & Rankin, D. J. Interaction effects of cell diffusion, cell density and public goods properties on the evolution of cooperation in digital microbes. *J. Evol. Biol.* **27**, 1869–1877 (2014).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

65. Popat, R. *et al.* Quorum-sensing and cheating in bacterial biofilms. *Proc. Biol. Sci.* **279**, 4765–4771 (2012).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

66. Hamilton, W. D. The genetical evolution of social behaviour I. *J. Theor. Biol.* **7**, 1–16 (1964).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

67. Hamilton, W. D. The genetical evolution of social behaviour II. *J. Theor. Biol.* **7**, 17–52 (1964).
Landmark papers in evolutionary biology, establishing the fundamental theory and broad-ranging importance of genetic identity between individuals for the evolution of cooperation.

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

68. Mitri, S., Xavier, J. B. & Foster, K. R. Social evolution in multispecies biofilms. *Proc. Natl Acad. Sci. USA* **108**, 10839–10846 (2011).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

69. Kümmerli, R., Griffin, A. S., West, S. A., Buckling, A. & Harrison, F. Viscous medium promotes cooperation in the pathogenic bacterium *Pseudomonas aeruginosa*. *Proc. Biol. Sci.* **276**, 3531–3538 (2009).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

70. Julou, T. *et al.* Cell–cell contacts confine public goods diffusion inside *Pseudomonas aeruginosa* clonal microcolonies. *Proc. Natl Acad. Sci. USA* **110**, 12577–12582 (2013).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

71. Seminara, A. *et al.* Osmotic spreading of *Bacillus subtilis* biofilms driven by an extracellular matrix. *Proc. Natl Acad. Sci. USA* **109**, 1116–1121 (2012).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

72. Datta, M. S., Korolev, K. S., Cvijovic, I., Dudley, C. & Gore, J. Range expansion promotes cooperation in an experimental microbial metapopulation. *Proc. Natl Acad. Sci. USA* **110**, 7354–7359 (2013). **This paper and reference 50 provide evidence that genetic drift in expanding metapopulations of *S. cerevisiae* generates a spatial structure which favours the use of a cooperative enzyme by a single genotype.**

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

73. Korolev, K. S., Xavier, J. B., Nelson, D. R. & Foster, K. R. A. Quantitative test of population genetics using spatiogenetic patterns in bacterial colonies. *Am. Nat.* **178**, 538–552 (2011).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

74. Hol, F. J. H. *et al.* Spatial structure facilitates cooperation in a social dilemma: empirical evidence from a bacterial community. *PLoS ONE* **8**, e77042 (2013).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
75. Mitri, S. & Foster, K. R. The genotypic view of social interactions in microbial communities. *Annu. Rev. Genet.* **47**, 247–273 (2013).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
76. Foster, K. R. & Bell, T. Competition, not cooperation, dominates interactions among culturable microbial species. *Curr. Biol.* **22**, 1845–1850 (2012).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
77. Oliveria, N. M. *et al.* Biofilm formation as a response to ecological competition. *PLoS Biol.* **13**, e1002191 (2015).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
78. Pfeiffer, T. Cooperation and competition in the evolution of ATP-producing pathways. *Science* **292**, 504–507 (2001).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
79. Xavier, J. B. & Foster, K. R. Cooperation and conflict in microbial biofilms. *Proc. Natl Acad. Sci. USA* **104**, 876–881 (2007).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
80. Durrett, R. & Levin, S. Allelopathy in spatially distributed populations. *J. Theor. Biol.* **185**, 165–171 (1997).

[Show context](#)[PubMed](#) [Article](#) [Google Scholar](#)

-
81. Ratcliff, W. & Denison, R. Alternative actions for antibiotics. *Science* **332**, 547–548 (2011).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
82. Abrudan, M. I. *et al.* Socially mediated induction and suppression of antibiosis during bacterial coexistence. *Proc. Natl Acad. Sci. USA* **112**, 11054–11059 (2015).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
83. Borgeaud, S., Metzger, L. C., Scignari, T. & Blokesch, M. The type VI secretion system of *Vibrio cholerae* fosters horizontal gene transfer. *Science* **347**, 63–67 (2015).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
84. Gardner, A. & West, S. A. Spite and the scale of competition. *J. Evol. Biol.* **17**, 1195–1203 (2004).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
85. Bucci, V., Nadell, C. D. & Xavier, J. B. The evolution of bacteriocin production in bacterial biofilms. *Am. Nat.* **178**, E162–E173 (2011).

[Show context](#)[PubMed](#) [Article](#) [Google Scholar](#)

-
86. Tait, K. & Sutherland, I. W. Antagonistic interactions amongst bacteriocin-producing enteric bacteria in dual species biofilms. *J. Appl. Microbiol.* **93**, 345–352 (2002).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
87. Borenstein, D. B., Ringel, P., Basler, M. & Wingreen, N. S. Established microbial colonies can survive type VI secretion assault. *PLoS Comput. Biol.* **11**, e1004520 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

88. Wexler, A. G. *et al.* Human symbionts inject and neutralize antibacterial toxins to persist in the gut. *Proc. Natl Acad. Sci. USA* **113**, 3639–3644 (2016).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

89. Alteri, C. J. *et al.* Multicellular bacteria deploy the type VI secretion system to preemptively strike neighboring cells. *PLoS Pathog.* **9**, e1003608 (2013).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

90. Karlsson, F. H., Nookaew, I., Petranovic, D. & Nielsen, J. Prospects for systems biology and modeling of the gut microbiome. *Trends Biotechnol.* **29**, 251–258 (2011).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

91. Morris, J. J., Lenski, R. E. & Zinser, E. R. The black queen hypothesis: evolution of dependencies through adaptive gene loss. *mBio* **3**, e00036-12 (2012).

[Show context](#)

[Google Scholar](#)

92. Tripp, H. J. *et al.* SAR11 marine bacteria require exogenous reduced sulphur for growth. *Nature* **452**, 741–744 (2008).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
93. Oliveira, N. M., Niehus, R. & Foster, K. R. Evolutionary limits to cooperation in microbial communities. *Proc. Natl Acad. Sci. USA* **111**, 17941–17946 (2014).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

94. Estrela, S. & Brown, S. P. Metabolic and demographic feedbacks shape the emergent spatial structure and function of microbial communities. *PLoS Comput. Biol.* **9**, e1003398 (2013).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

95. Momeni, B., Waite, A. J. & Shou, W. Spatial self-organization favors heterotypic cooperation over cheating. *eLife* **2**, e00960 (2013). **A study in which synthetic obligate mutualist strains of *S. cerevisiae* are found to spatially exclude a cheating strain in surface-bound colonies in a manner that promotes cooperation between mutualists.**

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

96. Morris, B. E. L., Henneberger, R., Huber, H. & Moissl-Eichinger, C. Microbial syntrophy: interaction for the common good. *FEMS Microbiol. Rev.* **37**, 384–406 (2013).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

97. Callaghan, A. *et al.* The genome sequence of *Desulfatibacillum alkenivorans* AK-01: a blueprint for anaerobic alkane oxidation. *Environ. Microbiol.* **14**, 101–113 (2012).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

98. Schink, B. Synergistic interactions in the microbial world. *Antonie Van Leeuwenhoek* **81**, 257–261 (2002).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
99. Pande, S. *et al.* Fitness and stability of obligate cross-feeding interactions that emerge upon gene loss in bacteria. *ISME J.* **8**, 953–962 (2014).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
100. Rakoff-Nahoum, S., Foster, K. R. & Comstock, L. The evolution of cooperation within the gut microbiota. *Nature* **533**, 255–259 (2016).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
101. Estrela, S., Trisos, C. H. & Brown, S. P. From metabolism to ecology: cross-feeding interactions shape the balance between polymicrobial conflict and mutualism. *Am. Nat.* **180**, 566–576 (2012).

[Show context](#)[PubMed](#) [Article](#) [Google Scholar](#)

-
102. Momeni, B., Briley, K. A., Fields, M. W. & Shou, W. Strong inter-population cooperation leads to partner intermixing in microbial communities. *eLife* **2**, e00230 (2013).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
103. Kümmerli, R., Jiricny, N., Clarke, L. S., West, S. A. & Griffin, A. S. Phenotypic plasticity of a cooperative behaviour in bacteria. *J. Evol. Biol.* **22**, 589–598 (2009).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
104. Kümmerli, R. & Brown, S. P. Molecular and regulatory properties of a public good shape the evolution of cooperation. *Proc. Natl Acad. Sci. USA* **107**, 18921–18926 (2010).

[Show context](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
105. Brown, S. P. & Taddei, F. The durability of public goods changes the dynamics and nature of social dilemmas. *PLoS ONE* **2**, e593 (2007).

[Show context](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
106. Mellbye, B. & Schuster, M. Physiological framework for the regulation of quorum sensing-dependent public goods in *Pseudomonas aeruginosa*. *J. Bacteriol.* **196**, 1155–1164 (2014).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
107. Cornforth, D. M. & Foster, K. R. Competition sensing: the social side of bacterial stress responses. *Nat. Rev. Microbiol.* **11**, 285–293 (2013).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
108. Schuster, M., Sexton, D. J., Diggle, S. P. & Greenberg, E. P. Acyl-homoserine lactone quorum sensing: from evolution to application. *Annu. Rev. Microbiol.* **67**, 43–63 (2013).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
109. Ng, W.-L. & Bassler, B. L. Bacterial quorum-sensing network architectures. *Annu. Rev. Genet.* **43**, 197–222 (2009).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
110. Redfield, R. J. Is quorum sensing a side effect of diffusion sensing? *Trends Microbiol.* **10**, 365–370 (2002).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
111. Cornforth, D. M. *et al.* Combinatorial quorum sensing allows bacteria to resolve their social and physical environment. *Proc. Natl Acad. Sci. USA* **111**, 4280–4284 (2014).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
112. Kim, M. K., Ingremeau, F., Zhao, A., Bassler, B. L. & Stone, H. A. Local and global consequences of flow on bacterial quorum sensing. *Nat. Microbiol.* **1**, 15005 (2016).

[Show context](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
113. Nadell, C. D., Xavier, J. B., Levin, S. A. & Foster, K. R. The evolution of quorum sensing in bacterial biofilms. *PLoS Biol.* **6**, e14 (2008).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
114. Schluter, J., Schoech, A., Foster, K. R. & Mitri, S. The evolution of quorum sensing as a mechanism to infer kinship. *PLoS Comput. Biol.* **12**, e1004848 (2016).

[Show context](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
115. van der Ploeg, J. R. Regulation of bacteriocin production in *Streptococcus mutans* by the quorum-sensing system required for development of genetic competence. *J. Bacteriol.* **187**, 3980–3989 (2005).

[Show context](#)[CAS](#)[PubMed](#)[Article](#)[Google Scholar](#)

-
116. Fontaine, L. *et al.* Quorum-sensing regulation of the production of B1p bacteriocins in *Streptococcus thermophilus*. *J. Bacteriol.* **189**, 7195–7205 (2007).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
117. Risøen, P. A., Brurberg, M. B., Eijsink, V. G. & Nes, I. F. Functional analysis of promoters involved in quorum sensing-based regulation of bacteriocin production in *Lactobacillus*. *Mol. Microbiol.* **37**, 619–628 (2000).

[Show context](#)[PubMed](#) [Google Scholar](#)

-
118. LeRoux, M., Peterson, S. B. & Mougous, J. D. Bacterial danger sensing. *J. Mol. Biol.* **427**, 3744–3753 (2015).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
119. Korgaonkar, A. K. & Whiteley, M. *Pseudomonas aeruginosa* enhances production of an antimicrobial in response to *N*-acetylglucosamine and peptidoglycan. *J. Bacteriol.* **193**, 909–917 (2011).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
120. Dong, T. G. *et al.* Generation of reactive oxygen species by lethal attacks from competing microbes. *Proc. Natl Acad. Sci. USA* **112**, 2181–2186 (2015).

[Show context](#)[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
121. LeRoux, M. *et al.* Kin cell lysis is a danger signal that activates antibacterial pathways of *Pseudomonas aeruginosa*. *eLife* **4**, e05701 (2015). **An investigation showing that cell lysate upregulates the T6SS of *P. aeruginosa* such that cells attack when they detect cues of clonemate death in the near surroundings.**

[Show context](#)[Article](#) [Google Scholar](#)

-
122. Nakamaru, M., Matsuda, H. & Iwasa, Y. The evolution of cooperation in a lattice-structured population. *J. Theor. Biol.* **184**, 65–81 (1997).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

123. Durrett, R. & Levin, S. The importance of being discrete (and spatial). *Theor. Popul. Biol.* **46**, 363–394 (1994).

[Show context](#)

[Article](#) [Google Scholar](#)

124. Mitteldorf, J. & Wilson, D. S. Population viscosity and the evolution of altruism. *J. Theor. Biol.* **204**, 481–496 (2000).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

125. Ratzke, C. & Gore, J. Self-organized patchiness facilitates survival in cooperatively growing *Bacillus subtilis* populations. *Nat. Microbiol.* **1**, 16022 (2016).

[Show context](#)

[Article](#) [Google Scholar](#)

126. Hallatschek, O. & Nelson, D. R. Gene surfing in expanding populations. *Theor. Popul. Biol.* **73**, 158–170 (2008).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

127. Kerr, B., Riley, M. A., Feldman, M. W. & Bohannan, B. J. M. Local dispersal promotes biodiversity in a real-life game of rock-paper-scissors. *Nature* **418**, 171–174 (2002).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
128. Pande, S. *et al.* Privatization of cooperative benefits stabilizes mutualistic cross-feeding interactions in spatially structured environments. *ISME J.* **10**, 1413–1423 (2016).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

129. Tolker-Nielsen, T. & Molin, S. Spatial organization of microbial biofilm communities. *Microb. Ecol.* **40**, 75–84 (2000).

[Show context](#)

[PubMed](#) [Google Scholar](#)

130. Rendueles, O. *et al.* Rapid and widespread *de novo* evolution of kin discrimination. *Proc. Natl Acad. Sci. USA* **112**, 9076–9081 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

131. Strassmann, J. E., Gilbert, O. M. & Queller, D. C. Kin discrimination and cooperation in microbes. *Annu. Rev. Microbiol.* **65**, 349–367 (2011).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

132. Oldewurtel, E. R., Kouzel, N., Dewenter, L., Henseler, K. & Maier, B. Differential interaction forces govern bacterial sorting in early biofilms. *eLife* **4**, e10811 (2015).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

133. Smukalla, S. *et al.* *FLO1* is a variable green beard gene that drives biofilm-like cooperation in budding yeast. *Cell* **135**, 726–737 (2008).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

134. Dawkins, R. *The Selfish Gene* (Oxford Univ. Press, 1989).

[Show context](#)

[Google Scholar](#)

135. Maynard Smith, J. & Szathmary, E. *The Major Transitions in Evolution* (Oxford Univ. Press, 1995).

[Show context](#)

[Google Scholar](#)

136. Tarnita, C. E., Taubes, C. H. & Nowak, M. A. Evolutionary construction by staying together and coming together. *J. Theor. Biol.* **320**, 10–22 (2013).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

137. Claessen, D., Rozen, D. E., Kuipers, O. P., Søgaard-Andersen, L. & van Wezel, G. P. Bacterial solutions to multicellularity: a tale of biofilms, filaments and fruiting bodies. *Nat. Rev. Microbiol.* **12**, 115–124 (2014).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

138. Ratcliff, W. C., Denison, R. F., Borrello, M. & Travisano, M. Experimental evolution of multicellularity. *Proc. Natl Acad. Sci. USA* **109**, 1595–1600 (2012).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

139. Koschwanez, J. H., Foster, K. R. & Murray, A. Improved use of a public good selects for the evolution of undifferentiated multicellularity. *eLife* **2**, e00367 (2013).

[Show context](#)

[PubMed](#) [Article](#) [Google Scholar](#)

140. Koschwanez, J. H., Foster, K. R. & Murray, A. W. Sucrose utilization in budding yeast as a model for the origin of undifferentiated multicellularity. *PLoS Biol.* **9**, e1001122 (2011).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

141. Justice, S. S., Hunstad, D. A., Cegelski, L. & Hultgren, S. J. Morphological plasticity as a bacterial survival strategy. *Nat. Rev. Microbiol.* **6**, 162–168 (2008).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

142. Persat, A., Stone, H. A. & Gitai, Z. The curved shape of *Caulobacter crescentus* enhances surface colonization in flow. *Nat. Commun.* **5**, 3824 (2014).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

143. Drescher, K. *et al.* Architectural transitions in *Vibrio cholerae* biofilms at single-cell resolution. *Proc. Natl Acad. Sci. USA* **113**, E2066–E2072 (2016).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

144. Teschler, J. K. *et al.* Living in the matrix: assembly and control of *Vibrio cholerae* biofilms. *Nat. Rev. Microbiol.* **13**, 255–268 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

145. Berk, V. *et al.* Molecular architecture and assembly principles of *Vibrio cholerae* biofilms. *Science* **337**, 236–239 (2012).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

146. Nadell, C. D., Drescher, K., Wingreen, N. S. & Bassler, B. L. Extracellular matrix structure governs invasion resistance in bacterial biofilms. *ISME J.* **9**, 1700–1709 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

147. Smith, D. R. *et al.* *In situ* proteolysis of the *Vibrio cholerae* matrix protein RbmA promotes biofilm recruitment. *Proc. Natl Acad. Sci. USA* **112**, 10491–10496 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

148. Costerton, J. W., Lewandowski, Z., Caldwell, D. E., Korber, D. R. & Lappin-scott, H. M. Microbial biofilms. *Annu. Rev. Microbiol.* **49**, 711–745 (1995).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

149. Roberts, A. E., Kragh, K. N., Bjarnsholt, T. & Diggle, S. P. The limitations of *in vitro* experimentation in understanding biofilms and chronic infection. *J. Mol. Biol.* **427**, 3646–3661 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

150. Rusconi, R., Garren, M. & Stocker, R. Microfluidics expanding the frontiers of microbial ecology. *Annu. Rev. Biophys.* **43**, 65–91 (2014).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

151. Earle, K. A. *et al.* Quantitative imaging of gut microbiota spatial organization. *Cell Host Microbe* **18**, 478–488 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
152. Harrison, F., Muruli, A., Higgins, S. & Diggle, S. P. Development of an *ex vivo* porcine lung model for studying growth, virulence, and signaling of *Pseudomonas aeruginosa*. *Infect. Immun.* **82**, 3312–3323 (2014).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

153. Welch, J. L. M., Rossetti, B. J., Rieken, C. W., Dewhirst, F. E. & Borisy, G. G. Biogeography of a human oral microbiome at the micron scale. *Proc. Natl Acad. Sci. USA* **113**, E791–E800 (2016).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

154. Coyte, K. Z., Schluter, J. & Foster, K. R. The ecology of the microbiome: network, competition, and stability. *Science* **350**, 663–666 (2015).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

155. Hamilton, W. D. Altruism and related phenomena, mainly in social insects. *Annu. Rev. Ecol. Evol. Syst.* **3**, 192–232 (1972).

[Show context](#)

[Google Scholar](#)

156. Foster, K. R. & Wenseleers, T. A general model for the evolution of mutualisms. *J. Evol. Biol.* **19**, 1283–1293 (2006).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

157. Kreft, J. U., Picioreanu, C., Wimpenny, J. W. T. & van Loosdrecht, M. C. M. Individual-based modelling of biofilms. *Microbiology* **147**, 2897–2912 (2001).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

-
158. Xavier, J. B., Picioreanu, C. & van Loosdrecht, M. C. M. A framework for multidimensional modelling of activity and structure of multispecies biofilms. *Environ. Microbiol.* **7**, 1085–1103 (2005).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

159. Kreft, J. U. Biofilms promote altruism. *Microbiology* **150**, 2751–2760 (2004). **A landmark individual-based modelling study demonstrating how the spatial structure of cell lineages can promote the evolution of cooperation in biofilms.**

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

160. Schluter, J. & Foster, K. R. The evolution of mutualism in gut microbiota via host epithelial selection. *PLoS Biol.* **10**, e1001424 (2012).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

161. Zhao, K. *et al.* Psl trails guide exploration and microcolony formation in *Pseudomonas aeruginosa* biofilms. *Nature* **497**, 388–391 (2013).

[Show context](#)

[CAS](#) [PubMed](#) [Article](#) [Google Scholar](#)

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Glossary

Microbiota

The community of microorganisms that live in association with a particular host organism (for example, the gut microbiota) or abiotic environment (for example, the soil microbiota).

Social phenotypes

Phenotypes that exert an effect (either positive or negative) on the reproductive output of other individuals and which evolved, in part, because of this fitness effect that they exert.

Type VI secretion system

(T6SS). A mechanism for killing neighbouring cells by the extension of a phage-tail-derived structure to putatively puncture adjacent cells and deliver toxic effectors.

Dispersal

The process by which cells depart from a community, either individually or collectively. Dispersal can be active, in response to stresses such as nutrient limitation, or passive, owing to biofilm erosion by fluid flow.

Genetic drift

A change in allele frequency in a population due to random sampling of organisms across generations (for example, due to stochasticity in reproductive success).

Public goods

Substances that are secreted into the extracellular space that provide a benefit to other cells in the vicinity.

Cheating mutants

Genotypes that gain a relative fitness advantage by receiving the benefits of an evolved cooperative trait of other genotypes, such as a public good, without contributing to the cooperative interaction themselves.

Ecological productivity

The total biomass produced by a strain or species in a given environmental setting

Antibiotics

Molecules that are produced by various microorganisms and act as toxins against other microorganisms; some antibiotics have been co-opted as pharmaceuticals for the treatment of microbial infections.

Bacteriocins

Antibiotics that are produced by bacteria and specifically target other bacteria. Bacteriocins often occur as toxin–antitoxin pairs that are encoded on the same plasmid or in the same genomic neighbourhood.

Contact-dependent inhibition

A mechanism of inhibiting the growth of neighbouring cells by the extension of a helical structure to contact target cells and deliver toxic effector molecules.

Syntrophic relationships

Interactions in which one species benefits by using the product of another as a nutrient source; the producing species may in turn benefit from the removal of this product.

Flocculation

Aggregation of yeast cells to form large multicellular groups that precipitate from liquid cultures and exhibit heightened stress tolerance.

Greenbeard gene

A gene (or a set of closely linked genes) that is responsible for both an identifying phenotypic trait and a cooperative behaviour that targets that identifying trait, ensuring that the greenbeard gene bearer preferentially benefits other bearers of the greenbeard gene.

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